

1 **Genetic dissection of adaptive form and function in**  
2 **rapidly-speciating cichlid fishes**

3 Frederico Henning\*, Gonzalo Machado-Schiaffino\*, Lukas Baumgarten & Axel Meyer

4 Department of Biology, University of Konstanz, 78464 Konstanz, Germany

5 \* These authors contributed equally to this work

6 Corresponding author: axel.meyer@uni-konstanz.de

7 ***Abstract***

8 Genes of major phenotypic effects and strong genetic correlations can facilitate adaptation,  
9 direct selective responses and potentially lead to phenotypic convergence. However, the  
10 preponderance of this type of genetic architecture in repeatedly-evolved adaptations remains  
11 unknown. Using hybrids between *Haplochromis chilotes* (thick-lipped) and *H. nyererei* (thin-  
12 lipped) we investigated the genetics underlying hypertrophied lips and elongated heads, traits  
13 that evolved repeatedly in cichlids. At least 25 loci of small-to-moderate and mainly additive  
14 effects were detected. Phenotypic variation in lip and head morphology was largely  
15 independent. Although several QTL overlapped for lip and head morphology traits, they were  
16 often of opposite effects. The distribution of effect signs suggests strong selection on lips.  
17 The fitness implications of several detected loci were demonstrated using a laboratory assay  
18 testing for the association between genotype and variation in foraging performance. The  
19 persistence of low fitness alleles in head morphology appears to be maintained through  
20 antagonistic pleiotropy/close linkage with positive-effect lip morphology alleles. Rather than  
21 being based on few major loci with strong positive genetic correlations, our results indicate  
22 that the evolution of the Lake Victoria thick-lipped ecomorph is the result of selection on  
23 numerous loci distributed throughout the genome.

24 **Key words:** adaptation genetics, convergent evolution, QTL mapping, RAD-seq, foraging  
25 performance, Haplochromines

## 26 *Introduction*

27 The fit between organisms and their environments is one of the most striking outcomes of  
28 adaptive evolution. Fundamental aspects of the genetic basis of adaptation such as the  
29 number of loci and the extent of genetic independence between traits affect the direction of  
30 adaptive responses (Schluter 1996; Losos 2011) and the contribution of traits to speciation  
31 (Servedio et al. 2011; Flaxman et al. 2013; Feder et al. 2014). Their importance in shaping  
32 adaptive radiations remains widely debated (Orr 2005; Hendry 2013; Laland et al. 2014;  
33 Wray et al. 2014). Adaptation itself is difficult to demonstrate (Endler 1986) and most  
34 investigations focus on measurable proxies (e.g. morphology, coloration) rather than on  
35 primary targets of selection (Arnold 1983; Losos 2011).

36 The number of loci that typically underlie adaptations is a longstanding debate in  
37 adaptation genetics. Fisher's geometric model was widely successful in promoting the view  
38 that adaptations typically have highly polygenic genetic bases and led to a consensus, which  
39 was in line with Darwin's original emphasis on slow and gradual change (Orr 2005). As late  
40 as 1992 however, it was realized that there was actually scarce empirical support for this  
41 consensus (Orr and Coyne 1992). To stimulate the collection of relevant data Orr and Coyne  
42 articulated three criteria: a) The study must be sufficiently powered; b) The phenotypic  
43 differences must be of adaptive significance, and; c) The trait must differ between natural  
44 populations or species.

45 There has been a surge of publications using both laboratory crosses and population-  
46 based association/divergence mapping in a variety of systems, which have shown mixed  
47 support for the notions of major genes vs. minor genes as the typical genetic basis of

48 adaptation (Hall et al. 2006; Chan et al. 2010; van't Hof et al. 2011; Ellegren et al. 2012;  
49 Greenwood et al. 2012; Nadeau et al. 2012; Greenwood et al. 2013; Kowalko et al. 2013;  
50 Linnen et al. 2013; Weber et al. 2013; Arnegard et al. 2014; Miller et al. 2014; Poelstra et al.  
51 2014), including in cichlids (Albertson et al. 2003b; Streelman et al. 2003; Roberts et al.  
52 2009; O'Quin et al. 2012; O'Quin et al. 2013). As pointed out recently by Rockman (2012),  
53 the increased attention given to genes of large effect is likely to reflect ascertainment bias in  
54 favor of more tractable traits, as well as technical aspects of genetic mapping that favor the  
55 discovery of major genes. Nevertheless, the incorporation of the effects of the time of  
56 recruitment, distance from adaptive optima, drift and more recently gene flow (Orr 1998a,  
57 2005; Dittmar et al. 2016 and references therein) to Fisher's geometric model leads to  
58 predictions that there are circumstances in which major genes are expected to form the bulk  
59 of the genetic bases of adaptations. Some of these predictions have recently received  
60 empirical support from studies of locally-adapted *Mimulus* species (Ferris et al. 2016) and in  
61 experimental manipulations done in sticklebacks (Rogers et al. 2012).

62 While evolutionary convergence is frequently seen as an illustration of predictable  
63 solutions to similar pressures that are found by natural selection, it is also recognized that  
64 genetic correlations can direct or constrain evolutionary responses (Schluter 1996; Losos  
65 2011). The extraordinary convergence of trophic morphologies, such as those found in cichlid  
66 fishes has led to the question of whether there are biases in the generation of phenotypic  
67 variation that direct adaptive evolution towards certain trajectories (Brakefield 2006), an  
68 issue that is a matter of current debate (Laland et al. 2014; Wray et al. 2014). Some potential  
69 sources of bias in the origin of variation include small mutational and genetic target sizes of  
70 the convergent phenotype (*i.e.* the number of loci that underlie traits) (Gompel and  
71 Prud'homme 2009), as well as genetic covariation in the case of multi-trait phenotypes.  
72 Genetic correlations are the result of tight linkage or pleiotropy between loci underlying

73 different traits (Lande 1984). Positive genetic correlations result from concordant effect  
74 signs, *i.e.* when a substitution of one allele at such a locus leads to an increase in the adaptive  
75 value of both traits and have the potential of facilitating the evolution of multi-trait  
76 phenotypes. Negative correlations result from discordant effect signs in tightly linked, or  
77 pleiotropic loci (*i.e.* antagonistic pleiotropy) and might constrain adaptation. A mixture of  
78 concordant and discordant signs can neutralize the overall impact of shared loci (Gardner and  
79 Latta 2007). Convergent evolution could be seen as the product of biases in the origin of  
80 variation if for instance, the genetic architecture of repeatedly-evolved adaptations is  
81 dominated by a few loci that have large and concordant effects on multiple sub-traits.

82         A specialized morphology consisting of hypertrophied lips, narrow and pointed heads  
83 (Fig. 1) is a striking example of convergent evolution that is replicated across several cichlid  
84 radiations (Kocher et al. 1993; Oliver and Arnegard 2010; Colombo et al. 2013; Manousaki et  
85 al. 2013; Burrell 2014; Henning and Meyer 2014, Machado-Schiaffino et al. 2017). Thick-  
86 lipped ecomorphs are typically suction-feeders that forage for prey within rocky crevices  
87 (Video S1) (Keenleyside 1991). The repeated evolution of the thick-lipped ecomorph is  
88 thought to reflect parallel adaptation to several sources of selection associated with foraging  
89 in narrow rocky crevices, including accessing prey (Baumgarten et al. 2015, Machado-  
90 Schiaffino et al. 2017), generating sufficient suction power and detecting prey through  
91 sensorial specializations (Oliver and Arnegard 2010). Transcriptomic evidence has shown  
92 that hypertrophied lips in African and Neotropical cichlids share molecular-developmental  
93 mechanisms (Colombo et al. 2013). However, it is unknown whether these parallel patterns  
94 of gene expression are restricted to downstream effects or also include upstream genes  
95 because the genetic basis of this suite of traits has not been investigated.

96         *Haplochromis (Paralabidochromis) chilotes* and *Haplochromis (Pundamilia) nyererei*  
97 are rock-restricted species that are widely-distributed in Lake Victoria (Witte and Van Oijen

1990; Seehausen and Bouton 1998). *H. chilotes* is a specialized insectivore that is characterized by thick, lobed lips. *H. nyererei* is more abundant and its diet consists of mainly zooplankton and a smaller proportion of insects that are obtained by picking, snapping and to a lesser extent pull-scraping (Seehausen and Bouton 1998). Previous work showed that differences in the performance of both species can be assessed experimentally by measuring the success of obtaining prey from angled substrates (Baumgarten et al. 2015). Natural populations of both species vary considerably regarding certain traits (e.g. coloration) but the between-species differences in lip and head morphology (hereafter LM and HM) are consistent regardless of the populations sampled (Seehausen 1996). The maximum divergence time between Lake Victorian haplochromines is generally accepted to be 15-100 thousand years (Johnson et al. 1996; Keller et al. 2013; Brawand et al. 2014). Consistent with their recent divergence, *H. nyererei* and *H. chilotes* can be crossed in the laboratory and generate fertile F<sub>1</sub> hybrids (Stelkens et al. 2010).

Here, we genetically dissect the divergent trophic morphologies of *Haplochromis chilotes* and *H. nyererei* and ask whether the repeated evolution of thick-lipped cichlids is likely to have been facilitated by the presence of major genes and strong, positive genetic correlations (Laland et al. 2014; Seehausen et al. 2014; Wray et al. 2014). To validate the fitness effects of the variation in morphological traits and the detected QTL, we developed an assay that yields a continuous measurement of foraging performance and is suitable for genetic mapping. Specifically, we a) describe the positions and effects of loci influencing between-species variation in morphology and foraging performance; b) test if the variation in head and lip morphologies is genetically independent and c) test the adaptive significance of both morphological variation and the detected genetic loci.

## 121 *Materials and Methods*

### 122 **EXPERIMENTAL CROSSES**

123 The genetic mapping panel was obtained by hybridizing a *Haplochromis chilotes* male and a  
124 *H. nyererei* female. Laboratory stocks were established using specimens obtained from  
125 commercial breeders and have been maintained by full-sibling mating in the Animal  
126 Research Facility (University of Konstanz) for over ten years. Species selection was based on  
127 the specialized morphology of *H. chilotes* and previous reports that it can be hybridized with  
128 *H. nyererei* (Stelkens et al. 2010). Large between-species differences regarding morphology  
129 and foraging performance are preserved in captivity and there are no indications that trait  
130 values were affected by the breeding scheme that was used during stock maintenance  
131 (Baumgarten et al. 2015). Inter-specific F<sub>1</sub> hybrids were obtained by housing a male *H.*  
132 *chilotes* with three female *H. nyererei* in a 360l tank. After a few weeks, one mouthbrooding  
133 female was spotted, transferred to a 360l tank and kept there until the larvae were free-  
134 swimming ( $\approx$ 15 days). Four F<sub>1</sub> hybrids (one male and three females) reached reproductive  
135 maturity and were fully viable and fertile. The F<sub>2</sub> generation was obtained by intercrossing  
136 the three F<sub>1</sub> females with the F<sub>1</sub> male multiple times. A total of 22 broods were isolated with  
137 an average brood size of 18 F<sub>2</sub> fish that reached maturity. Larvae were raised in 1/6, 1/3 or  
138 1/2 compartments of a 360l tank to minimize the effects of density and decrease the variance  
139 of body size. These species are more commonly referred to as *Pundamilia nyererei* and  
140 *Paralabidochromis chilotes* in the recent literature. Here, we opted to follow the “generic  
141 classification of the haplochromines” (van Oijen 1996) to avoid confusion until a taxonomic  
142 revision is carried out.

### 143 **MOLECULAR METHODS**

144 Genomic DNA was extracted from pectoral fin samples from 291 F<sub>2</sub>s, four F<sub>1</sub>s and both  
145 parental individuals (*H. chilotes* male and *H. nyererei* female) using the Zymo genomic DNA  
146 extraction kit. Genomic DNA was treated with RNase A. The parentals were included in  
147 three different libraries to increase coverage and guarantee sufficient confidence in assigning  
148 homozygous genotypes. Double digest RADseq libraries were prepared following Peterson et  
149 al. (2012). Briefly, 1 µg of Genomic DNA per sample was double-digested using the  
150 restriction enzymes PstI and MspI (New England BioLabs) for 3 hours at 37 °C. P1 and P2  
151 adapters were ligated to the digested DNA using T4 ligase for 30 minutes at room  
152 temperature. A total of 300 individually barcoded samples were pooled in six libraries. Size  
153 selection for each library was performed using the Pippin Prep (Sage Science, Beverly, MA)  
154 with a selected size-range of 325 to 400bp. Genomic libraries were single-end sequenced  
155 (100 bp length) in six lanes on an Illumina HiSeq 2000.

## 156 **ddRAD MARKER SELECTION**

157 Raw sequence reads were trimmed to a length of 100 bp – the last base was trimmed based on  
158 the drop in FastQC scores - and demultiplexed using STACKS (Catchen et al. 2011). Only  
159 high sequencing quality reads, with correct barcodes and unambiguous RAD site were  
160 retained. Demultiplexed reads were aligned to the *H. nyererei* reference genome using GSnap  
161 (Wu and Watanabe 2005). We required unique alignments allowing for a maximum of two  
162 mismatches and no terminal alignments. The ref\_map.pl parameters in STACKS were set as  
163 default except for the following parameters: minimum depth coverage to report a stack (-m 5)  
164 and upper bound for epsilon (--bound\_high 0.05, to reduce the probability of false-  
165 homozygotes). The genotypes for each marker were exported using the F2- design in the  
166 genotypes program in Stacks, requiring that both parentals were homozygous for different  
167 alleles and that at least 150 F<sub>2</sub> individuals were genotyped per marker (-r 150) with a

168 minimum coverage of 20 reads per individual (-m 20) and allowing for automatic corrections  
169 (-c). The variable sites were uniformly distributed across the entire read lengths.

## 170 **LINKAGE MAP ESTIMATION**

171 A total of 1687 markers passed the quality filters and were used for linkage map construction  
172 using the maximum likelihood algorithm implemented in JoinMap v.4 software (Van Ooijen  
173 2006) following guidelines for quality control (Van Ooijen 2006; Broman and Sen 2009) and  
174 the same procedures and thresholds that were thoroughly described elsewhere (Henning et al.  
175 2014). Briefly, individuals were excluded from linkage map construction if they had > 30%  
176 missing genotypes (n = 38) and loci were excluded if they were under severe segregation  
177 distortion ( $P$ -value < 0.01, n = 143) or had >20% missing genotypes (n = 434). The grouping  
178 of markers was determined based on an independence LOD threshold of 5 and orders were  
179 optimized by a) comparing maps obtained using the maximum likelihood and regression  
180 algorithms implemented in JoinMap (Henning et al. 2014); b) visually inspecting graphical  
181 genotypes and c) analyzing improbable genotypes as given by JoinMap. The cross-link of all  
182 markers were inspected using the plot.rf function in R/qtl and the recombination frequency  
183 per individual and per library was inspected using the countXO function to detect error-prone  
184 individuals or sequencing library batch effects (Broman and Sen 2009). Finally, the  
185 congruence between the genetic map and the *H. nyererei* draft genome  
186 (P\_nyererei\_broad\_scaffolds\_v1) was analyzed.

187 The number of markers in RAD datasets normally exceeds the number of observed  
188 crossovers. Furthermore, unique placements in the absence of observed crossovers can  
189 sometimes be the result of missing data alone (Henning et al. 2014). All of this results in  
190 marker redundancy (markers that map to the exact same genetic location and cannot be  
191 distinguished based on observed crossovers) and incorrect orders and distances. Redundancy



192 was eliminated by combining all the markers that mapped to identical positions and/or could  
193 not be placed with confidence owing to missing data. These concatenated markers were  
194 named with the prefix “c” followed by two digits indicating the LG and two digits identifying  
195 the order within each LG. This approach allowed us to i) increase computational efficiency  
196 and eliminate the need of random marker elimination or “jiggling” in the QTL mapping  
197 software; ii) reduce the effects of stochastic placement dependent on missing data (Henning  
198 et al. 2014); and iii) reduce the amount of overall missing data, since the combined markers  
199 consisted in the sum of the total genotypic observations from the linked markers.

## 200 **MORPHOLOGICAL TRAIT MEASUREMENTS**

201 Standard photographs were taken from the lateral and dorsal view of 15 fish from each of the  
202 parental populations, the F<sub>1</sub> hybrids and 291 F<sub>2</sub>s at 12-15 months of age. Fish were  
203 anaesthetized with MS-222 (Sigma) and standardized photographs were taken from the dorsal  
204 and lateral views. Measurements from standardized photographs were performed using  
205 ImageJ software. A combination of linear and geometric morphometric measurements were  
206 employed to assess morphological variation associated with hypertrophied lips and head  
207 shape (Fig. S1).

208 Morphological traits values were obtained for between 284 (“Lip PC”) and 291  
209 (“LA”) F<sub>2</sub>s. The following linear measurements were considered: lip area (“LA”); upper lip  
210 area (“ULA”); lower lip area (“LLA”); lip length (“LL”); head length (“HL”) and head angle  
211 (“HA”) (Fig. S1). Geometric morphometrics was carried out to measure head shape (“HS”)  
212 and lip shape (“LS”). Eight landmarks and eight semi-landmarks were placed on the dorsal  
213 view of the fish. The landmarks were: (A1), (A2), (A8) and (A7) posterior and anterior  
214 extreme of the right and left orbit, (A3) and (A6) right and left starting point of the upper lip,  
215 (A4) tip of the upper lip, (A5) tip of the snout at the base of the upper lip. Semi-landmarks

216 were placed on the outlines of the snout and the upper lip (Fig. S1). HS and LS were  
217 measured by placing equally spaced semi-landmarks on each side of the dorsal view (A9-A16  
218 for LS, A17-A26 for HS), in relation to landmarks A2, A4, A5 and A7. Each landmark was  
219 digitized in tpsDig version 2.16 (Rohlf 2010a). Relative warps analysis was performed to  
220 remove all non-shape variation in tpsRelw version 1.49 (Rohlf 2010b). The positions of the  
221 semi-landmarks were moved along an estimated curve between the neighboring points to  
222 minimize squared distances between the adjusted positions and the corresponding points in  
223 the consensus. Statistical analysis of each shape was performed with classifier variables  
224 (species, sex) and with SL as a covariate in MorphoJ version 1.05f. Allometric effects were  
225 detracted from the shape using a pooled regression within each species with the Procrustes  
226 coordinates as dependent variables and SL as independent variable.

227         The relationships of all traits with standard length was tested using linear models in R.  
228 Those traits where the relationship with size was significant, were size-corrected by obtaining  
229 the residuals from a regression of each measurement on standard length. Of all the traits we  
230 investigated, only HS varied between sexes, which might reflect female adaptations for  
231 mouthbrooding. However, correction was deemed unnecessary because sex-corrected  
232 phenotypic values resulted in identical QTL mapping results, presumably because the  
233 relationship with sex does not interact statistically with species assignment. This makes  
234 biological sense since both species are mouthbrooders. Area measurements are the residuals  
235 of the regression on body area. Normality was tested using the Shapiro-Wilks test  
236 (shapiro.test function in R) and all traits were visually inspected using qqnorm and hist  
237 functions in R and the suggested quality control procedures of R/qtl (Broman and Sen 2009).  
238 All traits were normally distributed and were scaled to units of  $F_2$  phenotypic standard  
239 deviations. The signs of the phenotypic values of HA was reversed (multiplied by -1) for the

240 QTL mapping analysis so that higher trait values were always present in *H. chilotes* to reflect  
241 the predicted adaptive direction (see the “Signatures of Natural Selection” section below).

242 The traits were grouped in the following two categories (Fig. S1), Lip morphology  
243 (LM) and Head morphology (HM). LM comprises measurements of lip area (“LA”, “ULA”,  
244 “LLA”) and lip length (“LL”). HM includes measurements of head length (“HL”), head angle  
245 (“HA”) and head shape (“HS”). “LIP PC” consists of the first principal component from a  
246 PCA of a series of lip traits (Fig. S1). LIP PC was not included in any trait category because  
247 the measurement includes lip shape (LS) and is not independent from head shape thus  
248 rendering the analysis of genetic correlations uninterpretable. Measurement-correlated traits  
249 are independently affected by measurement error and/or capture different aspects of the  
250 phenotypic variation in complex traits. Because slightly different aspects of the traits are  
251 measured, the analysis of measurement-correlated traits allows the detection of a greater  
252 number of QTL that underlie biologically-relevant traits that are too complex to be  
253 represented by a single measurement. Discussing the extent of the shared genetic basis in  
254 these traits is biologically meaningless, but including them in the analysis is  
255 methodologically relevant because it creates an internal control of phenotyping, QTL  
256 mapping and QTL co-localization analysis (e.g. these traits should be correlated, share a  
257 significant amount of QTL and have similar QTL effect distributions).

## 258 **FORAGING PERFORMANCE**

259 We previously developed a laboratory assay to measure performance using a series of  
260 discrete angles (Baumgarten et al. 2015). In the present study, the acrylic glass structures  
261 were designed to yield a continuous measurement that is suitable for QTL mapping: the  
262 minimum angle that each fish (both parental and F<sub>2</sub>S) can forage on. The acrylic structures  
263 consist of an angle of 60° at the base that gradually reduces with height (15 cm), finishing

264 with an angle of 15° (Fig. 2C and Fig. S2). Small equally-sized pieces of mosquito larvae  
265 were placed at regular intervals of 3 mm along the inner vertical axis and were fixed by  
266 drying at 50 °C for 5 minutes. The experimental tanks (60x120x50 cm) were divided into four  
267 compartments using Plexiglas dividers. The experimental fish were starved for two days prior  
268 to the experiment and were transferred into the experimental compartments 24 hours before  
269 the beginning of each trial for acclimation. Non-experimental fish were maintained in the  
270 background compartment in order to maintain social interactions and improve the acclimation  
271 of the experimental fish.

272 Foraging performance was measured in ten individuals of each of the parental species  
273 (*H. chilotes* and *H. nyererei*) and 162 F<sub>2</sub> individuals. Phenotypic values are the residuals of  
274 the linear regression of the maximum foraging height on standard length. Due to the  
275 feasibility of collecting foraging data, the sample size is reduced compared to the  
276 morphological traits, which invariably leads to a less precise estimation of QTL effects.  
277 Nevertheless, care was taken to avoid bias and to ensure that the full spectrum of variation in  
278 HM and LM was represented in the foraging data. A video showing examples of the foraging  
279 trials is available in the supplementary files (Video S2).

