

# Molecularly imprinted polymer-based electrochemical sensors for environmental analysis

Patrícia Rebelo<sup>a, c</sup>, Estefanía Costa-Rama<sup>a, b\*</sup>, Isabel Seguro<sup>a</sup>, João G. Pacheco<sup>a</sup>, Henri P.A.

Nouws<sup>a</sup>, M. Natália D.S. Cordeiro<sup>c</sup>, Cristina Delerue-Matos<sup>a</sup>

<sup>a</sup> REQUIMTE/LAQV, Instituto Superior de Engenharia do Porto, Instituto Politécnico do Porto, Dr. António Bernardino de Almeida 431, 4200-072 Porto, Portugal

<sup>b</sup> Departamento de Química Física y Analítica, Universidad de Oviedo, 33006 Oviedo, Spain

<sup>c</sup> REQUIMTE/LAQV, Departamento de Química e Bioquímica, Faculdade de Ciências, Universidade do Porto, Rua do Campo Alegre, s/n, 4619-007 Porto, Portugal

\*e-mail: costaestefania@uniovi.es

3

## 4 Abstract

5 The ever-increasing presence of contaminants in environmental waters is an alarming issue, not only because  
6 of their harmful effects in the environment but also because of their risk to human health. Pharmaceuticals  
7 and pesticides, among other compounds of daily use, such as personal care products or plasticisers, are being  
8 released into water bodies. This release mainly occurs through wastewater since the treatments applied in  
9 many wastewater treatment plants are not able to completely remove these substances. Therefore, the  
10 analysis of these contaminants is essential but this is difficult due to the great variety of contaminating  
11 substances. Facing this analytical challenge, electrochemical sensing based on molecularly imprinted  
12 polymers (MIPs) has become an interesting field for environmental monitoring. Benefiting from their  
13 superior chemical and physical stability, low-cost production, high selectivity and rapid response, MIPs  
14 combined with miniaturized electrochemical transducers offer the possibility to detect target analytes in-  
15 situ. In most reports, the construction of these sensors include nanomaterials to improve their analytical  
16 characteristics, especially their sensitivity. Moreover, these sensors have been successfully applied in real  
17 water samples without the need of laborious pre-treatment steps. This review provides a general overview  
18 of electrochemical MIP-based sensors that have been reported for the detection of pharmaceuticals,  
19 pesticides, heavy metals and other contaminants in water samples in the past decade. Special attention is  
20 given to the construction of the sensors, including different functional monomers, sensing platforms and  
21 materials employed to achieve the best sensitivity. Additionally, several parameters, such as the limit of  
22 detection, the linear concentration range and the type of water samples that were analysed are compiled.

23

24 **Keywords:** molecularly imprinted polymer; electrochemical sensors; emerging contaminants;  
25 pharmaceuticals; pesticides; heavy metals.

26

## 27 1. Introduction

28 Pollution is defined as the direct or indirect introduction of substances or energy into the environment, by  
29 man, that are liable to cause harm to human health, living resources or the biosphere's ecological and  
30 physical systems (Holdgate, 1979; Khanmohammadi et al., 2020; Manisalidis et al., 2020). Although  
31 pollutants can also enter the environment through natural processes, such as volcanic activity, this definition,  
32 contained in EU legislation (Directive, 2008/1/EC), assigns environmental responsibilities to human activities.  
33 Besides the evident detrimental effects (such as loss of biological diversity, introduction of invasive species,  
34 excessive amounts of hazardous chemicals in the food chain, and global climate change), environmental

35 pollution is a major cause of illness that leads to great economic costs for health care systems. The terms  
36 pollution and contamination are sometimes used as synonyms but they are not really the same. A  
37 contaminant is a substance that is not normally present in the environment or that is present in  
38 concentrations above the natural background but is not necessarily harmful. On the other hand, a pollutant  
39 is a contaminant that results in harmful effects to the environment or living organisms (Chapman, 2007).  
40 However, in some cases this distinction is a bit fuzzy because harmful effects may occur but are not observed.  
41 Over the past decades, the rise in population, globalisation, and industrialisation have led to the  
42 dissemination and the increase of the amount and the variety of hazardous substances in the environment.  
43 The harmful effects to living organisms and the ecosystem of many of these substances, which range from  
44 heavy metals and pesticides to phenolic compounds, xenobiotics, plasticisers and antibiotics, have been  
45 demonstrated (Chakraborty et al., 2020; Dulio et al., 2018; Richardson and Ternes, 2014). However, there is  
46 still insufficient knowledge about the toxicity of some of them to environmental, animal and human  
47 populations.

