

DE NOVO BIOSYNTHESIS OF O-METHYLATED FLAVONOIDS IN THE MICROBIAL FACTORY *STREPTOMYCES ALBUS*

Álvaro Pérez-Valero^{1,2,3}, Suhui Ye^{1,2,3}, Patricia Magadán-Corpas^{1,2,3}, Claudio J. Villar^{1,2,3}, Felipe Lombó^{1,2,3}.

¹ Research Unit BIONUC (Biotechnology in Nutraceuticals and Bioactive Compounds), Departamento de Biología Funcional, Área de Microbiología, Universidad de Oviedo, Oviedo, Spain.

² IUOPA (Instituto Universitario de Oncología del Principado de Asturias), Oviedo, Spain

³ ISPA (Instituto de Investigación Sanitaria del Principado de Asturias), Oviedo, Spain

Corresponding author email: lombofelipe@uniovi.es

Among natural products, flavonoids are highly attractive due to their promising effects in treating cancer, pathogenic bacteria, oxidative stress, cardio-vascular, inflammatory dysfunctions, etc. Particularly, methylated flavonoids are very interesting since the presence of methyl groups dramatically increases their stability and enhances the membrane transport, facilitating absorption and improving oral bioavailability [1]. Heterologous production of these plant compounds in engineered microbial cell factories is becoming a good alternative to traditional production in plants, saving time, space and avoiding plant requirements as specific soil or certain climatic conditions. The microbial host *Streptomyces albus* has demonstrated to be a good candidate for heterologous biosynthesis of flavonoids [2–4]. In this study, an engineered strain of *S. albus* (with various genomic editions, including native biosynthetic gene clusters deletions) has been selected to produce various O-methylated flavonoids due to its optimized production of the flavonoid intermediate naringenin. The naringenin biosynthesis gene cluster has been integrated into the chromosome of this bacteria using the Φ C31 phage integrase. In order to produce methylated derivatives, other genes, including methyltransferases, have been integrated into the Φ BT1 site of the bacterial chromosome, generating various strains able to produce methylated flavonoids such as sakuranetin, acacetin or genkwanin.

References

1. Koirala, N.; Thuan, N.H.; Ghimire, G.P.; Thang, D. Van; Sohng, J.K. Methylation of flavonoids: Chemical structures, bioactivities, progress and perspectives for biotechnological production. *Enzyme Microb. Technol.* 2016, *86*, 103–116.
2. Marín, L.; Gutiérrez-del-Río, I.; Yagüe, P.; Manteca, Á.; Villar, C.J.; Lombó, F. De novo biosynthesis of apigenin, luteolin, and eriodictyol in the actinomycete *Streptomyces albus* and production improvement by feeding and spore conditioning. *Front. Microbiol.* **2017**, *8*, 1–12, doi:10.3389/fmicb.2017.00921.
3. Marín, L.; Gutiérrez-del-Río, I.; Entrialgo-Cadierno, R.; Claudio, Villar, J.; Lombó, F. De novo biosynthesis of myricetin, kaempferol and quercetin in *Streptomyces albus* and *Streptomyces coelicolor*. *PLoS One* **2018**, *13*, 1–16, doi:10.1371/journal.pone.0207278.
4. Marín, L.; Gutiérrez-del-Río, I.; Villar, C.J.; Lombó, F. De novo biosynthesis of garbanzol and fustin in *Streptomyces albus* based on a potential flavanone 3-hydroxylase with 2-hydroxylase side activity. *Microb. Biotechnol.* **2021**, *14*, 2009–2024, doi:10.1111/1751-7915.13874.