## 280 **PHENOTYPIC CORRELATIONS IN F<sub>2</sub> HYBRIDS**

281 This analysis aimed at a) testing the contributions of the different morphological components  
282 to foraging performance, and; b) investigating the degree to which LM and HM segregate  
283 independently. The overall level of phenotypic correlation in the F<sub>2</sub> recombinant population  
284 was measured using Spearman's rho. The significance of the following correlations were  
285 tested: a) between morphological (LM, Lip PC and HM) and foraging traits and; b) between  
286 LM and HM. F<sub>2</sub> correlations emerge due to genetic correlations (when traits share QTL with  
287 concordant signs through linkage or pleiotropy) but also environmental correlations (*e.g.*

288 when one trait is the functional consequence of another trait or co-varies due to similar plastic  
289 responses).

## 290 **QUANTITATIVE TRAIT LOCI (QTL) MAPPING**

291 QTL mapping was performed for all traits using interval mapping (IM) and composite  
292 interval mapping (CIM) followed by a final evaluation using multiple interval mapping  
293 (MIM). In comparison to IM and CIM (or multiple QTL mapping – MQM), MIM has higher  
294 detection power, leads to more precise parameter estimates and allows for the simultaneous  
295 evaluation of interactions between detected QTL (Kao et al. 1999). However, QTL models  
296 based exclusively on MIM searches can be subject to over-parameterization as sample sizes  
297 decrease. To overcome these limitations, we took the following steps. The initial MIM model  
298 included all the QTL identified by IM and composite interval mapping, CIM using  
299 chromosome-wide LOD thresholds derived from 500 permutations and p-value cutoff of 0.05  
300 as inclusion criteria. IM was performed in R/qtl (Broman and Sen 2009) and WinQTL  
301 Cartographer v2.5 (available at <http://statgen.ncsu.edu/qtlcart/>). CIM and MIM analysis were  
302 conducted in WinQTL Cartographer v2.5. The positions of all QTL main effects included in  
303 the initial MIM models were optimized and the significance of each QTL main effect was  
304 tested. Non-significant QTL based on the BIC criteria and those with LOD scores below 2.5  
305 were excluded and model optimization proceeded as previously described (Silva et al. 2012).  
306 We aimed at the discovery of the maximum number of QTL and several observations suggest  
307 that the procedure we employed appropriately controlled for false-discovery: a) many of the  
308 suggestive QTL identified in the IM and CIM searches were eliminated from the initial model  
309 using the BIC model selection criteria; b) the amount of genetic variance explained does not  
310 suggest over-parametrization, and; c) the estimated QTL positions and effects are consistent  
311 across phenotypically correlated traits.

312 QTL were considered to co-localize when their 1-LOD intervals overlapped.  
313 Individual QTL effects, the total amount of phenotypic variance and the estimates of genetic  
314 variance (i.e. broad sense heritability) were obtained from the variance decomposition tables  
315 produced for the final MIM models in WinQTL Cartographer. Epistatic and dominant effects  
316 were grouped and analyzed as non-additive because the current implementation of MIM in  
317 WinQTL Cartographer only models epistatic interactions among QTL with significant main  
318 effects. Interactions that are unaccounted for in the QTL model will resemble dominant  
319 effects.

## 320 **SIGNATURES OF NATURAL SELECTION**

321 The distribution of effect signs was tested using the QTL sign test (QTLST) (Orr 1998b)  
322 using a custom R function written by Muir et al. (2014). The sign test was only applied to Lip  
323 PC and trait categories LM, HM because it only has power to reject the null hypothesis when  
324 the number of detected QTL  $> 6$ . No individual trait other than ULA had as many detected  
325 QTL with significant additive effects. Additive effects (in units of  $F_2$  standard deviation)  
326 were pooled in each trait category (Albertson et al. 2003a) and in the event of shared QTL,  
327 the effect with highest LOD support was selected for testing. Inclusion of the smallest co-  
328 localized effects led to congruent results. All tests were conducted in R version 3.1.1 (Team  
329 2014). In addition, the genome-wide additive effect estimates derived from interval mapping  
330 were used to compare the mean effects of adaptive and non-adaptive traits. The mean effect  
331 of LM, HM and FP was calculated from 10cM windows and was compared to the estimates  
332 for traits that also differ between the parentals but are likely non-adaptive (body depth, anal  
333 fin base and caudal peduncle length).

334 The designation of “positive” or “negative” allelic effects is based on the direction of  
335 adaptation for foraging in crevices (i.e., adaptive and maladaptive, respectively). “Positive”

336 effects are those that facilitate foraging in crevices. For most traits, the positive allelic effect  
337 increases the trait value, because hypertrophied lips and elongated heads are present in *H.*  
338 *chilotes*. The effects are reversed in the case of head angle (HA) because narrower and  
339 pointier heads facilitate foraging in crevices. To allow for the graphical comparison of the  
340 concordance of the effects of co-localizing QTL, the trait values of HA were reversed  
341 (multiplied by -1). Therefore, all alleles derived from *H. chilotes* (hereafter referred to as H  
342 alleles) are expected to increase trait values. Because we measured foraging performance in  
343 our mapping panel, the assignment of adaptive/maladaptive alleles was done directly by  
344 using the effect on foraging at the detected morphology QTL as a reference.

## 345 *Results*

### 346 **LINKAGE MAP CONSTRUCTION AND GENOME ANCHORING**

347 A saturated linkage map consisting of 1122 ddRAD markers distributed across 22 linkage  
348 groups with a total size of 1225.68cM was obtained, in agreement with the expectation based  
349 on the known haploid chromosome number in Haplochromine cichlids (Thompson 1981;  
350 Poletto et al. 2010) (Table S1). Eliminating redundancy led to the final linkage map used for  
351 QTL mapping that had 752 uniquely placed markers. The median interval size is 0.97cM,  
352 with 10 intervals larger than 10cM and a single interval (17cM) that is larger than 15cM  
353 (Table S2 and Fig. S3-S4). All but nine marker placements were congruent with the current  
354 *H. nyererei* draft genome sequence (Table S2). Two of these showed evidence of allelic  
355 dropout and were excluded from further analysis. Other incongruent markers showed no  
356 indications of genotyping errors and could be indicative of structural variations, genome  
357 fragmentation or mis-assembly. Comments highlighting the incongruences were added to  
358 Table S2.

359 The map of correspondence between our linkage map and the *H. nyererei* (Brawand et  
360 al. 2014) draft genome sequence shows a high level of congruency, which allowed for a high  
361 quality anchoring of the current scaffolds to our linkage map (Table S2). The comparison of  
362 genetic and physical distances did not point to the presence of large inversions segregating in  
363 our cross (*i.e.* no pairs of non-recombining markers that are separated by large physical  
364 distances were found). The physical distance between redundant markers and adjacent  
365 uniquely-placed markers ranged from 4bp to 1.86Mb (median = 83Kb) and 7Kb-6.36Mb  
366 (median = 424Kb), respectively. Therefore, marker redundancy is likely to have been caused  
367 by the close physical proximity of markers or small inversions in the parentals.

## 368 **PHENOTYPIC VARIATION AND CORRELATIONS IN F<sub>2</sub> HYBRIDS**

369 All traits, including foraging performance differed between the parental populations and  
370 segregated in the F<sub>2</sub> mapping panel (Fig. 2, Fig. S5). Trait values that facilitate foraging were  
371 present in *H. chilotes* (longer lips and narrower heads). Several LM measurements (LL, ULA  
372 and LA) and Lip PC were correlated with foraging performance with coefficients ranging  
373 between 0.19 and 0.27. HM traits showed no association with foraging performance, with the  
374 possible exception of HL, where a marginally non-significant relationship was found ( $P =$   
375 0.08,  $\rho = 0.13$ ). Variation in HM traits was also generally independent from variation in  
376 LM or LIPC traits ( $\rho = -0.08$  to 0.15), with the exception of two comparisons involving LA  
377 which were significant. All correlation coefficients are shown in Table 1.

## 378 **QTL MAPPING AND THE GENETIC ARCHITECTURE OF ADAPTATION**

379 The underlying genetic architecture was found to include many loci of small effect and a few  
380 of moderate effect (Table S3) that are distributed across all but three LGs (Fig. 3A). The  
381 number of detected QTL ranged from four to eleven. Some LGs have a high clumping of



382 QTL underlying all traits, indicating that these LGs have a moderate effect on most traits  
383 investigated. Specifically, LG11, LG13, LG20 and LG23 are associated with several traits  
384 across their entire length (Fig. 3A and Fig. S5). The majority of the detected foraging  
385 performance QTL co-localized with QTL underlying morphological traits, particularly LM  
386 (Fig. 3B) and in some cases, there was co-localization of QTL that influence all trait groups  
387 (Fig. 3C). The largest effect QTL account for 12.6%, 7.1% and 9.3% of the  $F_2$  phenotypic  
388 variance in LM, HM and foraging performance, respectively. The distribution of effect sizes  
389 we found suggests that our detection threshold is approximately 2% of  $F_2$  phenotypic  
390 variance (Fig. 4). In the final MIM QTL models, genetic effects on lip measurements are  
391 composed of mainly additive effects that explain on average 31% of the phenotypic variance  
392 (84% of the genetic variance). In contrast, additive effects accounted for only 22%  
393 phenotypic (64.95% genetic variance) in HM and 23% (67.24% of the genetic variance) in  
394 foraging performance traits (Table S4).

### 395 **GENOME-WIDE SIGNATURES OF NATURAL SELECTION**

396 H alleles were biased in their effect signs: The proportion of the positive additive effects was  
397 14/15 in LM, 10/13 in HM, 4/4 in foraging performance and 10/11 in the lip principal  
398 component trait (Table S3). The QTL sign test rejected the null hypothesis in LM indicating  
399 an excess of positive effect H alleles ( $P < 0.05$ ). In the case of the QTL with overlapping 1-  
400 LD intervals between HM and LM, all H alleles are adaptive for LM but nearly half were  
401 negative for HM (Fig. 4). In contrast, all of the H alleles at QTL unique to HM were adaptive.  
402 Furthermore, the inspection of genome-wide additive effect plots suggests that regions on  
403 different chromosomes or on the same chromosome but genetically distant from detected  
404 QTL also appear to have positive additive effect in the adaptive traits. As an example, the lip  
405 area trait (LA) mapped to at least five genomic regions (Fig. 5A). Not only are all of the

406 additive effects positive in the detected QTL, but also in LGs where no QTL were detected,  
407 such as LGs 11-16 (Fig. 5B).

## 408 *Discussion*

409 We investigated the genetic basis of foraging performance, lip and head morphology in a  
410 cross between two haplochromine cichlid species from one of the youngest and largest  
411 known cichlid radiations, *Haplochromis chilotes* and *H. nyererei* (Brawand et al. 2014). The  
412 first species combines a suite of morphological adaptations that are associated with foraging  
413 in rocky crevices for invertebrate larvae and that evolved in most cichlid adaptive radiations  
414 (Keenleyside 1991). Deciphering the contribution of traits to fitness can prove complicated  
415 even in model systems (Cook et al. 2012; Zeller et al. 2012)) and while it is clear that natural  
416 selection has shaped morphology in many textbook examples of adaptation (Albertson et al.  
417 2003a), the primary target of selection (e.g. foraging capacity) certainly involves diverse  
418 classes of traits (e.g. metabolic, behavioral) in addition to morphology. This multifaceted  
419 aspect of adaptation might be expected to involve more complex interactions between loci  
420 (Huang et al. 2012) as well as a higher number of them (Arnegard et al. 2014). Our results  
421 show that it is possible to measure adaptive significance directly (Arnold 1983) also at the  
422 genetic level.

423 We found strong evidence for a role of hypertrophied lips in foraging success and that  
424 numerous loci were recruited since the divergence between these two species. These findings  
425 constitute strong support for the adaptive significance of hypertrophic lips and highlight the  
426 genome-wide effects of the response to natural selection of polygenic traits in recent adaptive  
427 radiations (Flaxman et al. 2013; Feder et al. 2014). The evolution of this multi-trait  
428 phenotype does not appear to be dominated by positive genetic correlations or small genetic  
429 target sizes, which can bias phenotypic evolution. Rather, the genetics of these adaptive  
430 differences in trophic morphology is consistent with a model of mostly small effect loci,

431 where only a few loci explain more than 5% of phenotypic variation and an increasing  
432 number of loci smaller effects. Phenotypic correlations between trait groups were generally  
433 low. Despite the detection of several co-localizing QTL, the effects were not always  
434 concordant revealing potential genetic trade-offs in the evolution of hypertrophied lips and  
435 pointed heads.

436 It is predicted that major genes with pleiotropic function might be particularly  
437 important in local adaptation in the presence of gene flow (Seehausen et al. 2014; Dittmar et  
438 al. 2016; Ferris et al. 2016). We found evidence for the existence of positive genetic  
439 associations (through either pleiotropy or tight linkage) and some evidence for clustering on  
440 LG23. However, none of these factors seem to explain a large amount of between-species  
441 differences. The genetic architecture of these traits is more aptly described as uncorrelated,  
442 consisting of small-to-moderate additive effects across numerous loci. However, this does not  
443 rule out that though currently small, it is precisely the loci with positive correlation and larger  
444 effects that are important in the very early instances of divergence or under high levels of  
445 gene flow. This should be investigated with the comparison of the genome-wide pattern  
446 differentiation between *H. chilotes* and other sympatric haplochromines.

## 447 **THE GENETIC ARCHITECTURE OF ADAPTATION IN THICK-LIPPED** 448 **CICHLIDS**

449 The genetic basis of lip measurements is composed of mainly additive effects across  
450 numerous loci that are scattered throughout the genome. A large number of QTL of small  
451 effect that individually explained up to 12% of  $F_2$  phenotypic variation were detected on all  
452 but three LGs. LG11 and LG23 are associated with multiple traits in what appear to be  
453 multiple, closely-linked QTL. The genetic basis of the morphological traits we analyzed is  
454 consistent with that of other adaptive trophic morphologies analyzed in cichlids (*e.g.*

455 Albertson et al. 2003a; Parnell et al. 2012; Albertson et al. 2014) and also with what is  
456 thought to be the most common genetic architecture underlying quantitative phenotypes in  
457 general (Albert et al. 2008; Flint and Mackay 2009).

458         These conclusions are only strengthened by considering that our estimates of effect  
459 sizes and number of QTL are likely to be overestimates and underestimates, respectively. The  
460 actual genetic architecture underlying these traits is probably composed of many more  
461 undetected QTL with small effects (Flint and Mackay 2009). The development of next-  
462 generation sequencing technologies facilitated the use of forward-genetics on non-model  
463 organisms (Schneeberger 2014) and today a large number of studies meet Orr and Coyne's  
464 third criterion ("naturally occurring phenotypes"). However, considerable difficulties still  
465 exist for meeting Orr and Coyne's first criterion ("sufficient power") in QTL mapping using  
466 non-model organisms. The size of the F<sub>2</sub> panels are only a fraction of those used for genetic  
467 investigations in established models (Beavis 1994; Fishman et al. 2002; Laurie et al. 2004).  
468 The use of low sample sizes decreases the probability of detection of small effect QTL (*i.e.*  
469 increases the detection threshold), leads to biased estimates of effect sizes and insufficient  
470 power to disentangle the effects of closely-linked QTL (Beavis 1994; Xu 2003; Slate 2013).  
471 The size of the F<sub>2</sub> mapping panel determines the detection power threshold and the extent of  
472 the inflation of effect sizes introduced by factors such as the Beavis effect (Slate 2013).

473         The existence of multiple crossable cichlid species pairs with different divergence  
474 times that differ in these same traits offers a unique opportunity to test whether alleles of  
475 large additive effect are recruited in the earlier stages of adaptation as predicted by Fisher's  
476 geometric model of adaptive evolution (Orr 2005; Rockman 2012). This is supported, for  
477 example by work on sticklebacks (Rogers et al. 2012) and could be tested by further genetic  
478 mapping projects in the multiple thick-lipped ecomorphs that occur in other recent radiations  
479 such as the ones in Lake Malawi or the Midas cichlid radiation. Multiple ecologically-

480 divergent populations - from Lakes Nicaragua and Managua (*Amphilophus citrinellus* and *A.*  
481 *labiatus*), as well as the recently colonized crater lakes - are variable for lip morphology  
482 (Machado-Schiaffino et al. 2017). We have recently shown that phenotypic plasticity is an  
483 important component of between-morph variation (e.g. Machado-Schiaffino et al. 2014) and  
484 that genetic differences also exist between Neotropical morphs (Machado-Schiaffino et al.  
485 2017).

## 486 **THE CAUSES OF THE REPEATED EVOLUTION OF THICK-LIPPED** 487 **CICHLIDS**

488 If the evolution of cichlid thick-lipped ecomorphs were facilitated by biases in the origin of  
489 selectable variation, one would expect a large contribution from few loci to multiple traits. If  
490 covariance dominated the genetic basis of hypertrophic lips, then a) HM and LM would be  
491 expected to be largely positively correlated in  $F_2$ s; b) a significant portion of the covariation  
492 between HM and LM would be explained by co-localizing QTL and; c) the shared QTL  
493 would have concordant effects. In contrast, we found that LM and HM segregated largely  
494 independently, with the exception of two pairwise comparisons. Additional factors such as  
495 environmental variation or measurement error might have contributed to a failure to detect  
496 phenotypic associations between HM and the other classes of traits if the impact of these  
497 sources of errors be imagined to be largely independent in the different trait groups.  
498 However, the high degree of concordance between traits within the same trait groups suggests  
499 that measurement error did not have a major role in our analysis.

500         Seven (out of 13) QTL for HM co-localized with QTL for LM when considering an  
501 overlap of 1-LOD intervals but interestingly, three of them had negative effects for HM. The  
502 mixture of concordant and discordant effects at shared QTL can result in the masking of  
503 genetic correlations at the phenotypic level (Gardner and Latta 2007). Distinguishing between

504 close linkage and pleiotropy depends on the number of observed crossovers and is one of the  
505 main limitations of QTL mapping experiments. Nevertheless, the distinction between  
506 pleiotropy and linkage relates to *how little* recombination occurs between loci, with the  
507 former representing the extreme case of complete linkage. It is possible that close linkage has  
508 a similar effect to pleiotropy in rapid bursts of selection occurring in small populations  
509 (Gardner and Latta 2007).

510         Although the overall level of genetic covariance of LM and HM is unlikely to have a  
511 big effect in the response to selection, the presence of genetic trade-offs and antagonistic  
512 pleiotropy might still have an impact on trait evolution (Via and Hawthorne 2002).  
513 Overlapping QTL with concordant, positive effects were also found and it would be  
514 interesting to test whether these are among the first to be recruited in the initial adaptation or  
515 are important in adaptation through introgression. Likewise, lip area was weakly correlated  
516 with two measurements of head morphology and it would be interesting to test if this  
517 correlation is stronger in earlier instances of adaptation. These hypotheses can be tested for  
518 example by selection experiments in recombinant populations to analyze the fitness effects of  
519 individual QTL (e.g. Rogers et al. 2012; Arnegard et al. 2014).

520         The large genetic target size of the phenotypes that we investigated does not support  
521 the notion that similar phenotypes will be based on regions that are homologous to those that  
522 we have identified, particularly when compared to more divergent taxa (*i.e.* African vs.  
523 Neotropical cichlid radiations). However, because sharing of ancient genetic variation and  
524 incomplete lineage sorting is rampant in East African cichlids (Brawand et al. 2014) it could  
525 be true that the different African radiations have recruited ancient genetic variants. The  
526 accumulation of data linking genomic regions to evolutionarily relevant phenotypes in  
527 cichlids paves the way for exciting future research testing the importance of introgression and  
528 shared ancient genetic variation in cichlid adaptive radiations. It would be interesting to know

529 how often convergent phenotypic evolution between the haplochromine cichlid radiations in  
530 Lakes Victoria, Malawi and Tanganyika involves the recruitment of ancient shared variation,  
531 as was shown to be the case in the colonization of freshwater from marine environments in  
532 sticklebacks (Colosimo et al. 2005; Jones et al. 2012). Hybridization is a common  
533 phenomenon in many groups of organisms, particularly in recently diverged species and its  
534 role in adaptation to new environments has been debated for a long time (Lewontin and Birch  
535 1966). However, conclusive evidence of adaptive introgression is restricted to a few systems  
536 where the phylogenetic analysis of causal genetic regions in hybridizing species was  
537 performed, such as *Heliconius* (Pardo-Diaz et al. 2012). Both contemporary and ancient  
538 hybridization seem widespread in cichlid fish (e.g. Koblmuller et al. 2010; Joyce et al. 2011;  
539 Genner and Turner 2012; Keller et al. 2013) and it has been proposed to play a crucial role in  
540 cichlid adaptive radiations, the “hybrid swarm hypothesis” (Seehausen 2004). Testing for  
541 both the role of introgression and incomplete lineage sorting in adaptation can be achieved by  
542 *functional phylogenomics*, systematically contrasting the evolutionary histories of several  
543 genomic regions identified by forward-genetic screens with random genomic regions using  
544 target enrichment (outlined in Henning and Meyer 2014). Despite the decreasing costs for  
545 whole-genome sequencing, target enrichment is still more efficient for collecting high-  
546 coverage, population-level data from large contiguous genomic regions. It has been used for  
547 applications such as phylogenomics, exon sequencing or population-based fine-mapping  
548 (Burbano et al. 2010; Mamanova et al. 2010; Faircloth et al. 2012; Lemmon et al. 2012;  
549 Nadeau et al. 2012).

## 550 **SIGNATURES OF NATURAL SELECTION**

551 Morphological differences in LM, particularly in lip length were strongly associated with  
552 foraging success. Genetic variation at the loci underlying morphology could be demonstrated

553 to have an effect on foraging performance. Selection pressures in LM appear to be quite  
554 strong in natural conditions. This expectation was also confirmed by analyzing the  
555 distribution of effect signs. The null hypothesis of the distribution of QTL additive effect  
556 signs could be rejected for LM, thus supporting a role for directional natural selection in the  
557 evolution of these species differences. The QTL sign test we employed (QTL-ST) is  
558 conservative (Anderson and Slatkin 2003), tests for one particular scenario of natural  
559 selection (Orr 1998b) and is sensitive to variance in effect sizes (Rice and Townsend 2012).  
560 Therefore, the null hypothesis will only be rejected in extreme cases where the number of  
561 detected QTL is high and negative effects are virtually absent (e.g. Muir et al. 2014).  
562 Nevertheless, even with these restrictions it was possible to show that the observed  
563 abundance of positive effect alleles in LM traits is unlikely to have accumulated by chance.  
564 For comparison, although a large number of QTL responded to artificial selection for oil  
565 content in Maize in an even shorter timeframe, a great number of QTL (approx. 20%) had  
566 negative effects (Laurie et al. 2004). This suggests that the degree of selection for habitat  
567 partitioning in cichlid adaptive radiations is incredibly strong. In contrast, the persistence of  
568 negative effect alleles for HM, the overall distribution of additive effects and the weaker  
569 correlation with foraging performance all suggest a weaker or indirect selection pressure on  
570 HM. There are likely to be many additional QTL, given that the distribution of additive  
571 effects seems biased towards positive effects also in chromosomes where no QTL was  
572 detected. This suggests that many additional loci have diverged as the result of natural  
573 selection in an evolutionary timescale as short as 15 000 years.