48 Among these substances, the so-called “contaminants of emerging concern” (CECs) or “emerging  
49 contaminants” are receiving huge attention because of their increasing presence and extensive distribution  
50 in the environment (Dulio et al., 2018; Kroon et al., 2020; Naidu et al., 2016; Petrie et al., 2014). CECs, and  
51 their metabolites and/or transformation products, are a heterogeneous group of compounds that are  
52 present in a wide variety of products, including industrial chemicals, pharmaceuticals, drugs of abuse,  
53 personal care products, biocides, food additives, surfactants and plasticisers (Dulio et al., 2018; Lapworth et  
54 al., 2012; Ramírez-Malule et al., 2020). The term CECs does not only cover newly developed substances but  
55 also chemicals that have entered the environment for years, whose presence has only recently been detected  
56 (either because of the increase of their concentration or because of the advances in analytical methods)  
57 and/or studied (Richardson and Ternes, 2014; Zulkifli et al., 2018). Due to insufficient knowledge about the  
58 toxicity, impact, and behaviour of CECs, many of them are not yet regulated, so they are not routinely  
59 monitored and submitted to an emission control regulation. However, since their potential negative effects  
60 are increasingly recognised, relevant regulations are expected over the next years (Bilal et al., 2019). The  
61 variety and amount of CECs is so impressive that the list of priority substances created by NORMAN (the  
62 network of reference laboratories, research centres and related organisations for monitoring of emerging  
63 environmental substances funded by the European Commission) includes more than 900 (Dulio et al., 2018;  
64 norman-network).

65 Although CECs can enter the environment by the same routes as traditional contaminants, such as industrial  
66 processes and emissions, wastewater is recognised as a highly important contributor to the entry of CECs in  
67 the aquatic environment (Lapworth et al., 2012; Luo et al., 2014; Paíga et al., 2016), especially in  
68 countries/regions which do not have suitable regulation regarding wastewater treatment. However, even in  
69 regions with adequate regulation, removal of CECs may be incomplete because this largely depends on the  
70 physical-chemical characteristics of the contaminants and on the treatments applied in the wastewater  
71 treatment plants (WWTPs) (Lapworth et al., 2012; Luo et al., 2014; Patel et al., 2019). Other important  
72 sources of the release of CECs into the aquatic environment are landfill sites, since CECs may leach and reach  
73 groundwater, sewage sludge, and manure (that may contain veterinary pharmaceuticals) used in agriculture  
74 (Lapworth et al., 2012; Sui et al., 2015). Therefore, the determination of CECs levels in the aquatic  
75 environment is of great importance, not only to evaluate the water quality and security, but also to improve  
76 the knowledge on their pathways and fate.

77 As mentioned previously, the increasing amount and variety of contaminants has led to more stringent  
78 environmental regulations and, consequently, to the increase of the demands on environmental analysis.  
79 Vice versa, the latest improvements in environmental analysis results in legislation stringency. The analysis  
80 of contaminants in waters involves great challenges since they are a very large group with very different  
81 physical/chemical properties that are present at very low concentrations (Geissen et al., 2015; Rasheed et  
82 al., 2019). In this context, powerful analytical techniques (e.g. high-performance liquid chromatography and  
83 gas chromatography coupled to mass spectrometric detection (HPLC-MS and GC-MS, respectively)  
84 (Arismendi et al., 2019; Beldean-Galea et al., 2020; Benedetti et al., 2020; Liang et al., 2020; Merlo et al.,  
85 2020; Paíga et al., 2019) are used for the detection of contaminants in waters. However, these techniques  
86 are often not as fast and cost-effective as desired since they require complex and expensive instrumentation,  
87 laborious sample pre-treatment procedures and long analysis times (Rasheed et al., 2019; Zulkifli et al., 2018).

88 Spatial and temporal variations are also important issues in environmental analysis due to seasonality, inter-  
89 and intra-day variations and occasional events that can hamper a harmonised and representative screening  
90 (Luo et al., 2014; Paíga et al., 2019). Therefore, complete information of the presence of contaminants in  
91 water bodies and its spatial and temporal distribution requires the collection of several samples at different  
92 times. When conventional analytical methodologies, such as solid phase extraction followed by  
93 chromatographic analysis (Kuhn et al., 2020), are used, these samples must be sent to a laboratory, turning  
94 real-time information impossible. Hence, there is the need to develop technological solutions for easy, fast  
95 and on-site contamination monitoring that provide a representative spatial-temporal picture of  
96 environmental quality.

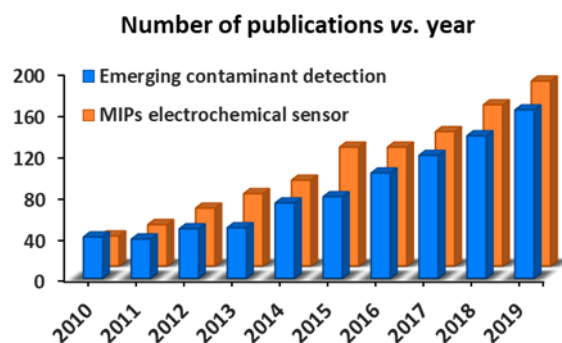
97 The growing demand for inexpensive and easy-to-use analytical devices capable of rapidly providing valuable  
98 qualitative and quantitative on-site information increases the interest in electrochemical sensors because of  
99 their reduced size, portability, low cost and low reagent and sample consumption. Moreover, the  
100 combination of cutting-edge technologies in digital communication networks and innovative sensors allows  
101 the construction of smart analytical tools. A robust electrochemical sensor requires the successful integration  
102 of a transducer (usually an electrochemical cell comprising a working- a reference- and a counter electrode)  
103 with a recognition element (e.g. biological (enzyme, antibody, protein...) or artificial (e.g. molecularly  
104 imprinted polymers (MIPs)). MIPs have attracted wide attention as recognition elements for sensor  
105 development due to their high selectivity towards the target analyte and their advantages compared to  
106 biological receptors: i) easy and low-cost preparation, ii) physical and chemical robustness when  
107 unfavourable conditions are used, such as organic solvents, extreme pH values, high temperatures and/or  
108 high pressures, iii) reusability, iv) stability and v) possibility of large scale production (Ansari and Karimi,  
109 2017a; Figueiredo et al., 2016; Gui et al., 2018).

110 The basis of today's molecular imprinting technology (MIT) started with the studies presented by Wulff and  
111 Sarhan (1972) almost 50 years ago (Beluomini et al., 2019). In the first decades the development and  
112 application of MIPs were mainly focused on separation and extraction techniques. Nowadays, MIPs are used  
113 in a wide variety of applications (Belbruno, 2019; Chen et al., 2016) including sample preparation (e.g. solid  
114 phase extraction) (Ansari and Karimi, 2017b), chromatographic separation (Boysen, 2019; Yang et al., 2016),  
115 drug delivery (Han et al., 2019; Luliński, 2017; Mokhtari and Ghaedi, 2019) and chemical sensing (Cai et al.,  
116 2019; Lopes et al., 2017; Pacheco et al., 2018; Rebelo et al., 2020). Electropolymerisation has facilitated the  
117 application of MIPs in electrochemical sensing by employing electroactive monomers that can polymerize by  
118 applying electric current. The first studies of selective MIPs towards several species produced in this way  
119 were done by Boyle et al. (1989), Dong et al. (1988) and Vinokurov (1992). After this, the first combination  
120 of electropolymerised MIPs and sensors was described by Hutchins and Bachas (1995) for the analysis of  
121 nitrate using a potentiometric sensor. The electrochemical synthesis approach was also successfully used to  
122 develop MIP-sensors by two different groups in 1999 (Deore et al., 1999; Malitesta et al., 1999). Malitesta et  
123 al. (1999) proposed the electropolymerisation of o-phenylenediamine (o-PD) in the presence of glucose  
124 (neutral molecule) in an aqueous environment on a gold-coated quartz crystal to assemble a piezoelectric  
125 sensor. Deore et al. (1999) reported the electrochemical preparation of polypyrrole (PPy) imprinted with L-  
126 glutamate anion. Along with electropolymerisation, the modification of electrodes with MIPs prepared by  
127 traditional methods such as bulk polymerisation was also explored. More recently, the MIP technology has  
128 been coupled to disposable and miniaturized screen-printed electrodes. So, in the future it is foreseen that  
129 some MIPs could be used in commercial sensing solutions.

130 Theoretically, MIPs can be prepared for any molecule of interest; indeed, the MIP database (available at  
131 <http://mipdatabase.com>) compiles MIPs for more than 10,000 target molecules. Therefore, in the last  
132 decade, the development of MIP-based electrochemical sensors has exploded as well as the concern about  
133 emerging contaminants (Fig. 1) (Ahmad et al., 2019; Ansari and Karimi, 2017a; Beluomini et al., 2019; Gui et  
134 al., 2018; Lahcen and Amine, 2019). Combining MIPs and electrochemical sensing strategies has shown great  
135 potential to benefit environmental pollution control (Ayankojo et al., 2020).

136 In this work, a comprehensive review of MIP-based electrochemical sensors for the analysis of contaminants  
137 in water is provided. There are some recent reviews about this kind of sensors, however, they are general  
138 regarding the applications of the sensors (Beluomini et al., 2019; Gui et al., 2018; Lahcen and Amine, 2019),  
139 or they are focused on the preparation and use of imprinted polymers for recognition and extraction, not  
140 only in sensing applications but also in solid phase (micro) extraction (Ansari and Karimi, 2017a; Figueiredo

141 et al., 2016). Thus, this review includes a summary of the main environmental contaminants and provides a  
142 general overview of the MIP-based electrochemical sensors reported for the analysis of these contaminants  
143 in water samples focusing on the polymerisation technique, functional monomers, types of electrodes and  
144 electrode modifiers and comparing their performances. Finally, the present challenges and prospects for  
145 advancing the use of this kind of sensors for environmental analysis are discussed.



146  
147 **Figure 1.** Evolution of publications regarding MIP-based electrochemical sensors and emerging contaminant detection  
148 in the last 10 years.