574 Lip traits had the highest overall genome-wide effect with a median genome-wide  
575 additive effect = 0.1 (in units of  $F_2$  standard deviation) and a range of 0.07-0.135 for each  
576 trait. Despite the low number of detected QTL, foraging performance also had a high overall  
577 positive effect (0.087). The net effect of HM was also positive, albeit lower than the previous



578 traits (median = 0.045, range = 0.038-0.05). Although the individual estimates are not  
579 independent owing to linkage, the overall median additive effects allows for a straightforward  
580 comparison of the influence of natural selection on different traits. When analyzing random  
581 traits, it is not possible to polarize trait values in relation to foraging in crevices as we have  
582 done for lip and head morphology traits. Nevertheless, species differences that are not the  
583 direct products of natural selection should not be biased towards any particular sign and  
584 should yield an overall value close to zero. This was the case with the traits that were taken  
585 for comparison (body depth, anal fin base and caudal peduncle length - phenotypic data not  
586 shown) which had a median effect very close to zero (-0.008). Note that this statement  
587 concerns only between-species differences and does not imply that traits evolve randomly.

588         It was hypothesized based on simulations that selection acting on a large fraction of  
589 the genome can lead to a non-linear and rapid build-up of reproductive isolation during  
590 speciation with gene-flow, leading to the process of whole-genome congealing (Flaxman et  
591 al. 2013; Feder et al. 2014). This pattern of QTL with biased effect signs throughout the  
592 genome has also been described in oral jaw traits that are important in the cichlid adaptive  
593 radiations (Albertson et al. 2003a) and could support the model of genome-wide congealing  
594 (Flaxman et al. 2013; Brawand et al. 2014; Feder et al. 2014), since the divergence of  
595 haplochromines occurred recently and under at least, partial gene flow. The accumulation of  
596 anchored genomes and QTL data pave the way for high-resolution studies on natural  
597 populations that could provide insights on the degree of genomic divergence that is  
598 associated with selection on lip morphology (Seehausen et al. 2014).

## 599 **CONCLUSIONS**

600 In summary, our results suggest that *i*) the loci underlying the morphological adaptations we  
601 investigated are numerous and have small additive effects; *ii*) foraging performance is

602 functionally and genetically associated with between-species morphological differences,  
603 particularly in lip morphology; *iii*) the distribution of additive effects suggests that natural  
604 selection had a genome-wide effect; and that *iv*) variation in lip and head morphology is  
605 largely genetically independent. Genetic correlations between lip and head morphology are  
606 unlikely to facilitate concerted evolution and in fact might have constrained trait evolution  
607 through the tight coupling of discordant alleles or antagonistic pleiotropy. While recent  
608 empirical and theoretical work has highlighted the role of large effect variants and pleiotropy  
609 in the repeated evolution and the maintenance of adaptations (Ferris et al. 2016 and  
610 references therein), the present results show that this is certainly not a requirement for  
611 evolutionary convergence in adaptive radiations.

## 612 **ACKNOWLEDGEMENTS**

613 This work was supported by an Alexander von Humboldt fellowship to GMS, and grants  
614 from the DFG to GMS (MA6144/1-1) and to AM (ME 1725/18-1), from the CNPq/DAAD to  
615 FH (GDE-290049/2007-5) and from the European Research Council (ERC advanced grant  
616 ‘GenAdap’ 293700) to AM. We thank C. Chang-Rudolf for technical assistance and J.  
617 Torres-Dowdall, Karl Radtke, A. Kautt, Darrin Husley and C. Kratochwil for critical  
618 comments. Chris Muir provided the R code to run the QTL sign test. Ad Konings kindly  
619 permitted the use of images from the DVD “Malawi Cichlid Feeding Behavior” (Cichlid  
620 Press).

## 621 **LITERATURE CITED**

- 622 Albert, A. Y. K., S. Sawaya, T. H. Vines, A. K. Knecht, C. T. Miller, B. R. Summers, S.  
623 Balabhadra, D. M. Kingsley, and D. Schluter. 2008. The genetics of adaptive shape  
624 shift in stickleback: Pleiotropy and effect size. *Evolution* 62:76-85.  
625 Albertson, R., J. Strelman, and T. Kocher. 2003a. Directional selection has shaped the oral  
626 jaws of Lake Malawi cichlid fishes. *Proc. Natl. Acad. Sci. U. S. A.* 100:5252-5257.

- 627 Albertson, R. C., K. E. Powder, Y. A. Hu, K. P. Coyle, R. B. Roberts, and K. J. Parsons.  
628 2014. Genetic basis of continuous variation in the levels and modular inheritance of  
629 pigmentation in cichlid fishes. *Mol. Ecol.* 23:5135-5150.
- 630 Albertson, R. C., J. T. Streebman, and T. D. Kocher. 2003b. Genetic basis of adaptive shape  
631 differences in the cichlid head. *J. Hered.* 94:291-301.
- 632 Anderson, E. C. and M. Slatkin. 2003. Orr's quantitative trait loci sign test under conditions  
633 of trait ascertainment. *Genetics* 165:445-446.
- 634 Arnegard, M. E., M. D. McGee, B. Matthews, K. B. Marchinko, G. L. Conte, S. Kabir, N.  
635 Bedford, S. Bergek, Y. F. Chan, F. C. Jones, D. M. Kingsley, C. L. Peichel, and D.  
636 Schluter. 2014. Genetics of ecological divergence during speciation. *Nature* 511:307-  
637 311.
- 638 Arnold, S. J. 1983. Morphology, Performance and Fitness. *Am. Zool.* 23:347-361.
- 639 Baumgarten, L., G. Machado-Schiaffino, F. Henning, and A. Meyer. 2015. What big lips are  
640 good for: on the adaptive function of repeatedly evolved hypertrophied lips of cichlid  
641 fishes. *Biol. J. Linn. Soc.* 115:448-455.
- 642 Beavis, A. D. 1994. The power and deceit of QTL experiments: Lessons from comparative  
643 QTL studies. Pp. 250-266. 49th Annual Corn & Sorghum Research Conference.
- 644 Brakefield, P. M. 2006. Evo-devo and constraints on selection. *Trends Ecol. Evol.* 21:362-  
645 368.
- 646 Brawand, D., C. E. Wagner, Y. I. Li, M. Malinsky, I. Keller, S. Fan, O. Simakov, A. Y. Ng,  
647 Z. W. Lim, E. Bezaul, J. Turner-Maier, J. Johnson, R. Alcazar, H. J. Noh, P. Russell,  
648 B. Aken, J. Alföldi, C. Amemiya, N. Azzouzi, J.-F. Baroiller, F. Barloy-Hubler, A.  
649 Berlin, R. Bloomquist, K. L. Carleton, M. A. Conte, H. D'Cotta, O. Eshel, L. Gaffney,  
650 F. Galibert, H. F. Gante, S. Gnerre, L. Greuter, R. Guyon, N. S. Haddad, W. Haerty,  
651 R. M. Harris, H. A. Hofmann, T. Hourlier, G. Hulata, D. B. Jaffe, M. Lara, A. P. Lee,  
652 I. MacCallum, S. Mwaiko, M. Nikaido, H. Nishihara, C. Ozouf-Costaz, D. J. Penman,  
653 D. Przybylski, M. Rakotomanga, S. C. P. Renn, F. J. Ribeiro, M. Ron, W. Salzburger,  
654 L. Sanchez-Pulido, M. E. Santos, S. Searle, T. Sharpe, R. Swofford, F. J. Tan, L.  
655 Williams, S. Young, S. Yin, N. Okada, T. D. Kocher, E. A. Miska, E. S. Lander, B.  
656 Venkatesh, R. D. Fernald, A. Meyer, C. P. Ponting, J. T. Streebman, K. Lindblad-Toh,  
657 O. Seehausen, and F. Di Palma. 2014. The genomic substrate for adaptive radiation in  
658 African cichlid fish. *Nature* 513:375-381.
- 659 Broman, K. W. and S. Sen. 2009. A guide to QTL mapping with R/qtl. Springer, Dordrecht.
- 660 Burbano, H. A., E. Hodges, R. E. Green, A. W. Briggs, J. Krause, M. Meyer, J. M. Good, T.  
661 Maricic, P. L. F. Johnson, Z. Y. Xuan, M. Rooks, A. Bhattacharjee, L. Brizuela, F. W.  
662 Albert, M. de la Rasilla, J. Fortea, A. Rosas, M. Lachmann, G. J. Hannon, and S.  
663 Paabo. 2010. Targeted investigation of the Neandertal genome by array-based  
664 sequence capture. *Science* 328:723-725.
- 665 Burress, E. 2014. Cichlid fishes as models of ecological diversification: patterns,  
666 mechanisms, and consequences. *Hydrobiologia*:1-21.
- 667 Catchen, J. M., A. Amores, P. Hohenlohe, W. Cresko, and J. H. Postlethwait. 2011. Stacks:  
668 Building and genotyping loci de novo from short-read sequences. *G3-Genes Genomes*  
669 *Genetics* 1:171-182.
- 670 Chan, Y. F., M. E. Marks, F. C. Jones, G. Villarreal, M. D. Shapiro, S. D. Brady, A. M.  
671 Southwick, D. M. Absher, J. Grimwood, J. Schmutz, R. M. Myers, D. Petrov, B.  
672 Jonsson, D. Schluter, M. A. Bell, and D. M. Kingsley. 2010. Adaptive evolution of  
673 pelvic reduction in sticklebacks by recurrent deletion of a Pitx1 enhancer. *Science*  
674 327:302-305.

- 675 Colombo, M., E. T. Diepeveen, M. Muschick, M. E. Santos, A. Indermaur, N. Boileau, M.  
676 Barluenga, and W. Salzburger. 2013. The ecological and genetic basis of convergent  
677 thick-lipped phenotypes in cichlid fishes. *Mol. Ecol.* 22:670-684.
- 678 Colosimo, P. F., K. E. Hosemann, S. Balabhadra, G. Villarreal, M. Dickson, J. Grimwood, J.  
679 Schmutz, R. M. Myers, D. Schluter, and D. M. Kingsley. 2005. Widespread parallel  
680 evolution in sticklebacks by repeated fixation of ectodyplasin alleles. *Science*  
681 307:1928-1933.
- 682 Cook, L. M., B. S. Grant, I. J. Saccheri, and J. Mallet. 2012. Selective bird predation on the  
683 peppered moth: the last experiment of Michael Majerus. *Biol. Lett.*
- 684 Dittmar, E. L., C. G. Oakley, J. K. Conner, B. A. Gould, and D. W. Schemske. 2016. Factors  
685 influencing the effect size distribution of adaptive substitutions. *Proc. R. Soc. B* 283.
- 686 Ellegren, H., L. Smeds, R. Burri, P. I. Olason, N. Backstrom, T. Kawakami, A. Kunstner, H.  
687 Makinen, K. Nadachowska-Brzyska, A. Qvarnstrom, S. Uebbing, and J. B. W. Wolf.  
688 2012. The genomic landscape of species divergence in *Ficedula* flycatchers. *Nature*  
689 491:756-760.
- 690 Endler, J. A. 1986. *Natural selection in the wild*. Princeton University Press, Princeton, N.J.
- 691 Faircloth, B. C., J. E. McCormack, N. G. Crawford, M. G. Harvey, R. T. Brumfield, and T.  
692 C. Glenn. 2012. Ultraconserved elements anchor thousands of genetic markers  
693 spanning multiple evolutionary timescales. *Syst. Biol.* 61:717-726.
- 694 Feder, J. L., P. Nosil, A. C. Wacholder, S. P. Egan, S. H. Berlocher, and S. M. Flaxman.  
695 2014. Genome-wide congealing and rapid transitions across the speciation continuum  
696 during speciation with gene flow. *J. Hered.* 105:810-820.
- 697 Ferris, K. G., L. L. Barnett, B. K. Blackman, and J. H. Willis. 2016. The genetic architecture  
698 of local adaptation and reproductive isolation in sympatry within the *Mimulus*  
699 *guttatus* species complex. *Mol. Ecol.* Early View (doi. 10.1111/mec.13763)
- 700 Fishman, L., A. J. Kelly, and J. H. Willis. 2002. Minor quantitative trait loci underlie floral  
701 traits associated with mating system divergence in *Mimulus*. *Evolution* 56:2138-2155.
- 702 Flaxman, S. M., J. L. Feder, and P. Nosil. 2013. Genetic hitchhiking and the dynamic buildup  
703 of genomic divergence during speciation with gene flow. *Evolution* 67:2577-2591.
- 704 Flint, J. and T. F. C. Mackay. 2009. Genetic architecture of quantitative traits in mice, flies,  
705 and humans. *Genome Res.* 19:723-733.
- 706 Gardner, K. M. and R. G. Latta. 2007. Shared quantitative trait loci underlying the genetic  
707 correlation between continuous traits. *Mol. Ecol.* 16:4195-4209.
- 708 Genner, M. J. and G. F. Turner. 2012. Ancient hybridization and phenotypic novelty within  
709 Lake Malawi's cichlid fish radiation. *Mol. Biol. Evol.* 29:195-206.
- 710 Gompel, N. and B. Prud'homme. 2009. The causes of repeated genetic evolution. *Dev. Biol.*  
711 332:36-47.
- 712 Greenwood, A. K., J. N. Cech, and C. L. Peichel. 2012. Molecular and developmental  
713 contributions to divergent pigment patterns in marine and freshwater sticklebacks.  
714 *Evolution & Development* 14:351-362.
- 715 Greenwood, A. K., A. R. Wark, K. Yoshida, and C. L. Peichel. 2013. Genetic and neural  
716 modularity underlie the evolution of schooling behavior in Threespine Sticklebacks.  
717 *Curr. Biol.* 23:1884-1888.
- 718 Hall, M. C., C. J. Basten, and J. H. Willis. 2006. Pleiotropic quantitative trait loci contribute  
719 to population divergence in traits associated with life-history variation in *Mimulus*  
720 *guttatus*. *Genetics* 172:1829-1844.
- 721 Hendry, A. P. 2013. Key questions in the genetics and genomics of eco-evolutionary  
722 dynamics. *Heredity* 111:456-466.

- 723 Henning, F., H. J. Lee, P. Franchini, and A. Meyer. 2014. Genetic mapping of horizontal  
724 stripes in Lake Victoria cichlid fishes: benefits and pitfalls of using RAD markers for  
725 dense linkage mapping. *Mol. Ecol.* 23:5224-5240.
- 726 Henning, F. and A. Meyer. 2014. The evolutionary genomics of cichlid fishes: Explosive  
727 speciation and adaptation in the postgenomic era. *Annu. Rev. Genomics Hum. Genet.*  
728 15:417-441.
- 729 Huang, W., S. Richards, M. A. Carbone, D. H. Zhu, R. R. H. Anholt, J. F. Ayroles, L.  
730 Duncan, K. W. Jordan, F. Lawrence, M. M. Magwire, C. B. Warner, K. Blankenburg,  
731 Y. Han, M. Javaid, J. Jayaseelan, S. N. Jhangiani, D. Muzny, F. Ongeru, L. Perales, Y.  
732 Q. Wu, Y. Q. Zhang, X. Y. Zou, E. A. Stone, R. A. Gibbs, and T. F. C. Mackay. 2012.  
733 Epistasis dominates the genetic architecture of *Drosophila* quantitative traits. *Proc.*  
734 *Natl. Acad. Sci. U. S. A.* 109:15553-15559.
- 735 Johnson, T. C., C. A. Scholz, M. R. Talbot, K. Kelts, R. D. Ricketts, G. Ngobi, K. Beuning, I.  
736 Ssemmanda, and J. W. McGill. 1996. Late pleistocene desiccation of Lake Victoria  
737 and rapid evolution of cichlid fishes. *Science* 273:1091-1093.
- 738 Jones, F. C., M. G. Grabherr, Y. F. Chan, P. Russell, E. Mauceli, J. Johnson, R. Swofford, M.  
739 Pirun, M. C. Zody, S. White, E. Birney, S. Searle, J. Schmutz, J. Grimwood, M. C.  
740 Dickson, R. M. Myers, C. T. Miller, B. R. Summers, A. K. Knecht, S. D. Brady, H. L.  
741 Zhang, A. A. Pollen, T. Howes, C. Amemiya, E. S. Lander, F. Di Palma, K. Lindblad-  
742 Toh, D. M. Kingsley, B. I. G. S. Platf, and W. G. A. Team. 2012. The genomic basis  
743 of adaptive evolution in threespine sticklebacks. *Nature* 484:55-61.
- 744 Joyce, D. A., D. H. Lunt, M. J. Genner, G. F. Turner, R. Bills, and O. Seehausen. 2011.  
745 Repeated colonization and hybridization in Lake Malawi cichlids. *Curr. Biol.*  
746 21:R108-R109.
- 747 Kao, C. H., Z. B. Zeng, and R. D. Teasdale. 1999. Multiple interval mapping for quantitative  
748 trait loci. *Genetics* 152:1203-1216.
- 749 Keenleyside, M. H. A. 1991. *Cichlid fishes : behaviour, ecology and evolution.* Chapman &  
750 Hall, London.
- 751 Keller, I., C. E. Wagner, L. Greuter, S. Mwaiko, O. M. Selz, A. Sivasundar, S. Wittwer, and  
752 O. Seehausen. 2013. Population genomic signatures of divergent adaptation, gene  
753 flow and hybrid speciation in the rapid radiation of Lake Victoria cichlid fishes. *Mol.*  
754 *Ecol.* 22:2848-2863.
- 755 Koblmuller, S., B. Egger, C. Sturmbauer, and K. M. Sefc. 2010. Rapid radiation, ancient  
756 incomplete lineage sorting and ancient hybridization in the endemic Lake Tanganyika  
757 cichlid tribe Tropheini. *Mol. Phylogenet. Evol.* 55:318-334.
- 758 Kocher, T. D., J. A. Conroy, K. R. Mckaye, and J. R. Stauffer. 1993. Similar Morphologies of  
759 Cichlid Fish in Lakes Tanganyika and Malawi Are Due to Convergence. *Mol.*  
760 *Phylogenet. Evol.* 2:158-165.
- 761 Kowalko, J. E., N. Rohner, Santiago B. Rompani, Brant K. Peterson, Tess A. Linden, M.  
762 Yoshizawa, Emily H. Kay, J. Weber, Hopi E. Hoekstra, William R. Jeffery, R.  
763 Borowsky, and Clifford J. Tabin. 2013. Loss of schooling behavior in cavefish  
764 through sight-dependent and sight-independent mechanisms. *Curr. Biol.* 23:1874-  
765 1883.
- 766 Laland, K., T. Uller, M. Feldman, K. Sterelny, G. B. Muller, A. Moczek, E. Jablonka, and J.  
767 Odling-Smee. 2014. Does evolutionary theory need a rethink? - POINT Yes, urgently.  
768 *Nature* 514:161-164.
- 769 Lande, R. 1984. The genetic correlation between characters maintained by selection, linkage  
770 and inbreeding. *Genet. Res.* 44:309-320.
- 771 Laurie, C. C., S. D. Chasalow, J. R. LeDeaux, R. McCarroll, D. Bush, B. Hauge, C. Q. Lai,  
772 D. Clark, T. R. Rocheford, and J. W. Dudley. 2004. The genetic architecture of



- 773 response to long-term artificial selection for oil concentration in the maize kernel.  
774 Genetics 168:2141-2155.
- 775 Lemmon, A. R., S. A. Emme, and E. M. Lemmon. 2012. Anchored hybrid enrichment for  
776 massively high-throughput phylogenomics. Syst. Biol. 61:727-744.
- 777 Lewontin, R. C. and L. C. Birch. 1966. Hybridization as a source of variation for adaptation  
778 to new environments. Evolution 20:315-&.
- 779 Linnen, C. R., Y. P. Poh, B. K. Peterson, R. D. H. Barrett, J. G. Larson, J. D. Jensen, and H.  
780 E. Hoekstra. 2013. Adaptive evolution of multiple traits through multiple mutations at  
781 a single gene. Science 339:1312-1316.
- 782 Losos, J. B. 2011. Convergence, Adaptation, and Constraint. Evolution 65:1827-1840.
- 783 Machado-Schiaffino, G., F. Henning, and A. Meyer. 2014. Species-specific differences in  
784 adaptive phenotypic plasticity in an ecologically relevant trophic trait: Hypertrophic  
785 lips in Midas cichlid fishes. Evolution:286-291.
- 786 Machado-Schiaffino, G., A. Kautt, J. Torres-Dowdall, L. Baumgarten, F. Henning, F and A.  
787 Meyer. 2017. Incipient speciation driven by hypertrophied lips in Midas cichlid  
788 fishes. Molecular Ecology (in press).
- 789 Mamanova, L., A. J. Coffey, C. E. Scott, I. Kozarewa, E. H. Turner, A. Kumar, E. Howard, J.  
790 Shendure, and D. J. Turner. 2010. Target-enrichment strategies for next-generation  
791 sequencing. Nat. Meth. 7:111-118.
- 792 Manousaki, T., P. M. Hull, H. Kusche, G. Machado-Schiaffino, P. Franchini, C. Harrod, K.  
793 R. Elmer, and A. Meyer. 2013. Parsing parallel evolution: ecological divergence and  
794 differential gene expression in the adaptive radiations of thick-lipped Midas cichlid  
795 fishes from Nicaragua. Mol. Ecol. 22:650-669.
- 796 Miller, C. T., A. M. Glazer, B. R. Summers, B. K. Blackman, A. R. Norman, M. D. Shapiro,  
797 B. L. Cole, C. L. Peichel, D. Schluter, and D. M. Kingsley. 2014. Modular skeletal  
798 evolution in sticklebacks is controlled by additive and clustered quantitative trait loci.  
799 Genetics 197:405-420.
- 800 Muir, C. D., J. B. Pease, and L. C. Moyle. 2014. Quantitative genetic analysis indicates  
801 natural selection on leaf phenotypes across wild tomato species (*Solanum sect.*  
802 *lycopersicon*; Solanaceae). Genetics 198:1629-1643.
- 803 Nadeau, N. J., A. Whibley, R. T. Jones, J. W. Davey, K. K. Dasmahapatra, S. W. Baxter, M.  
804 A. Quail, M. Joron, R. H. Ffrench-Constant, M. L. Blaxter, J. Mallet, and C. D.  
805 Jiggins. 2012. Genomic islands of divergence in hybridizing *Heliconius* butterflies  
806 identified by large-scale targeted sequencing. Philosophical Transactions of the Royal  
807 Society B-Biological Sciences 367:343-353.
- 808 O'Quin, C. T., A. C. Drilea, M. A. Conte, and T. D. Kocher. 2013. Mapping of pigmentation  
809 QTL on an anchored genome assembly of the cichlid fish, *Metriaclima zebra*. BMC  
810 Genomics 14:287.
- 811 O'Quin, C. T., A. C. Drilea, R. B. Roberts, and T. D. Kocher. 2012. A small number of genes  
812 underlie male pigmentation traits in Lake Malawi cichlid fishes. Journal of  
813 Experimental Zoology Part B-Molecular and Developmental Evolution 318B:199-  
814 208.
- 815 Oliver, M. K. and M. E. Arnegard. 2010. A new genus for *Melanochromis labrosus*, a  
816 problematic Lake Malawi cichlid with hypertrophied lips (Teleostei: Cichlidae).  
817 Ichthyol. Explor. Fres. 21:209-232.
- 818 Orr, H. A. 1998a. The population genetics of adaptation: The distribution of factors fixed  
819 during adaptive evolution. Evolution 52:935-949.
- 820 Orr, H. A. 1998b. Testing natural selection vs. genetic drift in phenotypic evolution using  
821 quantitative trait locus data. Genetics 149:2099-2104.

- 822 Orr, H. A. 2005. The genetic theory of adaptation: A brief history. *Nat. Rev. Genet.* 6:119-  
823 127.
- 824 Orr, H. A. and J. A. Coyne. 1992. The genetics of adaptation - a reassessment. *Am. Nat.*  
825 140:725-742.
- 826 Pardo-Diaz, C., C. Salazar, S. W. Baxter, C. Merot, W. Figueiredo-Ready, M. Joron, W. O.  
827 McMillan, and C. D. Jiggins. 2012. Adaptive introgression across species boundaries  
828 in *Heliconius* butterflies. *PLoS Genetics* 8.
- 829 Parnell, N. F., C. D. Hulsey, and J. T. Strelman. 2012. The genetic basis of a complex  
830 functional system. *Evolution* 66:3352-3366.
- 831 Peterson, B. K., J. N. Weber, E. H. Kay, H. S. Fisher, and H. E. Hoekstra. 2012. Double  
832 digest RADseq: An inexpensive method for *de novo* SNP discovery and genotyping in  
833 model and non-model species. *Plos One* 7.
- 834 Poelstra, J. W., N. Vijay, C. M. Bossu, H. Lantz, B. Ryll, I. Muller, V. Baglione, P.  
835 Unneberg, M. Wikelski, M. G. Grabherr, and J. B. W. Wolf. 2014. The genomic  
836 landscape underlying phenotypic integrity in the face of gene flow in crows. *Science*  
837 344:1410-1414.
- 838 Rice, D. P. and J. P. Townsend. 2012. Resampling QTL effects in the QTL sign test leads to  
839 incongruous sensitivity to variance in effect size. *G3-Genes Genomes Genetics* 2:905-  
840 911.
- 841 Roberts, R. B., J. R. Ser, and T. D. Kocher. 2009. Sexual conflict resolved by invasion of a  
842 novel sex determiner in Lake Malawi cichlid fishes. *Science* 326:998-1001.
- 843 Rockman, M. V. 2012. The QTN program and the alleles that matter for evolution: All that's  
844 gold does not glitter. *Evolution* 66:1-17.
- 845 Rogers, S. M., P. Tamkee, B. Summers, S. Balabhadra, M. Marks, D. M. Kingsley, and D.  
846 Schluter. 2012. Genetic Signature of Adaptive Peak Shift in Threespine Stickleback.  
847 *Evolution* 66:2439-2450.
- 848 Rohlf, F. 2010a. tpsDig version 2.16, Department of Ecology and Evolution, State University  
849 of New York at Stony Brook. Available at <http://life.bio.sunysb.edu/morph/>.
- 850 Rohlf, F. 2010b. tpsRelw, version 1.49, Department of Ecology and Evolution, State  
851 University of New York at Stony Brook. Available at  
852 <http://life.bio.sunysb.edu/morph/>.
- 853 Schluter, D. 1996. Adaptive radiation along genetic lines of least resistance. *Evolution*  
854 50:1766-1774.
- 855 Schneeberger, K. 2014. Using next-generation sequencing to isolate mutant genes from  
856 forward genetic screens. *Nat. Rev. Genet.* advance online publication.
- 857 Seehausen, O. 1996. Distribution of and reproductive isolation among color morphs of a  
858 rock-dwelling Lake Victoria cichlid (*Haplochromis nyererei*). *Ecol. Freshw. Fish.*  
859 5:195-202.
- 860 Seehausen, O. 2004. Hybridization and adaptive radiation. *Trends Ecol. Evol.* 19:198-207.
- 861 Seehausen, O. and N. Bouton. 1998. The community of rock-dwelling cichlids in Lake  
862 Victoria. *Bonner Zoologische Beiträge* 47:301-312.
- 863 Seehausen, O., R. K. Butlin, I. Keller, C. E. Wagner, J. W. Boughman, P. A. Hohenlohe, C.  
864 L. Peichel, G.-P. Saetre, C. Bank, A. Brannstrom, A. Brelsford, C. S. Clarkson, F.  
865 Eroukmanoff, J. L. Feder, M. C. Fischer, A. D. Foote, P. Franchini, C. D. Jiggins, F.  
866 C. Jones, A. K. Lindholm, K. Lucek, M. E. Maan, D. A. Marques, S. H. Martin, B.  
867 Matthews, J. I. Meier, M. Most, M. W. Nachman, E. Nonaka, D. J. Rennison, J.  
868 Schwarzer, E. T. Watson, A. M. Westram, and A. Widmer. 2014. Genomics and the  
869 origin of species. *Nat. Rev. Genet.* 15:176-192.
- 870 Servedio, M. R., G. S. Doorn, M. Kopp, A. M. Frame, and P. Nosil. 2011. Magic traits in  
871 speciation: 'magic' but not rare? *Trends in Ecology and Evolution* 26:389-397.

- 872 Silva, L. D. E., S. Wang, and Z. B. Zeng. 2012. Composite interval mapping and multiple  
 873 interval mapping: Procedures and guidelines for using windows QTL Cartographer.  
 874 Pp. 75-120 in S. A. Rifkin, ed. Quantitative trait loci (QTL) : methods and protocols.  
 875 Humana Press, Springer, New York.
- 876 Slate, J. 2013. From Beavis to beak color: A simulation study to examine how much QTL  
 877 mapping can reveal about the genetic architecture of quantitative traits. *Evolution*  
 878 67:1251-1262.
- 879 Stelkens, R. B., K. A. Young, and O. Seehausen. 2010. The accumulation of reproductive  
 880 incompatibilities in African cichlid fish. *Evolution* 64:617-632.
- 881 Streebman, J. T., R. C. Albertson, and T. D. Kocher. 2003. Genome mapping of the orange  
 882 blotch colour pattern in cichlid fishes. *Mol. Ecol.* 12:2465-2471.
- 883 Team, R. C. 2014. R: A Language and Environment for Statistical Computing. R Foundation  
 884 for Statistical Computing, Vienna, Austria.
- 885 van't Hof, A. E., N. Edmonds, M. Dalikova, F. Marec, and I. J. Saccheri. 2011. Industrial  
 886 melanism in British peppered moths has a singular and recent mutational origin.  
 887 *Science* 332:958-960.
- 888 van Oijen, M. J. P. 1996. The generic classification of the haplochromine cichlids of Lake  
 889 Victoria, East Africa *Zool. Verh. Leiden* 302:57-110.
- 890 Van Ooijen, J. W. 2006. JoinMap 4. Software for the calculation of genetic linkage maps in  
 891 experimental populations. Kyazma BV, Wageningen, The Netherlands: .
- 892 Via, S. and D. J. Hawthorne. 2002. The genetic architecture of ecological specialization:  
 893 Correlated gene effects on host use and habitat choice in pea aphids. *Am. Nat.*  
 894 159:S76-S88.
- 895 Weber, J. N., B. K. Peterson, and H. E. Hoekstra. 2013. Discrete genetic modules are  
 896 responsible for complex burrow evolution in *Peromyscus mice*. *Nature* 493:402-405.
- 897 Witte, F. and M. J. P. Van Oijen. 1990. Taxonomy, ecology and fishery of Lake Victoria  
 898 haplochromine trophic groups. *Zoologische Verhandelingen* 262:1-47.
- 899 Wray, G. A., H. E. Hoekstra, D. J. Futuyma, R. E. Lenski, T. F. C. Mackay, D. Schluter, and  
 900 J. E. Strassmann. 2014. Does evolutionary theory need a rethink? -  
 901 COUNTERPOINT No, all is well. *Nature* 514:161-164.
- 902 Wu, T. D. and C. K. Watanabe. 2005. GMAP: a genomic mapping and alignment program  
 903 for mRNA and EST sequences. *Bioinformatics* 21:1859-1875.
- 904 Xu, S. Z. 2003. Theoretical basis of the Beavis effect. *Genetics* 165:2259-2268.
- 905 Zeller, M., K. Lucek, M. Haesler, O. Seehausen, and A. Sivasundar. 2012. Little evidence for  
 906 a selective advantage of armour-reduced threespined stickleback individuals in an  
 907 invertebrate predation experiment. *Evol. Ecol.* 26:1293-1309.

908



909 **TABLES**

910 Table 1. Phenotypic correlation matrix of F<sub>2</sub> hybrids of all measured traits. Lip area (“LA”),  
 911 upper lip area (“ULA”), lower lip area (“LLA”), lip length (“LL”), lip principal component  
 912 (“LIP PC”), head length (“HL”), head angle (“HA”), head shape (“HS”) and foraging  
 913 performance (“FP”). The values of Spearman’s rank correlation coefficient are given below  
 914 the diagonal. Correlations between different trait groups are highlighted in bold. \*, \*\* and  
 915 \*\*\* represent *P* values of < 0.05, <0.01 and <0.001, respectively.

	LA	ULA	LLA	LL	LIPC	HA	HL	HS
ULA	0.579***							
LLA	0.545***	0.224***						
LL	0.497***	0.437***	0.298***					
LIPC	0.809***	0.716***	0.682***	0.729***				
HA	<b>-0.001</b>	<b>0.073</b>	<b>-0.059</b>	<b>-0.04</b>	-0.014			
HL	<b>0.149*</b>	<b>0.055</b>	<b>0.066</b>	<b>0.108</b>	0.078	-0.302***		
HS	<b>0.129*</b>	<b>-0.016</b>	<b>0.076</b>	<b>-0.042</b>	0.023	-0.241***	0.238***	
FP	<b>0.241**</b>	<b>0.186*</b>	<b>0.107</b>	<b>0.252***</b>	<b>0.266***</b>	<b>-0.087</b>	<b>0.13</b>	<b>0.075</b>

916 **FIGURE LEGENDS**

917 **Figure 1.** Convergent evolution and function of hypertrophic lips. Representative species are  
 918 shown from the cichlid radiations of the African great lakes, Central America and South  
 919 America. Photographs were kindly provided by Erwin Schraml, Ad Konings and Oliver  
 920 Lucanus. In the image sequence on the bottom, an individual *Placidochromis milomo*  
 921 (representative of the lake Malawi radiation) is seen searching for prey (left), targeting a  
 922 rocky crevice (center) and accessing the prey (right).

923 **Figure 2.** Foraging performance and morphological traits are correlated and segregate in F<sub>2</sub>  
 924 hybrids. A) Male specimens of both species used in the experiment. B) Distribution of  
 925 phenotypic values in representative traits in the parental and F<sub>2</sub> populations. C) The acrylic

926 device (left) and the experimental setting (right) developed to measure foraging performance.  
 927 D) Differences between the parental and F<sub>2</sub> populations in foraging performance. E) The  
 928 correlation between foraging performance and lip length. Spearman's correlation coefficient  
 929 is shown.

930 **Figure 3.** QTL map of foraging performance and associated morphological traits. A)  
 931 Distribution of all detected main effect QTL for all trait groups: foraging performance ("FP",  
 932 green), lip morphology ("LM", blue), head morphology ("HM", red) and lip principal  
 933 component ("LIP PC", black). The map distance in cM is given by the scales on the left.  
 934 Thick and thin bars represent the 1- and 2- LOD intervals, respectively. B-C) Overlapping  
 935 LOD profiles of QTL for different trait groups (shown with arrowheads in A). To avoid  
 936 redundancy, only the most highly supported QTL from each of the different trait groups  
 937 ("LM", "HM", "LIP PC" and "FP") are shown. The overlap of the 1-LOD intervals is  
 938 represented by the grey boxes.

939 **Figure 4.** Direction and distribution of QTL effects. A-B) Concordant and antagonistic allelic  
 940 effects at co-localizing QTL for lip and head morphology (in units of F<sub>2</sub> standard deviation).  
 941 Alleles inherited from *Haplochromis chilotes* and from *H. nyererei* are represented by "H"  
 942 and "N", respectively. Slopes of opposite signs are indicative of antagonistic effects because  
 943 all traits were polarized with regards to foraging in crevices. H alleles are expected to  
 944 increase phenotypic values for all traits (see Methods). Intersecting effect slopes are more  
 945 apparent in the comparison between homozygous genotypes (CC and NN) since non-additive  
 946 genetic variation can result in CN genotypes having phenotypic values above or below the  
 947 expected under a purely additive model (e.g. HS at LG17 or HA at LG13). C) Distribution of  
 948 detected additive effects. Effect sizes are expressed in percentage of explained F<sub>2</sub> phenotypic  
 949 variance.

950 **Figure 5.** QTL map of lip area (“LA”). A) At least five genomic regions underlie phenotypic  
951 variation in LA. The LOD profiles for the three different detection methods (IM, CIM and  
952 MIM) are shown and are largely congruent. The dark and light horizontal lines represent the  
953 genome-wide (3.7) and chromosome-wide (2.5) significance thresholds for IM and CIM. All  
954 MIM QTL that are shown are significant using the BIC criteria. B) All detected QTL have a  
955 positive additive effect (in standard deviation units).

For Peer Review Only

# 1 **Genetic dissection of adaptive form and function in** 2 **rapidly-speciating cichlid fishes**

3 Frederico Henning\*, Gonzalo Machado-Schiaffino\*, Lukas Baumgarten & Axel Meyer

4 Department of Biology, University of Konstanz, 78464 Konstanz, Germany

5 \* These authors contributed equally to this work

6 Corresponding author: [axel.meyer@uni-konstanz.de](mailto:axel.meyer@uni-konstanz.de)

## 7 **Abstract**

8 Genes of major phenotypic effects and strong genetic correlations can facilitate  
9 ~~convergence~~adaptation, direct selective responses and ~~might be particularly important during~~  
10 ~~adaptive divergence in the presence of gene flow~~potentially lead to phenotypic convergence.

11 However, the preponderance of this type of genetic architecture in repeatedly-evolved  
12 adaptations remains unknown. Using hybrids between *Haplochromis chilotes* (thick-lipped)  
13 and *H. nyererei* (thin-lipped) we investigated the genetics underlying hypertrophied lips and  
14 elongated heads, traits that evolved repeatedly in cichlids. At least 25 loci of small-to-  
15 moderate and mainly additive effects were detected. Phenotypic variation in lip and head  
16 morphology was largely independent. Although several QTL overlapped for lip and head  
17 morphology traits, they were often of opposite effects. The distribution of effect signs  
18 suggests strong selection on lips. The fitness implications of several detected loci were  
19 demonstrated using a laboratory assay testing for the association between genotype and  
20 variation in foraging performance. The persistence of low fitness alleles in head morphology  
21 appears to be maintained through antagonistic pleiotropy/close linkage with positive-effect  
22 lip morphology alleles. Rather than being based on few major loci with strong positive  
23 genetic correlations, our results indicate that the evolution of the Lake Victoria thick-lipped  
24 ecomorph is the result of selection on numerous loci distributed throughout the genome.

25 | **Key words:** adaptation genetics, convergent evolution, QTL mapping, RAD-seq, [foraging](#)  
26 | [performance](#), [Haplochromines](#)

## 27 | *Introduction*

28 | The fit between organisms and their environments is one of the most striking outcomes of  
29 | adaptive evolution. Fundamental aspects of the genetic basis of adaptation such as the  
30 | number of loci and the extent of genetic independence between traits affect the direction of  
31 | adaptive responses (Schluter 1996; Losos 2011) and the contribution of traits to speciation  
32 | (Servedio et al. 2011; Flaxman et al. 2013; Feder et al. 2014). Their importance in shaping  
33 | adaptive radiations remains widely debated (Orr 2005; Hendry 2013; Laland et al. 2014;  
34 | Wray et al. 2014). Adaptation itself is difficult to demonstrate (Endler 1986) and most  
35 | investigations focus on measurable proxies (e.g. morphology, coloration) rather than on  
36 | primary targets of selection (Arnold 1983; Losos 2011).


37 |         The number of loci that typically underlie adaptations is a longstanding debate in  
38 | adaptation genetics. Fisher's geometric model was widely successful in promoting the view  
39 | that adaptations typically have highly polygenic genetic bases and led to a consensus, which  
40 | was in line with Darwin's original emphasis on slow and gradual change (Orr 2005). As late  
41 | as 1992 however, it was realized that there was actually scarce empirical support for this  
42 | consensus (Orr and Coyne 1992). To stimulate the collection of relevant data Orr and Coyne  
43 | articulated three criteria: a) The study must be sufficiently powered; b) The phenotypic  
44 | differences must be of adaptive significance, and; c) The trait must differ between natural  
45 | populations or species.

46 |         There has been a surge of publications using both laboratory crosses and population-  
47 | based association/divergence mapping in a variety [of](#) systems, which have shown mixed  
48 | support for the notions of major genes vs. minor genes as the typical genetic basis of

49 adaptation (Hall et al. 2006; Chan et al. 2010; van't Hof et al. 2011; Ellegren et al. 2012;  
50 Greenwood et al. 2012; Nadeau et al. 2012; Greenwood et al. 2013; Kowalko et al. 2013;  
51 Linnen et al. 2013; Weber et al. 2013; Arnegard et al. 2014; Miller et al. 2014; Poelstra et al.  
52 2014), including in cichlids (Albertson et al. 2003b; Streelman et al. 2003; Roberts et al.  
53 2009; O'Quin et al. 2012; O'Quin et al. 2013). As pointed out recently by Rockman (2012),  
54 the increased attention given to genes of large effect is likely to reflect ascertainment bias in  
55 favor of more tractable traits, as well as technical aspects of genetic mapping that favor the  
56 discovery of major genes. Nevertheless, the incorporation of the effects of the time of  
57 recruitment, distance from adaptive optima, drift and more recently gene flow (Orr 1998a,  
58 2005; Dittmar et al. 2016 and references therein) to Fisher's geometric model leads to  
59 predictions that there are circumstances in which major genes are expected to form the bulk  
60 of the genetic bases of adaptations. Some of these predictions have recently received  
61 empirical support from studies of locally-adapted *Mimulus* species (Ferris et al. 2016) and in  
62 experimental manipulations done in sticklebacks (Rogers et al. 2012).

63 While evolutionary convergence is frequently seen as an illustration of predictable  
64 solutions to similar pressures that are found by natural selection, it is also recognized that  
65 genetic correlations can direct or constrain evolutionary responses (Schluter 1996; Losos  
66 2011). The extraordinary convergence of trophic morphologies, such as those found in cichlid  
67 fishes has led to the question of whether there are biases in the generation of phenotypic  
68 variation that direct adaptive evolution towards certain trajectories (Brakefield 2006), an  
69 issue that is a matter of current debate (Laland et al. 2014; Wray et al. 2014). Some potential  
70 sources of bias in the origin of variation include small mutational and genetic target sizes of  
71 the convergent phenotype (*i.e.* the number of loci that underlie traits) (Gompel and  
72 Prud'homme 2009), as well as genetic covariation in the case of multi-trait phenotypes.  
73 Genetic correlations are the result of tight linkage or pleiotropy between loci underlying

74 different traits (Lande 1984). Positive genetic correlations result from concordant effect  
75 signs, *i.e.* when a substitution of one allele at such a locus leads to an increase in the adaptive  
76 value of both traits and have the potential of facilitating the evolution of multi-trait  
77 phenotypes. Negative correlations result from discordant effect signs in tightly linked, or  
78 pleiotropic loci (*i.e.* antagonistic pleiotropy) and might constrain adaptation. A mixture of  
79 concordant and discordant signs can neutralize the overall impact of shared loci (Gardner and  
80 Latta 2007). Convergent evolution could be seen as the product of biases in the origin of  
81 variation if for instance, the genetic architecture of repeatedly-evolved adaptations is  
82 dominated by a few loci that have large and concordant effects on multiple sub-traits.

83 

84 A specialized morphology consisting of hypertrophied lips, narrow and pointed heads (Fig. 1)  
85 is a striking example of convergent evolution that is replicated across several cichlid  
86 radiations (Kocher et al. 1993; Oliver and Arnegard 2010; Colombo et al. 2013; Manousaki et  
87 al. 2013; Burrell 2014; Henning and Meyer 2014, [Machado-Schiaffino et al. 2017](#)). Thick-  
88 lipped ecomorphs are typically suction-feeders that forage for prey within rocky crevices  
89 (Video S1) (Keenleyside 1991). The repeated evolution of the thick-lipped ecomorph is  
90 thought to reflect parallel adaptation to several sources of selection associated with foraging  
91 in narrow rocky crevices, including accessing prey (Baumgarten et al. 2015, [Machado-  
92 Schiaffino et al. 2017](#)), generating sufficient suction power and detecting prey through  
93 sensorial specializations (Oliver and Arnegard 2010). Transcriptomic evidence has shown  
94 that ~~the convergent evolution of~~ hypertrophied lips in African and Neotropical cichlids shares  
95 molecular-developmental mechanisms (Colombo et al. 2013). However, it is unknown  
96 whether these parallel patterns of gene expression are restricted to downstream effects or also  
97 include upstream genes because the genetic basis of this suite of traits has not been  
98 investigated.