149

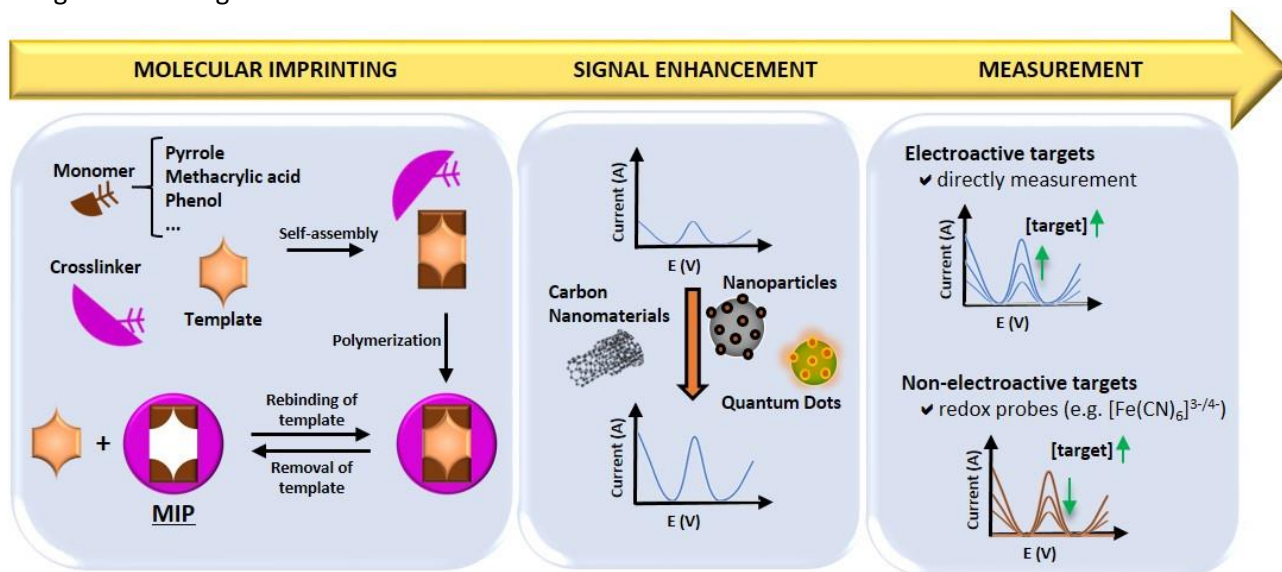
## 150 2. MIPs as recognition elements for electrochemical sensors

151 MIPs are tailor-made synthetic materials with selective binding cavities that act as recognition sites for a  
152 specific target molecule, mimicking natural receptors (Chen et al., 2016). MIPs are synthesized by mixing the  
153 template molecule with a functional monomer in an inert porogenic solvent, resulting in the formation of a  
154 pre-polymerisation complex. This complex polymerises in the presence of a cross-linking agent (that fixes the  
155 monomer around the template and allows the generation of the three-dimensional polymer network) and  
156 an initiator. After completion of the polymerisation process, the template is removed from the polymeric  
157 matrix leaving specific cavities whose shape, size and functional groups are complementary to the template  
158 molecule. Therefore, a molecular memory is generated inside the polymer, in which the target molecule is  
159 now able to rebind with a very high specificity. In this way, the MIP acquires the ability to selectively recognise  
160 the target molecule in the presence of other closely related molecules; a process that is very similar to the  
161 “lock and key” mechanism of enzymes (Chen et al., 2016; Madikizela et al., 2018a). A schematic summary of  
162 preparation of a MIP, signal enhancement strategies and detection mechanisms in electrochemical sensors  
163 is shown in Fig. 2.

164 The recognition properties of a MIP are compared with a non-imprinted polymer (NIP) that is prepared in the  
165 same conditions, but without the inclusion of the template molecule. Obviously, the great challenge is to  
166 obtain a MIP with the highest selectivity for the target molecule (Arrigo and Baroni, 2020).

167 The necessity of quantifying very low contaminant concentrations, the complexity of environmental samples  
168 and the limited availability of sensitive and selective methods are the principal difficulties for environmental  
169 monitoring (Madikizela et al., 2018b; Madikizela et al., 2018c; Namiesnik, 2000; Spietelun et al., 2013). In this  
170 context, suitable sample preparation techniques have been applied to selectively isolate and preconcentrate  
171 the analytes prior to their determination (Sarafray-Yazdi et al., 2012). Many research papers have shown the  
172 use of MIPs as selective sorbents for extraction of various contaminants from complex sample matrices  
173 (Madikizela et al., 2018c). Despite the advantages of this advanced technical application of MIPs, they are  
174 also excellent tools to be used as recognition elements in electrochemical sensing (Li et al., 2019b). As  
175 mentioned before, when compared with biological receptors, such as antibodies or enzymes, MIPs are  
176 cheaper, more robust and have a greater reusability. However, the optimisation of the synthesis process to  
177 obtain a selective MIP is often a laborious and time-consuming task that not always leads to a suitable MIP  
178 as recognition element in a sensor. In this context, to avoid the extent and trial-and-error of experimental  
179 procedures, computational simulations have been employed to describe, predict and analyse molecular

180 imprinting systems (Zhang et al., 2019). With a sustainable strategy, strong guidance regarding on the design  
 181 of MIPs (Khan et al., 2019; Rebelo et al., 2020; Xie et al., 2020).  
 182 Because of the differences in the templates' properties and the potential applications of the MIPs, a great  
 183 variety of strategies for their preparation, such as bulk, suspension, emulsion and precipitation  
 184 polymerisation, and surface imprinting, have extensively been reviewed (Ansari and Karimi, 2017b; Belbruno,  
 185 2019; Chen et al., 2016; Ertürk and Mattiasson, 2017; Lahcen and Amine, 2019; Mayes and Whitcombe,  
 186 2005). In the case of MIP-based electrochemical sensors, there are different approaches for the integration  
 187 of the MIPs with the transducer (Gui et al, 2018, 2019; Toro et al., 2015). The main ones are i) the mixture of  
 188 the MIP with the material of the working electrode before the preparation of the electrode; ii) the  
 189 immobilisation (usually by drop coating) of a previously prepared MIP or MIP-based composite with  
 190 conductive/nanomaterials on the transducer surface; and iii) electropolymerisation. These synthesis and  
 191 integration strategies are reviewed in the next sub-section.



192

193 **Figure 2.** Summary of the preparation procedure, signal enhancement and sensing mechanism of electrochemical  
 194 MIP-based sensors.

## 195 2.1. Preparation processes of MIPs

196 In the classical MIP preparation methods mentioned above, the bond between the template and the  
 197 monomer can either be covalent or non-covalent. Non-covalent binding is more flexible since the removal  
 198 and subsequent rebinding of the template is easier (Zaidi, 2017; Zhou et al., 2019). Some problems that still  
 199 need to be overcome are related to the elution of polar compounds, which have a low solubility in organic  
 200 solvents, the reduced number of effective recognition sites, the slow mass transfer rate and the  
 201 incompatibility of MIPs with aqueous media (Chen et al., 2011). So, alternative imprinting routes or new  
 202 imprinting techniques are constantly being developed.

203 As mentioned in the introduction, theoretically, MIPs can be synthesized for any analyte, but there is no  
 204 specific strategy for a particular class of analytes; i.e. the optimisation of the imprinting process by  
 205 experimental studies is crucial to produce the best MIP (or the MIP with the desired characteristics) for the  
 206 target molecule. Bulk polymerisation is the conventional and most popular approach due to its simple  
 207 operation and low production costs. In this method, also known as mass polymerisation, the template,  
 208 functional monomer, cross-linker and initiator are mixed in a single reactor (Farooq et al., 2018). This process  
 209 results in a rigid monolithic polymer matrix that, after crushing and grinding, may have some shortcomings  
 210 such as the loss of binding sites that can lead to a low yield (Chen et al., 2011; Farooq et al., 2018). Although  
 211 there are different protocols for the preparation of MIPs by bulk polymerisation, this method follows a typical  
 212 sequence involving five main steps: pre-polymerisation of the functional monomers with the template  
 213 molecules, addition of the cross-linking agents and initiators, polymerisation by heating for an extended  
 214 period of time, extraction of the template and crushing the crosslinked mass into powder. Precipitation  
 215 polymerisation, suspension polymerisation and emulsion polymerisation are three spherical MIP synthesis

216 methods. In these methods, the polymerisation procedure is similar to bulk polymerisation, but the post-  
217 treatment steps are not required, which reduces the number of steps and, more importantly, the probability  
218 of destroying the imprinted cavities (Chen et al., 2011; Xiao et al., 2018). Precipitation polymerisation is  
219 carried out in a large volume of an organic solvent in which the polymers are insoluble and precipitate. This  
220 method is free of surfactants and allows to control the size of the particles, but their shape remains irregular.  
221 Suspension and emulsion polymerisation are used to prepare micro spherical MIPs and to increase  
222 monodispersity. For emulsion polymerisation an oil/water biphasic system is prepared and surfactants are  
223 added to the organic phase to prevent diffusion across the continuous aqueous phase (Wackerlig and  
224 Schirhagl, 2016). Compared to the precipitation strategy, suspension and emulsion polymerisation require  
225 more reagents and the purity and performance of the products are affected by the presence of surfactants  
226 remnants (Chen et al., 2016).

227 Another process for the preparation of MIPs is surface imprinting, in which electropolymerisation is  
228 frequently used. In *in-situ* electropolymerisation, using cyclic voltammetry (CV), the imprinted polymers are  
229 produced by coating surfaces at a well-controlled rate, producing an ultrathin polymeric film. In this process,  
230 the CV cycles and time can be used to determine and regulate the adhesion and the morphology of the  
231 polymeric film on the surface of substrates, facilitating the optimisation of the synthesis and providing highly  
232 reproducible MIPs in terms of binding capacities and immobilisation on the substrate's surface. Moreover,  
233 this technique facilitates template removal since the obtained MIP film is thin and all the imprinting cavities  
234 are situated on or closely at the surface (Cieplak and Kutner, 2016). So, electropolymerisation is usually a  
235 simple and fast procedure involving three steps: solubilization and interaction of a functional monomer with  
236 the desired template in a solvent, and electrochemical coating followed by extraction of the template  
237 (Crapnell et al., 2019). Other strategies for surface imprinting have also been studied. For example, the sol-  
238 gel process is an approach that can facilitate MIP preparation in aqueous matrices (Moein et al., 2019; Zhou  
239 et al., 2019). This technique consists of the use of silica-based materials as supports that allow the use of  
240 green solvents and an easy construction at room temperature without the problem of chemical or thermal  
241 decomposition observed in bulk polymerisation (Moein et al., 2019; Pohanka, 2017; Zhou et al., 2019). There  
242 are two main steps for sol-gel processing: sequential hydrolysis of silane in the presence of an acidic or basic  
243 catalyst and condensation of a series of silane monomers to form siloxane bonds. With time, a porous and a  
244 strong three-dimensional network is formed by colloidal particles aggregation (Mujahid et al., 2010; Yang et  
245 al., 2013). Sol-gel materials offer high thermal stability and porosity, which combined with molecular  
246 imprinting systems play a significant role in the performance of MIPs. The lack of functional monomers, the  
247 low sensitivity, slow diffusion kinetics and long response times are some drawbacks of this process  
248 (Adumitrachioaie et al., 2018; Zhou et al., 2019).