99            *Haplochromis (Paralabidochromis) chilotes* and *Haplochromis (Pundamilia) nyererei*  
100 are rock-restricted species that are widely-distributed in Lake Victoria (Witte and Van Oijen  
101 1990; Seehausen and Bouton 1998). *H. chilotes* is a specialized insectivore that is  
102 characterized by thick, lobed lips. *H. nyererei* is more abundant and its diet consists of mainly  
103 zooplankton and a smaller proportion of insects that are obtained by picking, snapping and to  
104 a lesser extent pull-scraping (Seehausen and Bouton 1998). Previous work showed that  
105 differences in the performance of both species can be assessed experimentally by measuring  
106 the success of obtaining prey from angled substrates (Baumgarten et al. 2015). Natural  
107 populations of both species vary considerably regarding certain traits (e.g. coloration) but the  
108 between-species differences in lip and head morphology (hereafter LM and HM) are  
109 consistent regardless of the populations sampled (Seehausen 1996). The maximum  
110 divergence time between Lake Victorian haplochromines is generally accepted to be 15-100  
111 thousand years (Johnson et al. 1996; Keller et al. 2013; Brawand et al. 2014). Consistent with  
112 their recent divergence, *H. nyererei* and *H. chilotes* can be crossed in the laboratory and  
113 generate fertile F<sub>1</sub> hybrids (Stelkens et al. 2010).

114            Here, we genetically dissect the divergent trophic morphologies of *Haplochromis*  
115 *chilotes* and *H. nyererei* and ask whether the repeated evolution of thick-lipped cichlids is  
116 likely to have been facilitated by the presence of major genes and strong, positive genetic  
117 correlations (Laland et al. 2014; Seehausen et al. 2014; Wray et al. 2014). To validate the  
118 fitness effects of the variation in morphological traits and the detected QTL, we developed an  
119 assay that yields a continuous measurement of foraging performance and is suitable for  
120 genetic mapping. Specifically, we a) describe the positions and effects of loci influencing  
121 between-species variation in morphology and foraging performance; b) test if the variation in  
122 head and lip morphologies is genetically independent and c) test the adaptive significance of  
123 both morphological variation and the detected genetic loci.



## 124 *Materials and Methods*

### 125 **EXPERIMENTAL CROSSES**

126 The genetic mapping panel was obtained by hybridizing a *Haplochromis chilotes* male and a  
127 *H. nyererei* female. Laboratory stocks were established using specimens obtained from  
128 commercial breeders and have been maintained by full-sibling mating in the Animal  
129 Research Facility (University of Konstanz) for over ~~10~~ten years. Species selection was based  
130 on the specialized morphology of *H. chilotes* and previous reports that it can be hybridized  
131 with *H. nyererei* (Stelkens et al. 2010). Large between-species differences ~~in morphology~~  
132 regarding morphology and foraging performance are preserved in captivity and there are no  
133 indications that trait values were affected by the breeding scheme that was used during stock  
134 maintenance (Baumgarten et al. 2015). Inter-specific F<sub>1</sub> hybrids were obtained by housing a  
135 male *H. chilotes* with three female *H. nyererei* in a 360l tank. After a few weeks, one  
136 mouthbrooding female was spotted, transferred to a 360l tank and kept there until the larvae  
137 were free-swimming ( $\approx$ 15 days). Four F<sub>1</sub> hybrids (one male and three females) reached  
138 reproductive maturity and were fully viable and fertile. The F<sub>2</sub> generation was obtained by  
139 intercrossing the three F<sub>1</sub> females with the F<sub>1</sub> male multiple times. A total of 22 broods were  
140 isolated with an average brood size of 18 F<sub>2</sub> fish that reached maturity. -Larvae were raised in  
141 1/6, 1/3 or 1/2 compartments of a 360l tank to minimize the effects of density and decrease  
142 the variance of body size. These species are more commonly referred to as *Pundamilia*  
143 *nyererei* and *Paralabidochromis chilotes* in the recent literature. Here, we opted to follow the  
144 “generic classification of the haplochromines” (van Oijen 1996) to avoid confusion until a  
145 taxonomic revision is carried out.

### 146 **MOLECULAR METHODS**

147 Genomic DNA was extracted from pectoral fin samples from 291 F<sub>2</sub>s, four F<sub>1</sub>s and both  
148 parental individuals (*H. chilotes* male and *H. nyererei* female) using the Zymo genomic DNA  
149 extraction kit. Genomic DNA was treated with RNAse A. The parentals were included in  
150 three different libraries to increase coverage and guarantee sufficient confidence in assigning  
151 homozygous genotypes. Double digest RADseq libraries were prepared following Peterson et  
152 al. (2012). Briefly, 1 µg of Genomic DNA per sample was double-digested using the  
153 restriction enzymes PstI and MspI (New England BioLabs) for 3 hours at 37 °C. P1 and P2  
154 adapters were ligated to the digested DNA using T4 ligase for 30 minutes at room  
155 temperature. A total of 300 individually barcoded samples were pooled in six libraries. Size  
156 selection for each library was performed using the Pippin Prep (Sage Science, Beverly, MA)  
157 with a selected size-range of 325 to 400bp. Genomic libraries were single-end sequenced  
158 (100 bp length) in six lanes on an Illumina HiSeq 2000.

### 159 **ddRAD MARKER SELECTION**

160 Raw sequence reads were trimmed to a length of 100 bp – the last base was trimmed based on  
161 the drop in FastQC scores - and demultiplexed using STACKS (Catchen et al. 2011). Only  
162 high sequencing quality reads, with correct barcodes and unambiguous RAD site were  
163 retained. Demultiplexed reads were aligned to the *H. nyererei* reference genome using GSnap  
164 (Wu and Watanabe 2005). We required unique alignments allowing for a maximum of two  
165 mismatches and no terminal alignments. The ref\_map.pl parameters in STACKS were set as  
166 default except for the following parameters: minimum depth coverage to report a stack (-m 5)  
167 and upper bound for epsilon (--bound\_high 0.05, to reduce the probability of false-  
168 homozygotes). The genotypes for each marker were exported using the F2- design in the  
169 genotypes program in Stacks, requiring that both parentals were homozygous for different  
170 alleles and that at least 150 F<sub>2</sub> individuals were genotyped per marker (-r 150) with a

171 minimum coverage of 20 reads per individual (-m 20) and allowing for automatic corrections  
172 (-c). The variable sites were uniformly distributed across the entire read lengths.

### 173 **LINKAGE MAP ESTIMATION**

174 A total of 1687 markers passed the quality filters and were used for linkage map construction  
175 using the maximum likelihood algorithm implemented in JoinMap v.4 software (Van Ooijen  
176 2006) following guidelines for quality control (Van Ooijen 2006; Broman and Sen 2009) and  
177 the same procedures and thresholds that were thoroughly described elsewhere (Henning et al.  
178 2014). Briefly, individuals were excluded from linkage map construction if they had > 30%  
179 missing genotypes (n = 38) and loci were excluded if they were under severe segregation  
180 distortion ( $P$ -value < 0.01, n = 143) or had >20% missing genotypes (n = 434). The grouping  
181 of markers was determined based on an independence LOD threshold of 5 and orders were  
182 optimized by a) comparing maps obtained using the maximum likelihood and regression  
183 algorithms implemented in JoinMap (Henning et al. 2014); b) visually inspecting graphical  
184 genotypes and c) analyzing improbable genotypes as given by JoinMap. The cross-link of all  
185 markers were inspected using the plot.rf function in R/qtl and the recombination frequency  
186 per individual and per library was inspected using the countXO function to detect error-prone  
187 individuals or sequencing library batch effects (Broman and Sen 2009). Finally, the  
188 congruence between the genetic map and the *H. nyererei* draft genome  
189 (P\_nyererei\_broad\_scaffolds\_v1) was analyzed.

190 The number of markers in RAD datasets normally exceeds the number of observed  
191 crossovers. Furthermore, unique placements in the absence of observed crossovers can  
192 sometimes be the result of missing data alone (Henning et al. 2014). All of this results in  
193 marker redundancy (markers that map to the exact same genetic location and cannot be  
194 distinguished based on observed crossovers) and incorrect orders and distances. Redundancy

195 was eliminated by combining all the markers that mapped to identical positions and/or could  
196 not be placed with confidence owing to missing data. These concatenated markers were  
197 named with the prefix “c” followed by two digits indicating the LG and two digits identifying  
198 the order within each LG. This approach allowed us to i) increase computational efficiency  
199 and eliminate the need of random marker elimination or “jiggling” in the QTL mapping  
200 software; ii) reduce the effects of stochastic placement dependent on missing data (Henning  
201 et al. 2014); and iii) reduce the amount of overall missing data, since the combined markers  
202 consisted in the sum of the total genotypic observations from the linked markers.

### 203 **MORPHOLOGICAL TRAIT MEASUREMENTS**

204 Standard photographs were taken from the lateral and dorsal view of 15 fish from each of the  
205 parental populations, the F<sub>1</sub> hybrids and 291 F<sub>2</sub>s at 12-15 months of age. Fish were  
206 anaesthetized with MS-222 (Sigma) and standardized photographs were taken from the dorsal  
207 and lateral views. Measurements from standardized photographs were performed using  
208 ImageJ software. A combination of linear and geometric morphometric measurements were  
209 employed to assess morphological variation associated with hypertrophied lips and head  
210 shape (Fig. S1).

211 Morphological traits values were obtained for between 284 (“Lip PC”) and 291  
212 (“LA”) F<sub>2</sub>s. The following linear measurements were considered: lip area (“LA”); upper lip  
213 area (“ULA”); lower lip area (“LLA”); lip length (“LL”); head length (“HL”) and head angle  
214 (“HA”) (Fig. S1). Geometric morphometrics was carried out to measure head shape (“HS”)  
215 and lip shape (“LS”). Eight landmarks and eight semi-landmarks were placed on the dorsal  
216 view of the fish. The landmarks were: (A1), (A2), (A8) and (A7) posterior and anterior  
217 extreme of the right and left orbit, (A3) and (A6) right and left starting point of the upper lip,  
218 (A4) tip of the upper lip, (A5) tip of the snout at the base of the upper lip. Semi-landmarks

219 were placed on the outlines of the snout and the upper lip (Fig. S1). HS and LS were  
220 measured by placing equally spaced semi-landmarks on each side of the dorsal view (A9-A16  
221 for LS, A17-A26 for HS), in relation to landmarks A2, A4, A5 and A7. Each landmark was  
222 digitized in tpsDig version 2.16 (Rohlf 2010a). Relative warps analysis was performed to  
223 remove all non-shape variation in tpsRelw version 1.49 (Rohlf 2010b). The positions of the  
224 semi-landmarks were moved along an estimated curve between the neighboring points to  
225 minimize squared distances between the adjusted positions and the corresponding points in  
226 the consensus. Statistical analysis of each shape was performed with classifier variables  
227 (species, sex) and with SL as a covariate in MorphoJ version 1.05f. Allometric effects were  
228 detracted from the shape using a pooled regression within each species with the Procrustes  
229 coordinates as dependent variables and SL as independent variable.

230         The relationships of all traits with standard length was tested using linear models in R.  
231 Those traits where the relationship with size was significant, were size-corrected by obtaining  
232 the residuals from a regression of each measurement on standard length. Of all the traits we  
233 investigated, only HS varied between sexes, which might reflect female adaptations for  
234 mouthbrooding. However, correction was deemed unnecessary because sex-corrected  
235 phenotypic values resulted in identical QTL mapping results, presumably because the  
236 relationship with sex does not interact statistically with species assignment. This makes  
237 biological sense since both species are mouthbrooders. Area measurements are the residuals  
238 of the regression on body area. Normality was tested using the Shapiro-Wilks test  
239 (shapiro.test function in R) and all traits were visually inspected using qqnorm and hist  
240 functions in R and the suggested quality control procedures of R/qtl (Broman and Sen 2009).  
241 All traits were normally distributed and were scaled to units of  $F_2$  phenotypic standard  
242 deviations. The signs of the phenotypic values of HA was reversed (multiplied by -1) for the

243 QTL mapping analysis [so that higher trait values were always present in \*H. chilotes\*](#) to reflect  
244 the [predicted adaptive direction](#) (see the “[Signatures of Natural Selection](#)” section below).

245 The traits were grouped in the following two categories (Fig. S1), Lip morphology  
246 (LM) and Head morphology (HM). LM comprises measurements of lip area (“LA”, “ULA”,  
247 “LLA”) and lip length (“LL”). HM includes measurements of head length (“HL”), head angle  
248 (“HA”) and head shape (“HS”). “LIP PC” consists of the first principal component from a  
249 PCA of a series of lip traits (Fig. S1). LIP PC was not included in any trait category because  
250 the measurement includes lip shape (LS) and is not independent from head shape thus  
251 rendering the analysis of genetic correlations uninterpretable. Measurement-correlated traits  
252 are independently affected by measurement error and/or capture different aspects of the  
253 phenotypic variation in complex traits. Because slightly different aspects of the traits are  
254 measured, the analysis of measurement-correlated traits allows the detection of a greater  
255 number of QTL that underlie biologically-relevant traits that are too complex to be  
256 represented by a single measurement. Discussing the extent of the shared genetic basis in  
257 these traits is biologically meaningless, but including them in the analysis is  
258 methodologically relevant because it creates an internal control of phenotyping, QTL  
259 mapping and QTL co-localization analysis (e.g. these traits should be correlated, share a  
260 significant amount of QTL and have similar QTL effect distributions).

## 261 **FORAGING PERFORMANCE**

262 We previously developed a laboratory assay to measure performance using a series of  
263 discrete angles (Baumgarten et al. 2015). In the present study, the acrylic glass structures  
264 were designed to yield a continuous measurement that is suitable for QTL mapping: the  
265 minimum angle that each fish (both parental and F<sub>2</sub>S) can forage on. The acrylic structures  
266 consist of an angle of 60° at the base that gradually reduces with height (15 cm), finishing

267 with an angle of 15° (Fig. 2C and Fig. S2). Small equally-sized pieces of mosquito larvae  
268 were placed at regular intervals of 3 mm along the inner vertical axis and were fixed by  
269 drying at 50 °C for 5 minutes. The experimental tanks (60x120x50 cm) were divided into four  
270 compartments using Plexiglas dividers. The experimental fish were starved for two days prior  
271 to the experiment and were transferred into the experimental compartments 24 hours before  
272 the beginning of each trial for acclimation. Non-experimental fish were maintained in the  
273 background compartment in order to maintain social interactions and improve the acclimation  
274 of the experimental fish.

275 Foraging performance was measured in ten individuals of each of the parental species  
276 (*H. chilotes* and *H. nyererei*) and 162 F<sub>2</sub> individuals. Phenotypic values are the residuals of  
277 the linear regression of the maximum foraging height on standard length. Due to the  
278 feasibility of collecting foraging data, the sample size is reduced compared to the  
279 morphological traits, which invariably leads to a less precise estimation of QTL effects.  
280 Nevertheless, care was taken to avoid bias and to ensure that the full spectrum of variation in  
281 HM and LM was represented in the foraging data. A video showing examples of the foraging  
282 trials is available in the supplementary files (Video S2).

## 283 **PHENOTYPIC CORRELATIONS IN F<sub>2</sub> HYBRIDS**

284 This analysis aimed at a) testing the contributions of the different morphological components  
285 to foraging performance, and; b) investigating the degree to which LM and HM segregate  
286 independently. The overall level of phenotypic correlation in the F<sub>2</sub> recombinant population  
287 was measured using Spearman's rho. The significance of the following correlations were  
288 tested: a) between morphological (LM, Lip PC and HM) and foraging traits and; b) between  
289 LM and HM. F<sub>2</sub> correlations emerge due to genetic correlations (when traits share QTL with  
290 concordant signs through linkage or pleiotropy) but also environmental correlations (*e.g.*



291 when one trait is the functional consequence of another trait or co-varies due to similar plastic  
292 responses).

### 293 **QUANTITATIVE TRAIT LOCI (QTL) MAPPING**

294 QTL mapping was performed for all traits using interval mapping (IM) and composite  
295 interval mapping (CIM) followed by a final evaluation using multiple interval mapping  
296 (MIM). In comparison to IM and CIM (or multiple QTL mapping – MQM), MIM has higher  
297 detection power, leads to more precise parameter estimates and allows for the simultaneous  
298 evaluation of interactions between detected QTL (Kao et al. 1999). However, QTL models  
299 based exclusively on MIM searches can be subject to over-parameterization as sample sizes  
300 decrease. To overcome these limitations, we took the following steps. The initial MIM model  
301 included all the QTL identified by IM and composite interval mapping, CIM using  
302 chromosome-wide LOD thresholds derived from 500 permutations and p-value cutoff of 0.05  
303 as inclusion criteria. IM was performed in R/qtl (Broman and Sen 2009) and WinQTL  
304 Cartographer v2.5 (available at <http://statgen.ncsu.edu/qtlcart/>). CIM and MIM analysis were  
305 conducted in WinQTL Cartographer v2.5. The positions of all QTL main effects included in  
306 the initial MIM models were optimized and the significance of each QTL main effect was  
307 tested. Non-significant QTL based on the BIC criteria and those with LOD scores below 2.5  
308 were excluded and model optimization proceeded as previously described (Silva et al. 2012).  
309 We aimed at the discovery of the maximum number of QTL and several observations suggest  
310 that the procedure we employed appropriately controlled for false-discovery: a) many of the  
311 suggestive QTL identified in the IM and CIM searches were eliminated from the initial model  
312 using the BIC model selection criteria; b) the amount of genetic variance explained does not  
313 suggest over-parametrization, and; c) the estimated QTL positions and effects are consistent  
314 across phenotypically correlated traits.

315 | [QTL were considered to co-localize when their 1-LOD intervals overlapped.](#)  
316 | Individual QTL effects, the total amount of phenotypic variance and the estimates of genetic  
317 | variance (i.e. broad sense heritability) were obtained from the variance decomposition tables  
318 | produced for the final MIM models in WinQTL Cartographer. Epistatic and dominant effects  
319 | were grouped and analyzed as non-additive because the current implementation of MIM in  
320 | WinQTL Cartographer only models epistatic interactions among QTL with significant main  
321 | effects. Interactions that are unaccounted for in the QTL model will resemble dominant  
322 | effects.

### 323 | **SIGNATURES OF NATURAL SELECTION**

324 | The distribution of effect signs was tested using the QTL sign test (QTLST) (Orr 1998b)  
325 | using a custom R function written by Muir et al. (2014). The sign test was only applied to Lip  
326 | PC and trait categories LM, HM because it only has power to reject the null hypothesis when  
327 | the number of detected QTL > 6. No individual trait other than ULA had as many detected  
328 | QTL with significant additive effects. Additive effects (in units of  $F_2$  standard deviation)  
329 | were pooled in each trait category (Albertson et al. 2003a) and in the event of shared QTL,  
330 | the effect with highest LOD support was selected for testing. Inclusion of the smallest co-  
331 | localized effects led to congruent results. All tests were conducted in R version 3.1.1 (Team  
332 | 2014). -In addition, the genome-wide additive effect estimates derived from interval mapping  
333 | were used to compare the mean effects of adaptive and non-adaptive traits. The mean effect  
334 | of LM, HM and FP was calculated from 10cM windows and was compared to the estimates  
335 | for traits that also differ between the parentals but are likely non-adaptive (body depth, anal  
336 | fin base and caudal peduncle length).

337 | The designation of “positive” or “negative” [allelic](#) effects is based on the direction of  
338 | adaptation for foraging in crevices [\(i.e., adaptive and maladaptive, respectively\)](#). “Positive”

339 effects are those that facilitate foraging in crevices. For most traits, the positive allelic effect  
340 increases the trait value, because hypertrophied lips and elongated heads are present in *H.*  
341 *chilotes*. The effects are reversed in the case of head angle (HA), because ~~*H. chilotes* has~~  
342 narrower and pointier heads facilitate foraging in crevices. To allow for the graphical  
343 comparison of the concordance of the effects of co-localizing QTL, the trait values of HA  
344 were reversed (multiplied by -1). Therefore, all alleles derived from *H. chilotes* (hereafter  
345 referred to as H alleles) are expected to increase trait values. Because we measured foraging  
346 performance in our mapping panel, the assignment of adaptive/maladaptive alleles was done  
347 directly by using the effect on foraging at the detected morphology QTL as a reference.

## 348 *Results*

### 349 **LINKAGE MAP CONSTRUCTION AND GENOME ANCHORING**

350 A saturated linkage map consisting of 1122 ddRAD markers distributed across 22 linkage  
351 groups with a total size of 1225.68cM was obtained, in agreement with the expectation based  
352 on the known haploid chromosome number in Haplochromine cichlids (Thompson 1981;  
353 Poletto et al. 2010) (Table S1). Eliminating redundancy led to the final linkage map used for  
354 QTL mapping that had 752 uniquely placed markers. The median interval size is 0.97cM,  
355 with 10 intervals larger than 10cM and a single interval (17cM) that is larger than 15cM  
356 (Table S2 and Fig. S3-S4). All but nine marker placements were congruent with the current  
357 *H. nyererei* draft genome sequence (Table S2). Two of these showed evidence of allelic  
358 dropout and were excluded from further analysis. Other incongruent markers showed no  
359 indications of genotyping errors and could be indicative of structural variations, genome  
360 fragmentation or mis-assembly. Comments highlighting the incongruences were added to  
361 Table S2.

362 The map of correspondence between our linkage map and the *H. nyererei* (Brawand et  
363 al. 2014) draft genome sequence shows a high level of congruency, which allowed for a high  
364 quality anchoring of the current scaffolds to our linkage map (Table S2). The comparison of  
365 genetic and physical distances did not point to the presence of large inversions segregating in  
366 our cross (*i.e.* no pairs of non-recombining markers that are separated by large physical  
367 distances were found). The physical distance between redundant markers and adjacent  
368 uniquely-placed markers ranged from 4bp to 1.86Mb (median = 83Kb) and 7Kb-6.36Mb  
369 (median = 424Kb), respectively. Therefore, marker redundancy is likely to have been caused  
370 by the close physical proximity of markers or small inversions in the parentals.