249 Many materials for surface modification are also used in surface molecular imprinting processes such as  
250 Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles (Fe<sub>3</sub>O<sub>4</sub> MNPs) and mesoporous SiO<sub>2</sub> nanoparticles (SiO<sub>2</sub> NPs) (Fig. 2). Besides  
251 a larger specific surface area, magnetic nanoparticles have superior advantages in the removal of solvents  
252 and in the fast and easy isolation of the analyte from samples by applying a magnetic field (Huang et al.,  
253 2018; Ye, 2013). SiO<sub>2</sub> NPs provide a stable solid support due to their mesoporous properties, allowing higher  
254 stability and sorption capacities in comparison with ordinary imprinting methods (Wang et al., 2011). By  
255 virtue of the controllable methods and ease of modification, surface imprinting technologies have been  
256 explored by several research groups. Furthermore, they have also included functional nanomaterials in the  
257 imprinted shell layers to improve the MIP-based sensors' electron transfer rates (Beluomini et al., 2019;  
258 Maduraiveeran et al., 2018). Carbon nanotubes (CNTs), graphene and quantum dots (QDs) have received  
259 special attention in the preparation of MIPs. Among the carbon materials, graphene is the most widely used  
260 because of its unique physicochemical properties. Besides its low electronic noise and thin 2-dimensional  
261 surface, graphene has strong interaction with carbon-based ring compounds, such as some contaminants  
262 (Zaidi, 2017). QDs are a kind of conducting nanocrystals that have excellent photoluminescence properties  
263 (Shi et al., 2019). QDs possess high luminescence efficiency and photostability, a broad absorption spectrum,  
264 narrow fluorescence emission bands and quantum-size effects (Nsibande and Forbes, 2019). The  
265 combination of their excellent properties with the specific recognition of MIPs is gaining more attention as  
266 an alternative for sensitive and selective detection of contaminants (Nsibande and Forbes, 2019; Sobiech et  
267 al., 2019; Wang et al., 2019; Zhou et al., 2018). Prior to molecular imprinting, QDs can be loaded on an

268 electrode surface or incorporated in the surface of supporting nanomaterials such as SiO<sub>2</sub> NPs (Tan et al.,  
269 2016; Yola and Atar, 2017).

## 270 **2.2 Functional monomers**

271 The selection of an appropriate functional monomer is an essential step for the successful construction of  
272 MIPs with desired affinity and selectivity. Template-monomer interactions can be changed by the  
273 combination of different functional groups. So, many types of commercially available functional monomers  
274 have been studied, based on their potential to establish hydrogen bonds or electrostatic interactions with  
275 the template. Among the functional monomers reported in the literature, methacrylic acid (MAA), pyrrole  
276 (Py), and phenol (Ph) and derivatives are the most widely used in electrochemical MIP-based sensors for the  
277 detection of contaminants. Due its nature, MAA is the most popular carboxylic acid-based monomer used in  
278 bulk polymerisation. MAA not has only the ability to serve as both hydrogen bond donor or acceptor, but it  
279 can also interact with the template in various ways, including strong ion-pair and dipole-dipole interactions,  
280 and van der Waals forces, increasing its broad applicability (Mayes and Whitcombe, 2005). Py is a conductive  
281 (to some extent) and electropolymerisable monomer, well known to partially cross-link (Sadriu et al., 2020).  
282 Without the need to add a cross-linker, electropolymerisation is the most efficient method to synthesize PPy  
283 MIPs. The formation of the PPy film is fast, and a chemically and mechanically stable polymer is obtained  
284 (Sadriu et al., 2020). As a result of its good permeability, the use of PPy also allows easier extraction and  
285 rebinding of the template. Nevertheless, there are several experimental variables that can affect the  
286 chemical and physical properties of PPy formed on the electrode's surface, such as solvent, electrolyte,  
287 temperature and pH (Sadki et al., 2000). In contrast, Ph and p-aminothiophenol (PATP) are electronically non-  
288 conducting polymers which are also extensively used in electropolymerisation. The insulation properties of  
289 the formed polymer films can be improved by using self-assembled monolayers (Sharma et al., 2012);  
290 particularly PATP, which contains thiol groups, has been used for strong binding on gold surfaces. All these  
291 functional monomers can establish strong hydrogen bonds with templates, demonstrating that this effective  
292 interaction is a promising option to take into account during MIP preparations (Ansari, 2017).