## 371 **PHENOTYPIC VARIATION AND CORRELATIONS IN F<sub>2</sub> HYBRIDS**

372 All traits, including foraging performance differed between the parental populations and  
373 segregated in the F<sub>2</sub> mapping panel (Fig. 2, Fig. S5). [Trait values that facilitate foraging were](#)  
374 [present in \*H. chilotes\* \(longer lips and narrower heads\)](#). Several LM measurements (LL, ULA  
375 and LA) and Lip PC were correlated with foraging performance with coefficients ranging  
376 between 0.19 and 0.27. HM traits showed no association with foraging performance, with the  
377 possible exception of HL, where a marginally non-significant relationship was found ( $P =$   
378  $0.08$ ,  $\rho = 0.13$ ). Variation in HM traits was also generally independent from variation in  
379 LM or LIPC traits ( $\rho = -0.08$  to  $0.15$ ), with the exception of two comparisons involving LA  
380 which were significant. All correlation coefficients are shown in Table 1.

## 381 **QTL MAPPING AND THE GENETIC ARCHITECTURE OF ADAPTATION**

382 The underlying genetic architecture was found to include many loci of small effect and a few  
383 of moderate effect (Table S3) that are distributed across all but three LGs (Fig. 3A). The  
384 number of detected QTL ranged from four to eleven. Some LGs have a high clumping of

385 QTL underlying all traits, indicating that these LGs have a moderate effect on most traits  
 386 investigated. Specifically, LG11, LG13, LG20 and LG23 are associated with several traits  
 387 across their entire length (Fig. 3A and Fig. S5). [The majority of the detected foraging](#)  
 388 [performance QTL co-localized with QTL underlying morphological traits, particularly LM](#)  
 389 [\(Fig. 3B\) and in some cases, there was co-localization of QTL that influence all trait groups](#)  
 390 [\(Fig. 3C\).](#) The largest effect QTL account for 12.6%, 7.1% and 9.3% of the F<sub>2</sub> phenotypic  
 391 variance in LM, HM and foraging performance, respectively. The distribution of effect sizes  
 392 we found suggests that our detection threshold is approximately 2% of F<sub>2</sub> phenotypic  
 393 variance (Fig. 4). In the final MIM QTL models, genetic effects on lip measurements are  
 394 composed of mainly additive effects that explain on average 31% of the phenotypic variance  
 395 (84% of the genetic variance). In contrast, additive effects accounted for only 22%  
 396 phenotypic (64.95% genetic variance) in HM and 23% (67.24% of the genetic variance) in  
 397 foraging performance traits (Table S4).

## 398 **GENOME-WIDE SIGNATURES OF NATURAL SELECTION**

399 **Heh** alleles were biased in their effect signs: The proportion of the positive additive effects  
 400 was 14/15 in LM, 10/13 in HM, 4/4 in foraging performance and 10/11 in the lip principal  
 401 component trait (Table S3). The QTL sign test rejected the null hypothesis in LM indicating  
 402 an excess of positive effect **Heh** alleles ( $P < 0.05$ ). [In the case of the QTL with overlapping](#)  
 403 [1-LD intervals between HM and LM, all H alleles are adaptive for LM but nearly half were](#)  
 404 [negative for HM \(Fig. 4\). In contrast, all of the H alleles at QTL unique to HM were adaptive.](#)  
 405 Furthermore, the inspection of genome-wide additive effect plots suggests that regions on  
 406 different chromosomes or on the same chromosome but genetically distant from detected  
 407 QTL also appear to have positive additive effect in the adaptive traits. As an example, the lip  
 408 area trait (LA) mapped to at least five genomic regions (Fig. 5A). Not only are all of the

409 additive effects positive in the detected QTL, but also in LGs where no QTL were detected,  
410 such as LGs 11-16 (Fig. 5B).

## 411 *Discussion*

412 We investigated the genetic basis of foraging performance, lip and head morphology in a  
413 cross between two haplochromine cichlid species from one of the youngest and largest  
414 known cichlid radiations, *Haplochromis chilotes* and *H. nyererei* (Brawand et al. 2014). The  
415 first species combines a suite of morphological adaptations that are associated with foraging  
416 in rocky crevices for invertebrate larvae and that evolved in most cichlid adaptive radiations  
417 (Keenleyside 1991). Deciphering the contribution of traits to fitness can prove complicated  
418 even in model systems (Cook et al. 2012; Zeller et al. 2012)) and while it is clear that natural  
419 selection has shaped morphology in many textbook examples of adaptation (Albertson et al.  
420 2003a), the primary target of selection (e.g. foraging capacity) certainly involves diverse  
421 classes of traits (e.g. metabolic, behavioral) in addition to morphology. This multifaceted  
422 aspect of adaptation might be expected to involve more complex interactions between loci  
423 (Huang et al. 2012) as well as a higher number of them (Arnegard et al. 2014). Our results  
424 show that it is possible to measure adaptive significance directly (Arnold 1983) also at the  
425 genetic level.

426 We found strong evidence for a role of hypertrophied lips in foraging success and that  
427 numerous loci were recruited since the divergence between these two species. These findings  
428 constitute strong support for the adaptive significance of hypertrophic lips (Endler 1986) and  
429 highlight the genome-wide effects of the response to natural selection of polygenic traits in  
430 recent adaptive radiations (Flaxman et al. 2013; Feder et al. 2014). The evolution of this  
431 multi-trait phenotype does not appear to be dominated by positive genetic correlations or  
432 small genetic target sizes, which can bias phenotypic evolution. Rather, the genetics of these  
433 adaptive differences in trophic morphology is consistent with a model of mostly small effect

434 loci, where only a few loci explain more than 5% of phenotypic variation and an increasing  
 435 number of loci smaller effects. Phenotypic correlations between trait groups were generally  
 436 low. Despite the detection of several co-localizing QTL, the effects ~~were many times~~  
 437 ~~antagonistic~~ were not always concordant revealing potential genetic trade-offs in the evolution  
 438 of hypertrophied lips and pointed heads.

439 It is predicted that major genes with pleiotropic function might be particularly  
 440 important in local adaptation in the presence of gene flow (Seehausen et al. 2014; Dittmar et  
 441 al. 2016; Ferris et al. 2016). We found evidence for the existence of positive genetic  
 442 associations (through either pleiotropy or tight linkage) and some evidence for clustering on  
 443 LG23. However, none of these factors seem to explain a large amount of between-species  
 444 differences. The genetic architecture of these traits is more aptly described as uncorrelated,  
 445 consisting of small-to-moderate additive effects across numerous loci. ~~This is in fact the~~  
 446 ~~genetic architecture that is thought to most commonly form the basis of quantitative traits~~  
 447 ~~(Flint and Mackay 2009)~~. However, this does not rule out that though currently small, it is  
 448 precisely the loci with positive correlation and larger effects that are important in the very  
 449 early instances of divergence or under high levels of gene flow. This should be investigated  
 450 with the comparison of the genome-wide pattern differentiation between *H. chilotes* and other  
 451 sympatric haplochromines.

## 452 **THE GENETIC ARCHITECTURE OF ADAPTATION IN THICK-LIPPED**

### 453 **CICHLIDS**

454 The genetic basis of lip measurements is composed of mainly additive effects across  
 455 numerous loci that are scattered throughout the genome. A large number of QTL of small  
 456 effect that individually explained up to 12% of F<sub>2</sub> phenotypic variation were detected on all  
 457 but three LGs. LG11 and LG23 are associated with multiple traits in what appear to be



458 multiple, closely-linked QTL. The genetic basis of the morphological traits we analyzed is  
459 consistent with that of other adaptive trophic morphologies analyzed in cichlids (e.g.  
460 Albertson et al. 2003a; Parnell et al. 2012; Albertson et al. 2014) and also with what is  
461 thought to be the most common genetic architecture underlying quantitative phenotypes in  
462 general (Albert et al. 2008; Flint and Mackay 2009).

463 These conclusions are only strengthened by considering that our estimates of effect  
464 sizes and number of QTL are likely to be overestimates and underestimates, respectively. The  
465 actual genetic architecture underlying these traits is probably composed of many more  
466 undetected QTL with small effects (Flint and Mackay 2009). The development of next-  
467 generation sequencing technologies facilitated the use of forward-genetics on non-model  
468 organisms (Schneeberger 2014) and today a large number of studies meet Orr and Coyne's  
469 third ~~criteria~~-criterion ("naturally occurring phenotypes"). However, considerable difficulties  
470 still exist for meeting Orr and Coyne's first ~~criteria~~-criterion ("sufficient power") in QTL  
471 mapping using non-model organisms. The size of the  $F_2$  panels are ~~but~~-only a fraction of  
472 those used for genetic investigations in established models (Beavis 1994; Fishman et al.  
473 2002; Laurie et al. 2004). The use of low sample sizes decreases the probability of detection  
474 of small effect QTL (*i.e.* increases the detection threshold), leads to biased estimates of effect  
475 sizes and insufficient power to disentangle the effects of closely-linked QTL (Beavis 1994;  
476 Xu 2003; Slate 2013). The size of the  $F_2$  mapping panel determines the detection power  
477 threshold and the extent of the inflation of effect sizes introduced by factors such as the  
478 Beavis effect (Slate 2013).

479 The existence of multiple crossable cichlid species pairs with different divergence  
480 times that differ in these same traits offers a unique opportunity to test whether alleles of  
481 large additive effect are recruited in the earlier stages of adaptation as predicted by Fisher's  
482 geometric model of adaptive evolution (Orr 2005; Rockman 2012). This is supported, for

483 example by work on sticklebacks (Rogers et al. 2012) and could be tested by further genetic  
 484 mapping projects in the multiple thick-lipped ecomorphs that occur in other recent radiations  
 485 such as the ones in Lake Malawi or the Midas cichlid radiation. Multiple ecologically-  
 486 divergent populations - from Lakes Nicaragua and Managua (*Amphilophus citrinellus* and *A.*  
 487 *labiatus*), as well as the recently colonized crater lakes - are variable for lip morphology  
 488 (Machado-Schiaffino et al. 2017). We have recently shown that phenotypic plasticity is an  
 489 important component of between-morph variation (e.g. Machado-Schiaffino et al. 2014) and  
 490 that genetic differences also exist between Neotropical morphs (Machado-Schiaffino et al.  
 491 2017).

## 492 **THE CAUSES OF THE REPEATED EVOLUTION OF THICK-LIPPED**

### 493 **CICHLIDS**

494 If the evolution of cichlid thick-lipped ecomorphs were facilitated by biases in the origin of  
 495 selectable variation, one would expect a large contribution from few loci to multiple traits. If  
 496 covariance dominated the genetic basis of ~~the lippy hypertrophic lipsecomorph~~, then a) HM  
 497 and LM would be expected to be largely positively correlated in F<sub>2</sub>s; b) a significant portion  
 498 of ~~the QTL for HM and LM would co-localize~~ the covariation between HM and LM would  
 499 be explained by co-localizing QTL and; c) the shared QTL would have concordant effects. In  
 500 contrast, we found that LM and HM segregated largely independently, with the exception of  
 501 two pairwise comparisons. Additional factors such as environmental variation or  
 502 measurement error might have contributed to a failure to detect phenotypic associations  
 503 between HM and the other classes of traits if the impact of these sources of errors be  
 504 imagined to be largely independent in the different trait groups. However, the high degree of  
 505 concordance between traits within the same trait groups suggests that measurement error did  
 506 not have a major role in our analysis.

507 |       Seven (out of 13) QTL for HM ~~overlapped~~ co-localized with QTL for LM when  
508 | considering an overlap of 1-LOD intervals but interestingly, three of them had negative  
509 | effects for HM. The mixture of concordant and discordant effects at shared QTL can result in  
510 | the masking of genetic correlations at the phenotypic level (Gardner and Latta 2007).  
511 | Distinguishing between close linkage and pleiotropy depends on the number of observed  
512 | crossovers and is one of the main limitations of QTL mapping experiments. Nevertheless, the  
513 | distinction between pleiotropy and linkage relates to *how little* recombination occurs between  
514 | loci, with the former representing the extreme case of complete linkage. It is possible that  
515 | close linkage has a similar effect to pleiotropy in rapid bursts of selection occurring in small  
516 | populations (Gardner and Latta 2007).

517 |       Although the overall level of genetic covariance of LM and HM is unlikely to have a  
518 | big effect in the response to selection, the presence of genetic trade-offs and antagonistic  
519 | pleiotropy might still have an impact on trait evolution (Via and Hawthorne 2002).  
520 | Overlapping QTL with concordant, positive effects were also found and it would be  
521 | interesting to test whether these are among the first to be recruited in the initial adaptation or  
522 | are important in adaptation through introgression. Likewise, lip area was weakly correlated  
523 | with two measurements of head morphology; and it would be interesting to test if this  
524 | correlation is stronger in earlier instances of adaptation. These hypotheses can be tested for  
525 | example by selection experiments in recombinant populations to analyze the fitness effects of  
526 | individual QTL (e.g. Rogers et al. 2012; Arnegard et al. 2014).

527 |       The large genetic target size of the phenotypes that we investigated does not support  
528 | the notion that similar phenotypes will be based on regions that are homologous to those that  
529 | we have identified, particularly when compared to more divergent taxa (*i.e.* African vs.  
530 | Neotropical cichlid radiations). However, because sharing of ancient genetic variation and  
531 | incomplete lineage sorting is rampant in East African cichlids (Brawand et al. 2014) it could

532 be true that the different African radiations have recruited ancient genetic variants. The  
533 accumulation of data linking genomic regions to evolutionarily relevant phenotypes in  
534 cichlids paves the way for exciting future research testing the importance of introgression and  
535 shared ancient genetic variation in cichlid adaptive radiations. It would be interesting to know  
536 how often convergent phenotypic evolution between the haplochromine cichlid radiations in  
537 Lakes Victoria, Malawi and Tanganyika involves the recruitment of ancient shared variation,  
538 as was shown to be the case in the colonization of freshwater from marine environments in  
539 sticklebacks (Colosimo et al. 2005; Jones et al. 2012). Hybridization is a common  
540 phenomenon in many groups of organisms, particularly in recently diverged species and its  
541 role in adaptation to new environments has been debated for a long time (Lewontin and Birch  
542 1966). However, conclusive evidence of adaptive introgression is restricted to a few systems  
543 where the phylogenetic analysis of causal genetic regions in hybridizing species was  
544 performed, such as *Heliconius* (Pardo-Diaz et al. 2012). Both contemporary and ancient  
545 hybridization seem widespread in cichlid fish (e.g. Koblmuller et al. 2010; Joyce et al. 2011;  
546 Genner and Turner 2012; Keller et al. 2013) and it has been proposed to play a crucial role in  
547 cichlid adaptive radiations, the “hybrid swarm hypothesis” (Seehausen 2004). Testing for  
548 both the role of introgression and incomplete lineage sorting in adaptation can be achieved by  
549 *functional phylogenomics*, systematically contrasting the evolutionary histories of several  
550 genomic regions identified by forward-genetic screens with random genomic regions using  
551 target enrichment (outlined in Henning and Meyer 2014). Despite the decreasing costs for  
552 whole-genome sequencing, target enrichment is still more efficient for collecting high-  
553 coverage, population-level data from large contiguous genomic regions. It has been used for  
554 applications such as phylogenomics, exon sequencing or population-based fine-mapping  
555 (Burbano et al. 2010; Mamanova et al. 2010; Faircloth et al. 2012; Lemmon et al. 2012;  
556 Nadeau et al. 2012).

557 **SIGNATURES OF NATURAL SELECTION**

558 Morphological differences in LM, particularly in lip length were strongly associated with  
559 foraging success. Genetic variation at the loci underlying morphology could be demonstrated  
560 to have an effect on foraging performance. Selection pressures in LM appear to be quite  
561 strong in natural conditions. This expectation was also confirmed by analyzing the  
562 distribution of effect signs. The null hypothesis of the distribution of QTL additive effect  
563 signs could be rejected for LM, thus supporting a role for directional natural selection in the  
564 evolution of these species differences. The QTL sign test we employed (QTL-ST) is  
565 conservative (Anderson and Slatkin 2003), tests for one particular scenario of natural  
566 selection (Orr 1998b) and is sensitive to variance in effect sizes (Rice and Townsend 2012).  
567 Therefore, the null hypothesis will only be rejected in extreme cases where the number of  
568 detected QTL is high and negative effects are virtually absent (e.g. Muir et al. 2014).  
569 Nevertheless, even with these restrictions it was possible to show that the observed  
570 abundance of positive effect alleles in LM traits is unlikely to have accumulated by chance.  
571 For comparison, although a large number of QTL responded to artificial selection for oil  
572 content in Maize in an even shorter timeframe, a great number of QTL (approx. 20%) had  
573 negative effects (Laurie et al. 2004). This suggests that the degree of selection for habitat  
574 partitioning in cichlid adaptive radiations is incredibly strong. In contrast, the persistence of  
575 negative effect alleles for HM, the overall distribution of additive effects and the weaker  
576 correlation with foraging performance all suggest a weaker or indirect selection pressure on  
577 HM. There are likely to be many additional QTL, given that the distribution of additive  
578 effects seems biased towards positive effects also in chromosomes where no QTL was  
579 detected. This suggests that many additional loci have diverged as the result of natural  
580 selection in an evolutionary timescale as short as 15 000 years.

581 Lip traits had the highest overall genome-wide effect with a median genome-wide  
582 additive effect = 0.1 (in units of  $F_2$  standard deviation) and a range of 0.07-0.135 for each  
583 trait. Despite the low number of detected QTL, foraging performance also had a high overall  
584 positive effect (0.087). The net effect of HM was also positive, albeit lower than the previous  
585 traits (median = 0.045, range = 0.038-0.05). Although the individual estimates are not  
586 independent owing to linkage, the overall median additive effects allows for a straightforward  
587 comparison of the influence of natural selection on different traits. When analyzing random  
588 traits, it is not possible to polarize trait values in relation to foraging in crevices as we have  
589 done for lip and head morphology traits. Nevertheless, species differences that are not the  
590 direct products of natural selection should not be biased towards any particular sign and  
591 should yield an overall value close to zero. This was the case with the traits that were taken  
592 for comparison (body depth, anal fin base and caudal peduncle length - phenotypic data not  
593 shown) which had a median effect very close to zero (-0.008). Note that this statement  
594 concerns only between-species differences and does not imply that traits evolve randomly.

595 It was hypothesized based on simulations that selection acting on a large fraction of  
596 the genome can lead to a non-linear and rapid build-up of reproductive isolation during  
597 speciation with gene-flow, leading to the process of whole-genome congealing (Flaxman et  
598 al. 2013; Feder et al. 2014). This pattern of QTL with biased effect signs throughout the  
599 genome has also been described in oral jaw traits that are important in the cichlid adaptive  
600 radiations (Albertson et al. 2003a) and could support the model of genome-wide congealing  
601 (Flaxman et al. 2013; Brawand et al. 2014; Feder et al. 2014), since the divergence of  
602 haplochromines occurred recently and under at least, partial gene flow. The accumulation of  
603 anchored genomes and QTL data pave the way for high-resolution studies on natural  
604 populations that could provide insights on the degree of genomic divergence that is  
605 associated with selection on lip morphology (Seehausen et al. 2014).

## 606 **CONCLUSIONS**

607 In summary, our results suggest that *i*) the loci underlying the morphological adaptations we  
608 investigated are numerous and have small additive effects; *ii*) foraging performance is  
609 functionally and genetically associated with between-species morphological differences,  
610 particularly in lip morphology; *iii*) the distribution of additive effects suggests that natural  
611 selection had a genome-wide effect; and that *iv*) variation in lip and head morphology is  
612 largely genetically independent. Genetic correlations between lip and head morphology are  
613 unlikely to facilitate concerted evolution and in fact might have constrained trait evolution  
614 through the tight coupling of discordant alleles or antagonistic pleiotropy. While recent  
615 empirical and theoretical work has highlighted the role of large effect variants and pleiotropy  
616 in the repeated evolution and the maintenance of adaptations (Ferris et al. 2016 and  
617 references therein), the present results show that this is certainly not a requirement for  
618 evolutionary convergence in adaptive radiations.

## 619 **ACKNOWLEDGEMENTS**

620 This work was supported by an Alexander von Humboldt fellowship to GMS, and grants  
621 from the DFG to GMS (MA6144/1-1) and to AM (ME 1725/18-1), from the CNPq/DAAD to  
622 FH (GDE-290049/2007-5) and from the European Research Council (ERC advanced grant  
623 ‘GenAdap’ 293700) to AM. We thank C. Chang-Rudolf for technical assistance and J.  
624 Torres-Dowdall, Karl Radtke, A. Kautt, Darrin Husley and C. Kratochwil for critical  
625 comments. Chris Muir provided the R code to run the QTL sign test. Ad Konings kindly  
626 permitted the use of images from the DVD “Malawi Cichlid Feeding Behavior” (Cichlid  
627 Press).