## 293 **2.3 Detection mechanisms**

294 When using electrochemical MIP-based sensors, electrical signal (current or potential) changes, which are  
295 proportional to the concentration of the target analyte, are recorded (Li et al., 2012). Two principal  
296 techniques are used to detect the specific recognition events: voltammetry and electrochemical impedance  
297 spectroscopy (EIS) (Li et al, 2019b).

298 The voltammetric methods include linear sweep voltammetry (LSV), CV, differential pulse voltammetry (DPV)  
299 and square wave voltammetry (SWV). In these methods, the electroactivity of the analyte conditions the  
300 detection mechanism. For electroactive molecules, the current is measured directly, where the signal outputs  
301 ideally are the faradaic currents resulting from the oxidation/reduction of the analytes after binding in the  
302 MIP. In this case, under controlled conditions a correlation can be made between the concentration of the  
303 analyte and the measured current (Chen et al., 2016). When non-electroactive targets are involved, the signal  
304 can be produced using redox probes such as [Fe (CN)<sub>6</sub>]<sup>3-/4-</sup> (Li et al., 2012). In this case, also called the "gate-  
305 controlled" mechanism, the current response of the redox probe is inversely proportional to the  
306 concentration of the analyte since higher concentrations lead to fewer channels for the diffusion of the redox  
307 probe to the electrode surface, and consequently to the decrease of the signal (Gui et al., 2019) (Fig. 2).

308 EIS is a powerful tool to investigate electron transfer and diffusion processes that occur at the  
309 electrode/electrolyte interface. The typical Nyquist plot obtained in EIS has two areas: at high frequencies a  
310 semi-circle portion corresponds to the electron transfer resistance, and at low frequencies diffusion limited  
311 process are observed by a linear portion (Li et al., 2018). During the MIP's assembly process, changes in the  
312 surface properties of the electrode occur and different Nyquist plots are observed. Generally, the rebinding  
313 of the template in the cavities of the polymeric film results in an increase of the charge transfer resistance  
314 on the surface of the electrode, thus semi-circles with larger diameters are observed.

315

### 316 3. MIP-based electrochemical sensors for contaminant analysis

#### 317 3.1 Pharmaceuticals

318 Pharmaceuticals are widespread micropollutants and are ubiquitous in waters and soils (Afonso-Olivares et al., 2017; Kamba et al., 2017). They have been released into the environment for decades through various  
319 ways, such as metabolic excretion in its original form or as metabolites, improper disposal of expired  
320 medication, and escaping the modest removal efficiencies of WWTPs (Björnlén et al., 2018; Rivera-Jaimes  
321 et al., 2018). Despite their low concentrations, their long half-lives amplifies effects from drug-drug  
322 interactions that can potentially be hazardous (Evgenidou et al., 2015; Kamba et al., 2017). Subtle effects of  
323 pharmaceutical compounds on aquatic and terrestrial organisms have been reported (Boxall, 2004) and,  
324 more recently, a major concern of the public and the scientific community includes bacterial resistance  
325 (Ayukekbong et al., 2017; Richardson and Ternes, 2014).

327 There is no definitive date that marks the emergence of the identification of pharmaceuticals in the  
328 environment (Daughton, 2016), but over the past three decades various methods and approaches have been  
329 developed to assess their profiles and occurrence patterns in different environmental compartments  
330 (Evgenidou et al., 2015; Yang et al., 2017). Undoubtedly, the knowledge of concentration levels of  
331 pharmaceuticals is essential as a starting point to apply more advanced treatments to improve their removal  
332 and, consequently, minimise their environmental risk (Afonso-Olivares et al., 2017).

333 Since 2009 several works have been dedicated to the development of electrochemical MIP-based sensors for  
334 the determination of a wide variety of pharmaceuticals including antibiotics, anti-inflammatories, analgesics,  
335 hormones, blood lipid regulators,  $\beta$ -blockers, antidepressants, antiepileptics and cytostatic drugs (Des  
336 Azevedo et al., 2013; Florea et al., 2015; Futra et al., 2016; Han et al., 2016; Miege et al., 2009; Özcan and  
337 Topçuoğulları, 2017; Xiao et al., 2017; Yang et al., 2017). Many different strategies based on new imprinting  
338 techniques and polymerisation methods, and the use of innovative nanomaterials have been studied to  
339 improve the performance of the sensors (Adumitrachioaie et al., 2018; Uzun and Turner, 2016). However,  
340 only a few electrochemical MIP-based sensors were applied to environmental samples. This is mainly because  
341 of the low levels of contaminants in the environment. Table 1 summarises the main characteristics of  
342 electrochemical MIP-based sensors for the detection of different classes of pharmaceuticals in water  
343 samples.

#### 344 3.1.1 Antibiotics

345 Antibiotics are the most frequently studied group of pharmaceuticals for electrochemical detection using  
346 MIP-based sensors, showing the concern about their presence in waters and the importance of their  
347 detection.