## 628 **LITERATURE CITED**



- 629 Albert, A. Y. K., S. Sawaya, T. H. Vines, A. K. Knecht, C. T. Miller, B. R. Summers, S.  
630 Balabhadra, D. M. Kingsley, and D. Schluter. 2008. The genetics of adaptive shape  
631 shift in stickleback: Pleiotropy and effect size. *Evolution* 62:76-85.
- 632 Albertson, R., J. Strelman, and T. Kocher. 2003a. Directional selection has shaped the oral  
633 jaws of Lake Malawi cichlid fishes. *Proc. Natl. Acad. Sci. U. S. A.* 100:5252-5257.
- 634 Albertson, R. C., K. E. Powder, Y. A. Hu, K. P. Coyle, R. B. Roberts, and K. J. Parsons.  
635 2014. Genetic basis of continuous variation in the levels and modular inheritance of  
636 pigmentation in cichlid fishes. *Mol. Ecol.* 23:5135-5150.
- 637 Albertson, R. C., J. T. Strelman, and T. D. Kocher. 2003b. Genetic basis of adaptive shape  
638 differences in the cichlid head. *J. Hered.* 94:291-301.
- 639 Anderson, E. C. and M. Slatkin. 2003. Orr's quantitative trait loci sign test under conditions  
640 of trait ascertainment. *Genetics* 165:445-446.
- 641 Arnegard, M. E., M. D. McGee, B. Matthews, K. B. Marchinko, G. L. Conte, S. Kabir, N.  
642 Bedford, S. Bergek, Y. F. Chan, F. C. Jones, D. M. Kingsley, C. L. Peichel, and D.  
643 Schluter. 2014. Genetics of ecological divergence during speciation. *Nature* 511:307-  
644 311.
- 645 Arnold, S. J. 1983. Morphology, Performance and Fitness. *Am. Zool.* 23:347-361.
- 646 Baumgarten, L., G. Machado-Schiaffino, F. Henning, and A. Meyer. 2015. What big lips are  
647 good for: on the adaptive function of repeatedly evolved hypertrophied lips of cichlid  
648 fishes. *Biol. J. Linn. Soc.* 115:448-455.
- 649 Beavis, A. D. 1994. The power and deceit of QTL experiments: Lessons from comparative  
650 QTL studies. Pp. 250-266. 49th Annual Corn & Sorghum Research Conference.
- 651 Brakefield, P. M. 2006. Evo-devo and constraints on selection. *Trends Ecol. Evol.* 21:362-  
652 368.
- 653 Brawand, D., C. E. Wagner, Y. I. Li, M. Malinsky, I. Keller, S. Fan, O. Simakov, A. Y. Ng,  
654 Z. W. Lim, E. Bezault, J. Turner-Maier, J. Johnson, R. Alcazar, H. J. Noh, P. Russell,  
655 B. Aken, J. Alföldi, C. Amemiya, N. Azzouzi, J.-F. Baroiller, F. Barloy-Hubler, A.  
656 Berlin, R. Bloomquist, K. L. Carleton, M. A. Conte, H. D'Cotta, O. Eshel, L. Gaffney,  
657 F. Galibert, H. F. Gante, S. Gnerre, L. Greuter, R. Guyon, N. S. Haddad, W. Haerty,  
658 R. M. Harris, H. A. Hofmann, T. Hourlier, G. Hulata, D. B. Jaffe, M. Lara, A. P. Lee,  
659 I. MacCallum, S. Mwaiko, M. Nikaido, H. Nishihara, C. Ozouf-Costaz, D. J. Penman,  
660 D. Przybylski, M. Rakotomanga, S. C. P. Renn, F. J. Ribeiro, M. Ron, W. Salzburger,  
661 L. Sanchez-Pulido, M. E. Santos, S. Searle, T. Sharpe, R. Swofford, F. J. Tan, L.  
662 Williams, S. Young, S. Yin, N. Okada, T. D. Kocher, E. A. Miska, E. S. Lander, B.  
663 Venkatesh, R. D. Fernald, A. Meyer, C. P. Ponting, J. T. Strelman, K. Lindblad-Toh,  
664 O. Seehausen, and F. Di Palma. 2014. The genomic substrate for adaptive radiation in  
665 African cichlid fish. *Nature* 513:375-381.
- 666 Broman, K. W. and S. Sen. 2009. A guide to QTL mapping with R/qlt. Springer, Dordrecht.
- 667 Burbano, H. A., E. Hodges, R. E. Green, A. W. Briggs, J. Krause, M. Meyer, J. M. Good, T.  
668 Maricic, P. L. F. Johnson, Z. Y. Xuan, M. Rooks, A. Bhattacharjee, L. Brizuela, F. W.  
669 Albert, M. de la Rasilla, J. Fortea, A. Rosas, M. Lachmann, G. J. Hannon, and S.  
670 Paabo. 2010. Targeted investigation of the Neandertal genome by array-based  
671 sequence capture. *Science* 328:723-725.
- 672 Burress, E. 2014. Cichlid fishes as models of ecological diversification: patterns,  
673 mechanisms, and consequences. *Hydrobiologia*:1-21.
- 674 Catchen, J. M., A. Amores, P. Hohenlohe, W. Cresko, and J. H. Postlethwait. 2011. Stacks:  
675 Building and genotyping loci de novo from short-read sequences. *G3-Genes Genomes*  
676 *Genetics* 1:171-182.
- 677 Chan, Y. F., M. E. Marks, F. C. Jones, G. Villarreal, M. D. Shapiro, S. D. Brady, A. M.  
678 Southwick, D. M. Absher, J. Grimwood, J. Schmutz, R. M. Myers, D. Petrov, B.

- 679 Jonsson, D. Schluter, M. A. Bell, and D. M. Kingsley. 2010. Adaptive evolution of  
680 pelvic reduction in sticklebacks by recurrent deletion of a *Pitx1* enhancer. *Science*  
681 327:302-305.
- 682 Colombo, M., E. T. Diepeveen, M. Muschick, M. E. Santos, A. Indermaur, N. Boileau, M.  
683 Barluenga, and W. Salzburger. 2013. The ecological and genetic basis of convergent  
684 thick-lipped phenotypes in cichlid fishes. *Mol. Ecol.* 22:670-684.
- 685 Colosimo, P. F., K. E. Hosemann, S. Balabhadra, G. Villarreal, M. Dickson, J. Grimwood, J.  
686 Schmutz, R. M. Myers, D. Schluter, and D. M. Kingsley. 2005. Widespread parallel  
687 evolution in sticklebacks by repeated fixation of ectodyplasin alleles. *Science*  
688 307:1928-1933.
- 689 Cook, L. M., B. S. Grant, I. J. Saccheri, and J. Mallet. 2012. Selective bird predation on the  
690 peppered moth: the last experiment of Michael Majerus. *Biol. Lett.*
- 691 Dittmar, E. L., C. G. Oakley, J. K. Conner, B. A. Gould, and D. W. Schemske. 2016. Factors  
692 influencing the effect size distribution of adaptive substitutions. *Proc. R. Soc. B* 283.
- 693 Ellegren, H., L. Smeds, R. Burri, P. I. Olason, N. Backstrom, T. Kawakami, A. Kunstner, H.  
694 Makinen, K. Nadachowska-Brzyska, A. Qvarnstrom, S. Uebbing, and J. B. W. Wolf.  
695 2012. The genomic landscape of species divergence in *Ficedula* flycatchers. *Nature*  
696 491:756-760.
- 697 Endler, J. A. 1986. Natural selection in the wild. Princeton University Press, Princeton, N.J.
- 698 Faircloth, B. C., J. E. McCormack, N. G. Crawford, M. G. Harvey, R. T. Brumfield, and T.  
699 C. Glenn. 2012. Ultraconserved elements anchor thousands of genetic markers  
700 spanning multiple evolutionary timescales. *Syst. Biol.* 61:717-726.
- 701 Feder, J. L., P. Nosil, A. C. Wacholder, S. P. Egan, S. H. Berlocher, and S. M. Flaxman.  
702 2014. Genome-wide congealing and rapid transitions across the speciation continuum  
703 during speciation with gene flow. *J. Hered.* 105:810-820.
- 704 Ferris, K. G., L. L. Barnett, B. K. Blackman, and J. H. Willis. 2016. The genetic architecture  
705 of local adaptation and reproductive isolation in sympatry within the *Mimulus*  
706 *guttatus* species complex. *Mol. Ecol.* [Early View \(doi. 10.1111/mec.13763\)](https://doi.org/10.1111/mec.13763)
- 707 Fishman, L., A. J. Kelly, and J. H. Willis. 2002. Minor quantitative trait loci underlie floral  
708 traits associated with mating system divergence in *Mimulus*. *Evolution* 56:2138-2155.
- 709 Flaxman, S. M., J. L. Feder, and P. Nosil. 2013. Genetic hitchhiking and the dynamic buildup  
710 of genomic divergence during speciation with gene flow. *Evolution* 67:2577-2591.
- 711 Flint, J. and T. F. C. Mackay. 2009. Genetic architecture of quantitative traits in mice, flies,  
712 and humans. *Genome Res.* 19:723-733.
- 713 Gardner, K. M. and R. G. Latta. 2007. Shared quantitative trait loci underlying the genetic  
714 correlation between continuous traits. *Mol. Ecol.* 16:4195-4209.
- 715 Genner, M. J. and G. F. Turner. 2012. Ancient hybridization and phenotypic novelty within  
716 Lake Malawi's cichlid fish radiation. *Mol. Biol. Evol.* 29:195-206.
- 717 Gompel, N. and B. Prud'homme. 2009. The causes of repeated genetic evolution. *Dev. Biol.*  
718 332:36-47.
- 719 Greenwood, A. K., J. N. Cech, and C. L. Peichel. 2012. Molecular and developmental  
720 contributions to divergent pigment patterns in marine and freshwater sticklebacks.  
721 *Evolution & Development* 14:351-362.
- 722 Greenwood, A. K., A. R. Wark, K. Yoshida, and C. L. Peichel. 2013. Genetic and neural  
723 modularity underlie the evolution of schooling behavior in Threespine Sticklebacks.  
724 *Curr. Biol.* 23:1884-1888.
- 725 Hall, M. C., C. J. Basten, and J. H. Willis. 2006. Pleiotropic quantitative trait loci contribute  
726 to population divergence in traits associated with life-history variation in *Mimulus*  
727 *guttatus*. *Genetics* 172:1829-1844.

- 728 Hendry, A. P. 2013. Key questions in the genetics and genomics of eco-evolutionary  
729 dynamics. *Heredity* 111:456-466.
- 730 Henning, F., H. J. Lee, P. Franchini, and A. Meyer. 2014. Genetic mapping of horizontal  
731 stripes in Lake Victoria cichlid fishes: benefits and pitfalls of using RAD markers for  
732 dense linkage mapping. *Mol. Ecol.* 23:5224-5240.
- 733 Henning, F. and A. Meyer. 2014. The evolutionary genomics of cichlid fishes: Explosive  
734 speciation and adaptation in the postgenomic era. *Annu. Rev. Genomics Hum. Genet.*  
735 15:417-441.
- 736 Huang, W., S. Richards, M. A. Carbone, D. H. Zhu, R. R. H. Anholt, J. F. Ayroles, L.  
737 Duncan, K. W. Jordan, F. Lawrence, M. M. Magwire, C. B. Warner, K. Blankenburg,  
738 Y. Han, M. Javaid, J. Jayaseelan, S. N. Jhangiani, D. Muzny, F. Onger, L. Perales, Y.  
739 Q. Wu, Y. Q. Zhang, X. Y. Zou, E. A. Stone, R. A. Gibbs, and T. F. C. Mackay. 2012.  
740 Epistasis dominates the genetic architecture of *Drosophila* quantitative traits. *Proc.*  
741 *Natl. Acad. Sci. U. S. A.* 109:15553-15559.
- 742 Johnson, T. C., C. A. Scholz, M. R. Talbot, K. Kelts, R. D. Ricketts, G. Ngobi, K. Beuning, I.  
743 Ssemmanda, and J. W. McGill. 1996. Late pleistocene desiccation of Lake Victoria  
744 and rapid evolution of cichlid fishes. *Science* 273:1091-1093.
- 745 Jones, F. C., M. G. Grabherr, Y. F. Chan, P. Russell, E. Mauceli, J. Johnson, R. Swofford, M.  
746 Pirun, M. C. Zody, S. White, E. Birney, S. Searle, J. Schmutz, J. Grimwood, M. C.  
747 Dickson, R. M. Myers, C. T. Miller, B. R. Summers, A. K. Knecht, S. D. Brady, H. L.  
748 Zhang, A. A. Pollen, T. Howes, C. Amemiya, E. S. Lander, F. Di Palma, K. Lindblad-  
749 Toh, D. M. Kingsley, B. I. G. S. Platf, and W. G. A. Team. 2012. The genomic basis  
750 of adaptive evolution in threespine sticklebacks. *Nature* 484:55-61.
- 751 Joyce, D. A., D. H. Lunt, M. J. Genner, G. F. Turner, R. Bills, and O. Seehausen. 2011.  
752 Repeated colonization and hybridization in Lake Malawi cichlids. *Curr. Biol.*  
753 21:R108-R109.
- 754 Kao, C. H., Z. B. Zeng, and R. D. Teasdale. 1999. Multiple interval mapping for quantitative  
755 trait loci. *Genetics* 152:1203-1216.
- 756 Keenleyside, M. H. A. 1991. *Cichlid fishes : behaviour, ecology and evolution*. Chapman &  
757 Hall, London.
- 758 Keller, I., C. E. Wagner, L. Greuter, S. Mwaiko, O. M. Selz, A. Sivasundar, S. Wittwer, and  
759 O. Seehausen. 2013. Population genomic signatures of divergent adaptation, gene  
760 flow and hybrid speciation in the rapid radiation of Lake Victoria cichlid fishes. *Mol.*  
761 *Ecol.* 22:2848-2863.
- 762 Koblmuller, S., B. Egger, C. Sturmbauer, and K. M. Sefc. 2010. Rapid radiation, ancient  
763 incomplete lineage sorting and ancient hybridization in the endemic Lake Tanganyika  
764 cichlid tribe Tropheini. *Mol. Phylogenet. Evol.* 55:318-334.
- 765 Kocher, T. D., J. A. Conroy, K. R. Mckaye, and J. R. Stauffer. 1993. Similar Morphologies of  
766 Cichlid Fish in Lakes Tanganyika and Malawi Are Due to Convergence. *Mol.*  
767 *Phylogenet. Evol.* 2:158-165.
- 768 Kowalko, J. E., N. Rohner, Santiago B. Rompani, Brant K. Peterson, Tess A. Linden, M.  
769 Yoshizawa, Emily H. Kay, J. Weber, Hopi E. Hoekstra, William R. Jeffery, R.  
770 Borowsky, and Clifford J. Tabin. 2013. Loss of schooling behavior in cavefish  
771 through sight-dependent and sight-independent mechanisms. *Curr. Biol.* 23:1874-  
772 1883.
- 773 Laland, K., T. Uller, M. Feldman, K. Sterelny, G. B. Muller, A. Moczek, E. Jablonka, and J.  
774 Odling-Smee. 2014. Does evolutionary theory need a rethink? - POINT Yes, urgently.  
775 *Nature* 514:161-164.
- 776 Lande, R. 1984. The genetic correlation between characters maintained by selection, linkage  
777 and inbreeding. *Genet. Res.* 44:309-320.

- 778 Laurie, C. C., S. D. Chasalow, J. R. LeDeaux, R. McCarroll, D. Bush, B. Hauge, C. Q. Lai,  
779 D. Clark, T. R. Rocheford, and J. W. Dudley. 2004. The genetic architecture of  
780 response to long-term artificial selection for oil concentration in the maize kernel.  
781 *Genetics* 168:2141-2155.
- 782 Lemmon, A. R., S. A. Emme, and E. M. Lemmon. 2012. Anchored hybrid enrichment for  
783 massively high-throughput phylogenomics. *Syst. Biol.* 61:727-744.
- 784 Lewontin, R. C. and L. C. Birch. 1966. Hybridization as a source of variation for adaptation  
785 to new environments. *Evolution* 20:315-&.
- 786 Linnen, C. R., Y. P. Poh, B. K. Peterson, R. D. H. Barrett, J. G. Larson, J. D. Jensen, and H.  
787 E. Hoekstra. 2013. Adaptive evolution of multiple traits through multiple mutations at  
788 a single gene. *Science* 339:1312-1316.
- 789 Losos, J. B. 2011. Convergence, Adaptation, and Constraint. *Evolution* 65:1827-1840.
- 790 Machado-Schiaffino, G., F. Henning, and A. Meyer. 2014. Species-specific differences in  
791 adaptive phenotypic plasticity in an ecologically relevant trophic trait: Hypertrophic  
792 lips in Midas cichlid fishes. *Evolution*:286-291.
- 793 Machado-Schiaffino, G., A. Kautt, J. Torres-Dowdall, L. Baumgarten, F. Henning, F and A.  
794 Meyer. 2017. Incipient speciation driven by hypertrophied lips in Midas cichlid  
795 fishes. *Molecular Ecology* (in press).
- 796 Mamanova, L., A. J. Coffey, C. E. Scott, I. Kozarewa, E. H. Turner, A. Kumar, E. Howard, J.  
797 Shendure, and D. J. Turner. 2010. Target-enrichment strategies for next-generation  
798 sequencing. *Nat. Meth.* 7:111-118.
- 799 Manousaki, T., P. M. Hull, H. Kusche, G. Machado-Schiaffino, P. Franchini, C. Harrod, K.  
800 R. Elmer, and A. Meyer. 2013. Parsing parallel evolution: ecological divergence and  
801 differential gene expression in the adaptive radiations of thick-lipped Midas cichlid  
802 fishes from Nicaragua. *Mol. Ecol.* 22:650-669.
- 803 Miller, C. T., A. M. Glazer, B. R. Summers, B. K. Blackman, A. R. Norman, M. D. Shapiro,  
804 B. L. Cole, C. L. Peichel, D. Schluter, and D. M. Kingsley. 2014. Modular skeletal  
805 evolution in sticklebacks is controlled by additive and clustered quantitative trait loci.  
806 *Genetics* 197:405-420.
- 807 Muir, C. D., J. B. Pease, and L. C. Moyle. 2014. Quantitative genetic analysis indicates  
808 natural selection on leaf phenotypes across wild tomato species (*Solanum sect.*  
809 *lycopersicon*; Solanaceae). *Genetics* 198:1629-1643.
- 810 Nadeau, N. J., A. Whibley, R. T. Jones, J. W. Davey, K. K. Dasmahapatra, S. W. Baxter, M.  
811 A. Quail, M. Joron, R. H. Ffrench-Constant, M. L. Blaxter, J. Mallet, and C. D.  
812 Jiggins. 2012. Genomic islands of divergence in hybridizing *Heliconius* butterflies  
813 identified by large-scale targeted sequencing. *Philosophical Transactions of the Royal*  
814 *Society B-Biological Sciences* 367:343-353.
- 815 O'Quin, C. T., A. C. Drilea, M. A. Conte, and T. D. Kocher. 2013. Mapping of pigmentation  
816 QTL on an anchored genome assembly of the cichlid fish, *Metriaclima zebra*. *BMC*  
817 *Genomics* 14:287.
- 818 O'Quin, C. T., A. C. Drilea, R. B. Roberts, and T. D. Kocher. 2012. A small number of genes  
819 underlie male pigmentation traits in Lake Malawi cichlid fishes. *Journal of*  
820 *Experimental Zoology Part B-Molecular and Developmental Evolution* 318B:199-  
821 208.
- 822 Oliver, M. K. and M. E. Arnegard. 2010. A new genus for *Melanochromis labrosus*, a  
823 problematic Lake Malawi cichlid with hypertrophied lips (Teleostei: Cichlidae).  
824 *Ichthyol. Explor. Fres.* 21:209-232.
- 825 Orr, H. A. 1998a. The population genetics of adaptation: The distribution of factors fixed  
826 during adaptive evolution. *Evolution* 52:935-949.



- 827 Orr, H. A. 1998b. Testing natural selection vs. genetic drift in phenotypic evolution using  
828 quantitative trait locus data. *Genetics* 149:2099-2104.
- 829 Orr, H. A. 2005. The genetic theory of adaptation: A brief history. *Nat. Rev. Genet.* 6:119-  
830 127.
- 831 Orr, H. A. and J. A. Coyne. 1992. The genetics of adaptation - a reassessment. *Am. Nat.*  
832 140:725-742.
- 833 Pardo-Diaz, C., C. Salazar, S. W. Baxter, C. Merot, W. Figueiredo-Ready, M. Joron, W. O.  
834 McMillan, and C. D. Jiggins. 2012. Adaptive introgression across species boundaries  
835 in *Heliconius* butterflies. *PLoS Genetics* 8.
- 836 Parnell, N. F., C. D. Hulsey, and J. T. Streebman. 2012. The genetic basis of a complex  
837 functional system. *Evolution* 66:3352-3366.
- 838 Peterson, B. K., J. N. Weber, E. H. Kay, H. S. Fisher, and H. E. Hoekstra. 2012. Double  
839 digest RADseq: An inexpensive method for *de novo* SNP discovery and genotyping in  
840 model and non-model species. *Plos One* 7.
- 841 Poelstra, J. W., N. Vijay, C. M. Bossu, H. Lantz, B. Ryll, I. Muller, V. Baglione, P.  
842 Unneberg, M. Wikelski, M. G. Grabherr, and J. B. W. Wolf. 2014. The genomic  
843 landscape underlying phenotypic integrity in the face of gene flow in crows. *Science*  
844 344:1410-1414.
- 845 Rice, D. P. and J. P. Townsend. 2012. Resampling QTL effects in the QTL sign test leads to  
846 incongruous sensitivity to variance in effect size. *G3-Genes Genomes Genetics* 2:905-  
847 911.
- 848 Roberts, R. B., J. R. Ser, and T. D. Kocher. 2009. Sexual conflict resolved by invasion of a  
849 novel sex determiner in Lake Malawi cichlid fishes. *Science* 326:998-1001.
- 850 Rockman, M. V. 2012. The QTN program and the alleles that matter for evolution: All that's  
851 gold does not glitter. *Evolution* 66:1-17.
- 852 Rogers, S. M., P. Tamkee, B. Summers, S. Balabhadra, M. Marks, D. M. Kingsley, and D.  
853 Schluter. 2012. Genetic Signature of Adaptive Peak Shift in Threespine Stickleback.  
854 *Evolution* 66:2439-2450.
- 855 Rohlf, F. 2010a. tpsDig version 2.16, Department of Ecology and Evolution, State University  
856 of New York at Stony Brook. Available at <http://life.bio.sunysb.edu/morph/>.
- 857 Rohlf, F. 2010b. tpsRelw, version 1.49, Department of Ecology and Evolution, State  
858 University of New York at Stony Brook. Available at  
859 <http://life.bio.sunysb.edu/morph/>.
- 860 Schluter, D. 1996. Adaptive radiation along genetic lines of least resistance. *Evolution*  
861 50:1766-1774.
- 862 Schneeberger, K. 2014. Using next-generation sequencing to isolate mutant genes from  
863 forward genetic screens. *Nat. Rev. Genet.* advance online publication.
- 864 Seehausen, O. 1996. Distribution of and reproductive isolation among color morphs of a  
865 rock-dwelling Lake Victoria cichlid (*Haplochromis nyererei*). *Ecol. Freshw. Fish.*  
866 5:195-202.
- 867 Seehausen, O. 2004. Hybridization and adaptive radiation. *Trends Ecol. Evol.* 19:198-207.
- 868 Seehausen, O. and N. Bouton. 1998. The community of rock-dwelling cichlids in Lake  
869 Victoria. *Bonner Zoologische Beiträge* 47:301-312.
- 870 Seehausen, O., R. K. Butlin, I. Keller, C. E. Wagner, J. W. Boughman, P. A. Hohenlohe, C.  
871 L. Peichel, G.-P. Saetre, C. Bank, A. Brannstrom, A. Brelsford, C. S. Clarkson, F.  
872 Eroukhanoff, J. L. Feder, M. C. Fischer, A. D. Foote, P. Franchini, C. D. Jiggins, F.  
873 C. Jones, A. K. Lindholm, K. Lucek, M. E. Maan, D. A. Marques, S. H. Martin, B.  
874 Matthews, J. I. Meier, M. Most, M. W. Nachman, E. Nonaka, D. J. Rennison, J.  
875 Schwarzer, E. T. Watson, A. M. Westram, and A. Widmer. 2014. Genomics and the  
876 origin of species. *Nat. Rev. Genet.* 15:176-192.

- 877 Servedio, M. R., G. S. Doorn, M. Kopp, A. M. Frame, and P. Nosil. 2011. Magic traits in  
878 speciation: 'magic' but not rare? *Trends in Ecology and Evolution* 26:389-397.
- 879 Silva, L. D. E., S. Wang, and Z. B. Zeng. 2012. Composite interval mapping and multiple  
880 interval mapping: Procedures and guidelines for using windows QTL Cartographer.  
881 Pp. 75-120 in S. A. Rifkin, ed. *Quantitative trait loci (QTL) : methods and protocols*.  
882 Humana Press, Springer, New York.
- 883 Slate, J. 2013. From Beavis to beak color: A simulation study to examine how much QTL  
884 mapping can reveal about the genetic architecture of quantitative traits. *Evolution*  
885 67:1251-1262.
- 886 Stelkens, R. B., K. A. Young, and O. Seehausen. 2010. The accumulation of reproductive  
887 incompatibilities in African cichlid fish. *Evolution* 64:617-632.
- 888 Streelman, J. T., R. C. Albertson, and T. D. Kocher. 2003. Genome mapping of the orange  
889 blotch colour pattern in cichlid fishes. *Mol. Ecol.* 12:2465-2471.
- 890 Team, R. C. 2014. R: A Language and Environment for Statistical Computing. R Foundation  
891 for Statistical Computing, Vienna, Austria.
- 892 van't Hof, A. E., N. Edmonds, M. Dalikova, F. Marec, and I. J. Saccheri. 2011. Industrial  
893 melanism in British peppered moths has a singular and recent mutational origin.  
894 *Science* 332:958-960.
- 895 van Oijen, M. J. P. 1996. The generic classification of the haplochromine cichlids of Lake  
896 Victoria, East Africa. *Zool. Verh. Leiden* 302:57-110.
- 897 Van Ooijen, J. W. 2006. JoinMap 4. Software for the calculation of genetic linkage maps in  
898 experimental populations. Kyazma BV, Wageningen, The Netherlands: .
- 899 Via, S. and D. J. Hawthorne. 2002. The genetic architecture of ecological specialization:  
900 Correlated gene effects on host use and habitat choice in pea aphids. *Am. Nat.*  
901 159:S76-S88.
- 902 Weber, J. N., B. K. Peterson, and H. E. Hoekstra. 2013. Discrete genetic modules are  
903 responsible for complex burrow evolution in *Peromyscus mice*. *Nature* 493:402-405.
- 904 Witte, F. and M. J. P. Van Oijen. 1990. Taxonomy, ecology and fishery of Lake Victoria  
905 haplochromine trophic groups. *Zoologische Verhandelingen* 262:1-47.
- 906 Wray, G. A., H. E. Hoekstra, D. J. Futuyma, R. E. Lenski, T. F. C. Mackay, D. Schluter, and  
907 J. E. Strassmann. 2014. Does evolutionary theory need a rethink? -  
908 COUNTERPOINT No, all is well. *Nature* 514:161-164.
- 909 Wu, T. D. and C. K. Watanabe. 2005. GMAP: a genomic mapping and alignment program  
910 for mRNA and EST sequences. *Bioinformatics* 21:1859-1875.
- 911 Xu, S. Z. 2003. Theoretical basis of the Beavis effect. *Genetics* 165:2259-2268.
- 912 Zeller, M., K. Lucek, M. Haesler, O. Seehausen, and A. Sivasundar. 2012. Little evidence for  
913 a selective advantage of armour-reduced threespined stickleback individuals in an  
914 invertebrate predation experiment. *Evol. Ecol.* 26:1293-1309.

915

916 **TABLES**

917 Table 1. Phenotypic correlation matrix of F<sub>2</sub> hybrids of all measured traits. Lip area (“LA”),  
 918 upper lip area (“ULA”), lower lip area (“LLA”), lip length (“LL”), lip principal component  
 919 (“LIP PC”), head length (“HL”), head angle (“HA”), head shape (“HS”) and foraging  
 920 performance (“FP”). The values of Spearman’s rank correlation coefficient are given below  
 921 the diagonal. Correlations between different trait groups are highlighted in bold. \*, \*\* and  
 922 \*\*\* represent *P* values of < 0.05, <0.01 and <0.001, respectively.

	LA	ULA	LLA	LL	LIPC	HA	HL	HS
ULA	0.579***							
LLA	0.545***	0.224***						
LL	0.497***	0.437***	0.298***					
LIPC	0.809***	0.716***	0.682***	0.729***				
HA	<b>-0.001</b>	<b>0.073</b>	<b>-0.059</b>	<b>-0.04</b>	-0.014			
HL	<b>0.149*</b>	<b>0.055</b>	<b>0.066</b>	<b>0.108</b>	0.078	-0.302***		
HS	<b>0.129*</b>	<b>-0.016</b>	<b>0.076</b>	<b>-0.042</b>	0.023	-0.241***	0.238***	
FP	<b>0.241**</b>	<b>0.186*</b>	<b>0.107</b>	<b>0.252***</b>	<b>0.266***</b>	<b>-0.087</b>	<b>0.13</b>	<b>0.075</b>

923 **FIGURE LEGENDS**

924 **Figure 1.** Convergent evolution and function of hypertrophic lips. Representative species are  
 925 shown from the cichlid radiations of the African great lakes, Central America and South  
 926 America. Photographs were kindly provided by Erwin Schraml, Ad Konings and Oliver  
 927 Lucanus. In the image sequence on the bottom, an individual *Placidochromis milomo*  
 928 (representative of the lake Malawi radiation) is seen searching for prey (left), targeting a  
 929 rocky crevice (center) and accessing the prey (right).

930 **Figure 2.** Foraging performance and morphological traits are correlated and segregate in F<sub>2</sub>  
 931 hybrids. A) Male specimens of both species used in the experiment. B) Distribution of  
 932 phenotypic values in representative traits in the parental and F<sub>2</sub> populations. C) -The acrylic



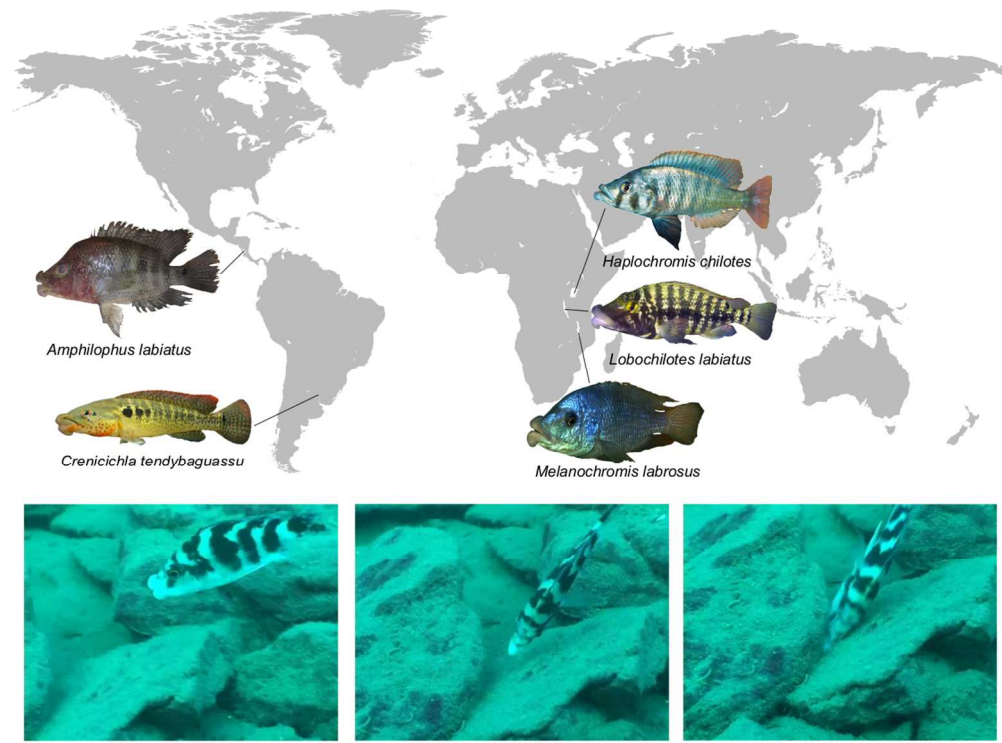
933 device (left) and the experimental setting (right) developed to measure foraging performance.  
934 D) Differences between the parental and F<sub>2</sub> populations in foraging performance. E) The  
935 correlation between foraging performance and lip length. Spearman's correlation coefficient  
936 is shown.

937 **Figure 3.** QTL map of foraging performance and associated morphological traits. A)  
938 Distribution of all detected main effect QTL for [all trait groups](#): foraging performance ("FP",  
939 green), lip morphology ("LM", blue), head morphology ("HM", red) and lip principal  
940 component ("LIP PC", black). The map distance in cM is given by the scales on the left.  
941 Thick and thin bars represent the 1- and 2- LOD intervals, respectively. [B-C\) Overlapping](#)  
942 [LOD profiles of QTL for different trait groups \(shown with arrowheads in A\). To avoid](#)  
943 [redundancy, only the most highly supported QTL from each of the different trait groups](#)  
944 [\("LM", "HM", "LIP PC" and "FP"\) are shown.](#) The overlap of the 1-LOD intervals is  
945 represented by the grey boxes.

946 [Figure 4. Direction and distribution of QTL effects. A-B\) Concordant and antagonistic allelic](#)  
947 [effects at co-localizing QTL for lip and head morphology \(in units of F<sub>2</sub> standard deviation\).](#)  
948 [Alleles inherited from \*Haplochromis chilotes\* and from \*H. nyererei\* are represented by "H"](#)  
949 [and "N", respectively. Slopes of opposite signs are indicative of antagonistic effects because](#)  
950 [all traits were polarized with regards to foraging in crevices. H alleles are expected to](#)  
951 [increase phenotypic values for all traits \(see Methods\). Intersecting effect slopes are more](#)  
952 [apparent in the comparison between homozygous genotypes \(CC and NN\) since non-additive](#)  
953 [genetic variation can result in CN genotypes having phenotypic values above or below the](#)  
954 [expected under a purely additive model \(e.g. HS at LG17 or HA at LG13\). C\) Distribution of](#)  
955 detected additive effects. Effect sizes are expressed in percentage of explained F<sub>2</sub> phenotypic  
956 variance.

957 **Figure 5.** QTL map of lip area (“LA”). A) At least five genomic regions underlie phenotypic  
958 variation in LA. The LOD profiles for the three different detection methods (IM, CIM and  
959 MIM) are shown and are largely congruent. The dark and light horizontal lines represent the  
960 genome-wide (3.7) and chromosome-wide (2.5) significance thresholds for IM and CIM. -All  
961 MIM QTL that are shown are significant using the BIC criteria. B) All detected QTL have a  
962 positive additive effect (in standard deviation units).

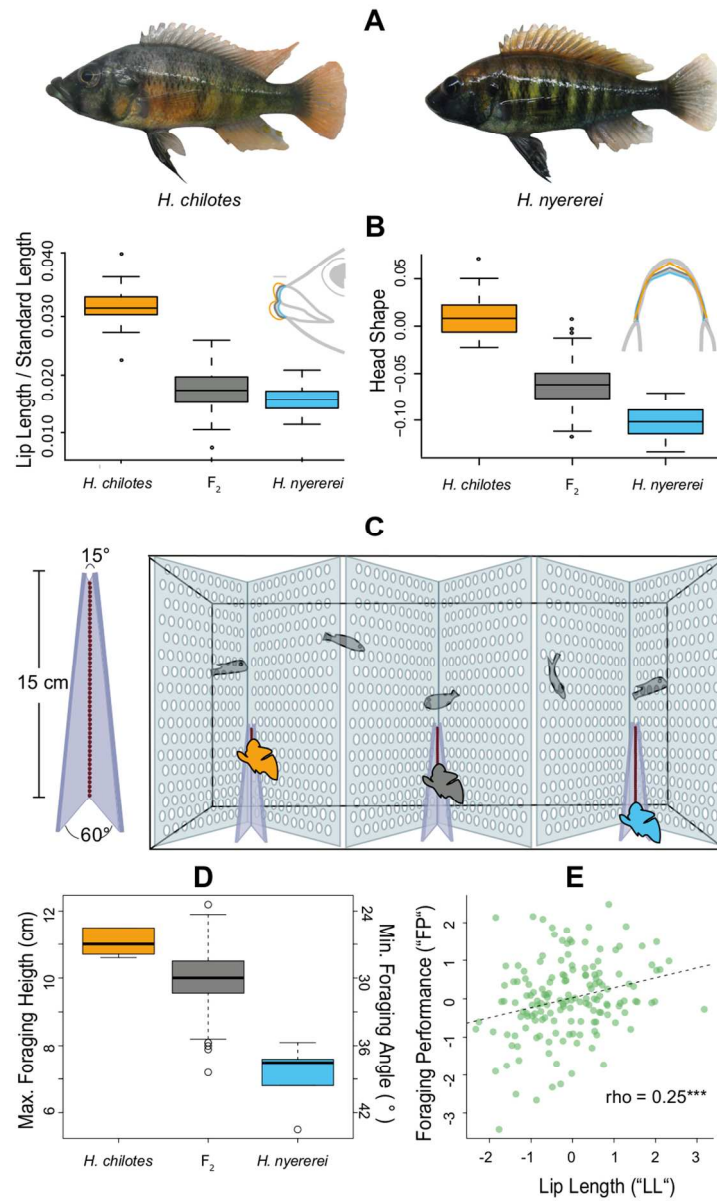
For Peer Review Only



**Figure 1.** Convergent evolution and function of hypertrophic lips. Representative species are shown from the cichlid radiations of the African great lakes, Central America and South America. Photographs were kindly provided by Erwin Schraml, Ad Konings and Oliver Lucanus. In the image sequence on the bottom, an individual *Placidochromis milomo* (representative of the lake Malawi radiation) is seen searching for prey (left), targeting a rocky crevice (center) and accessing the prey (right).

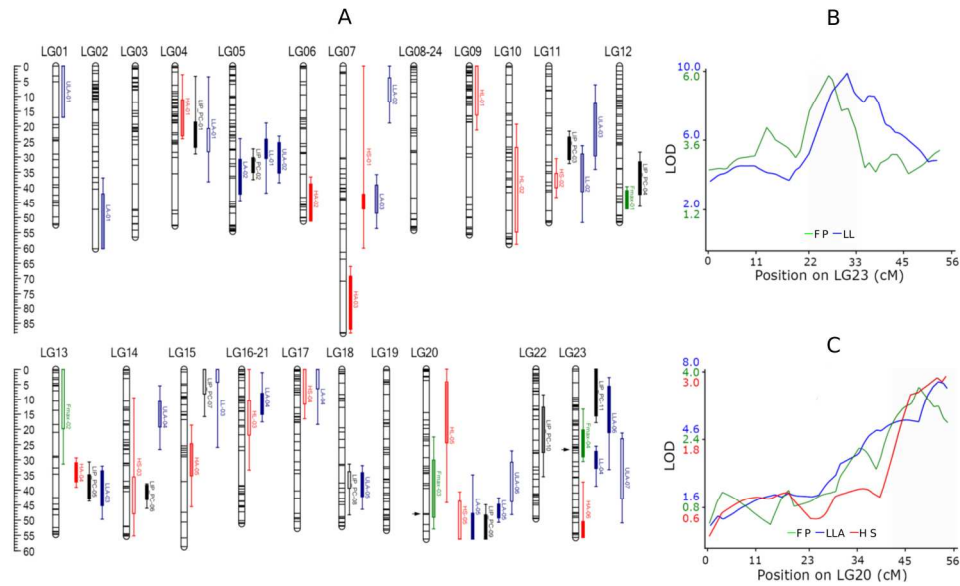
Fig. 1  
115x87mm (300 x 300 DPI)

Only



**Figure 2.** Foraging performance and morphological traits are correlated and segregate in F2 hybrids. A) Male specimens of both species used in the experiment. B) Distribution of phenotypic values in representative traits in the parental and F2 populations. C) The acrylic device (left) and the experimental setting (right) developed to measure foraging performance. D) Differences between the parental and F2 populations in foraging performance. Spearman's correlation coefficient is shown. E) The correlation between foraging performance and lip length.

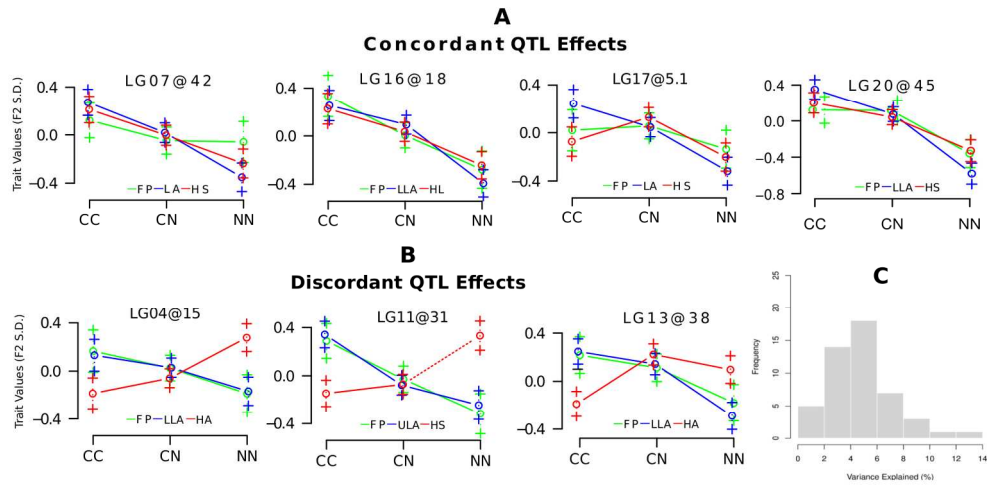
Fig. 2  
86x143mm (300 x 300 DPI)



**Figure 3.** QTL map of foraging performance and associated morphological traits. A) Distribution of all detected main effect QTL for all trait groups: foraging performance ("FP", green), lip morphology ("LM", blue), head morphology ("HM", red) and lip principal component ("LIP PC", black). The map distance in cM is given by the scales on the left. Thick and thin bars represent the 1- and 2- LOD intervals, respectively. B-C) Overlapping LOD profiles of QTL for different trait groups (shown with arrowheads in A). To avoid redundancy, only the most highly supported QTL from each of the different trait groups ("LM", "HM", "LIP PC" and "FP") are shown. The overlap of the 1-LOD intervals is represented by the grey boxes.

Fig. 3

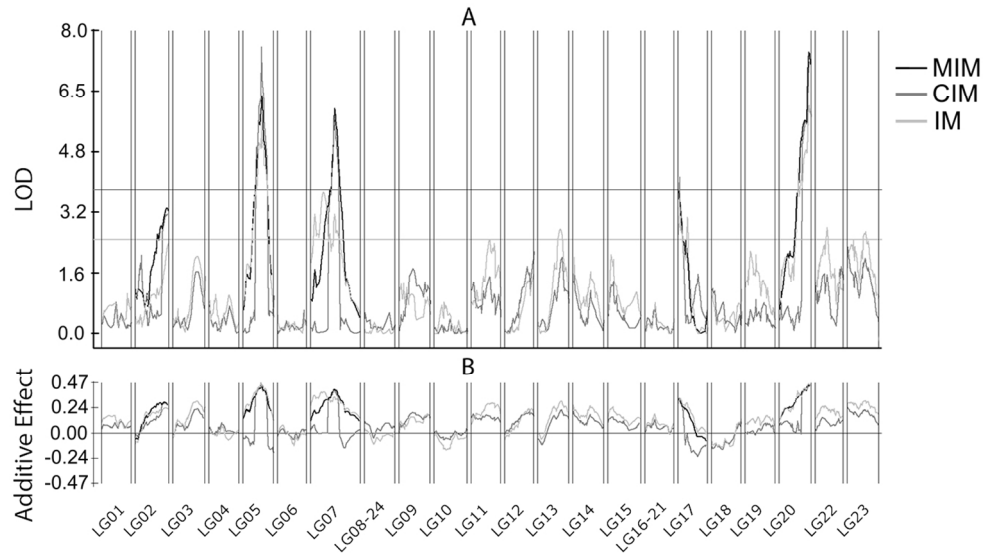
178x106mm (300 x 300 DPI)



**Figure 4.** Direction and distribution of QTL effects. A-B) Concordant and antagonistic allelic effects at co-localizing QTL for lip and head morphology (in units of F2 standard deviation). Alleles inherited from *Haplochromis chilotes* and from *H. nyererei* are represented by “H” and “N”, respectively. Slopes of opposite signs are indicative of antagonistic effects because all traits were polarized with regards to foraging in crevices. H alleles are expected to increase phenotypic values for all traits (see Methods). Intersecting effect slopes are more apparent in the comparison between homozygous genotypes (CC and NN) since non-additive genetic variation can result in CN genotypes having phenotypic values above or below the expected under a purely additive model (e.g. HS at LG17 or HA at LG13). C) Distribution of detected additive effects. Effect sizes are expressed in percentage of explained F2 phenotypic variance.

Fig. 4

177x92mm (300 x 300 DPI)



**Figure 5.** QTL map of lip area ("LA"). A) At least five genomic regions underlie phenotypic variation in LA. The LOD profiles for the three different detection methods (IM, CIM and MIM) are shown and are largely congruent. The dark and light horizontal lines represent the genome-wide (3.7) and chromosome-wide (2.5) significance thresholds for IM and CIM. All MIM QTL that are shown are significant using the BIC criteria. B) All detected QTL have a positive additive effect (in standard deviation units).

Fig. 5

114x63mm (300 x 300 DPI)