348 Sulfamethoxazole (SMX) is one of the most frequently sulfonamide bacteriostatic antibiotic detected in the  
349 aquatic environment, soils and sediments due to its wide spectrum of applications and poor ability to be  
350 metabolised (Fekadu et al., 2019; Tao et al., 2019; Zhao et al., 2015). SMX has been detected in industrial  
351 effluents, in WWTP effluents and in surface water at concentrations between 0.036 nM and 7.2 nM with  
352 detection frequencies of more than 80% (García-Espinoza and Mijaylova Nacheva, 2019). In Africa, it was the  
353 most detected compound in water samples, reaching concentrations far higher than the ecotoxicity  
354 endpoints (Fekadu et al., 2019). Its presence revealed high risk to sensitive aquatic organisms (Zhang et al.,  
355 2012) with possible mutagenic activity and the potential to disrupt the endocrine system (Archer et al., 2017;  
356 Zhao et al., 2015). Therefore, its monitoring is relevant to guarantee the health of all organisms. In 2015, the  
357 first electrochemical MIP-sensor, based on a PPy modified Boron Doped Diamond Electrode (BDDE), was  
358 developed for the determination of SMX in spiked lake water (Zhao et al., 2015). This sensor was prepared  
359 by electropolymerisation of Py and the determination of SMX was achieved through its direct oxidation using  
360 SWV. A good sensitivity, a limit of detection (LOD) of 24 nM and minimal interferences of structurally similar  
361 sulfonamides (sulfadimethoxine, sulfadiazine and sulfafurazole) were observed. Although BDDEs have  
362 excellent properties, some researchers avoid their use because of the very heterogeneous composition of  
363 the BDDE's surface (Feier et al., 2019) and because of the possibility of its degradation in alkaline solutions  
364 (Luong et al., 2009). Sulfanilamide (SN) is another member of the sulfonamide family and has mainly been



365 detected in surface waters, with high detection rates in Chinese rivers in comparison with other countries.  
366 Tadi et al. (2014) also used electropolymerisation of Py for the fabrication of a sensor for the analysis of SN.  
367 In this case, a pencil graphite electrode (PGE) was used. Py was selected as the best functional monomer  
368 compared with furan, thiophene, methylthiophene and methylpyrrole since computational studies showed it  
369 has the highest binding interaction energy with SN. DPV was used for its direct analysis (oxidation), achieving  
370 an LOD of 20 nM. This sensor showed good selectivity towards SN; species with analogue structures such as  
371 SMX, sulfathiazole and sulfadiazine did not significantly interfere in the analysis. The applicability of this  
372 sensor was tested in spiked human serum and ground water samples, obtaining recoveries between 98 and  
373 115%.

374 The use of nanomaterials is one of the most widely used strategies to increase the electrode's surface area  
375 and consequently the sensitivity of the method. Zamora-Gálvez et al. (2016) reported a specific and highly  
376 sensitive composite-based sensing system, based on MIP-modified Fe<sub>3</sub>O<sub>4</sub> MNPs, to detect SMX via EIS (Fig.  
377 3A). In this work, nano-sized MIP cavities were constructed on the surface of the Fe<sub>3</sub>O<sub>4</sub> nanoparticles by bulk  
378 polymerisation using MAA as the functional monomer. Because of the superparamagnetic properties of the  
379 formed composite, this sensing system allowed easy separation, pre-concentration and manipulation of the  
380 target analyte. After magnetic deposition of the composite on the surface of a screen printed carbon  
381 electrode (SPCE), the electron transfer resistance (R<sub>ct</sub>) was monitored by EIS in the presence of [Fe(CN)<sub>6</sub>]<sup>3-</sup>/  
382 <sup>4-</sup>, which increased with increasing SMX concentrations. This sensing system showed an excellent LOD of 1  
383 × 10<sup>-3</sup> nM and was applied to spiked seawater samples, achieving recoveries between 87 and 106%. In  
384 comparison with the previously described SMX sensor (Zhao et al., 2015), and although higher amounts of  
385 reagents were required, a much lower LOD was obtained. In this work MIP-modified Fe<sub>3</sub>O<sub>4</sub> MNPs were used  
386 to separate and pre-concentrate SMX and their magnetic deposition onto the surface of SPCEs offered an  
387 innovative design in the field of disposable sensors.

388 Azithromycin (AZY) and erythromycin (ERY) are frequently prescribed macrolide antibiotics to treat many  
389 different bacterial infections. Due to their wide use, the difficulty of removing them by common wastewater  
390 treatments and their frequent detection in water bodies, AZY and ERY were included in the EU Watch List  
391 (Decision, 2018/840/ EU) of substances that could pose a significant risk to aquatic environments (Ayankojo  
392 et al., 2020; Rebelo et al., 2020). Recently, a low-cost and user-friendly electrochemical MIP sensor to detect  
393 AZY in water was reported (Rebelo et al., 2020). By a computational study based on density functional theory  
394 (DFT), 4-aminobenzoic acid (4-ABA) was chosen as the most suitable monomer, which was  
395 electropolymerised on the surface of a SPCE. The analysis of AZY was performed by its oxidation using DPV  
396 (LOD = 80 nM) and the sensor displayed great recognition behaviour in the presence of other interfering  
397 compounds. It was successfully applied to the analysis of spiked tap water and water samples collected  
398 upstream of a WWTP output in the Ave river (Portugal). A similar system was used by Ayankojo et al. (2020)  
399 to obtain the first ERY-selective MIP film integrated with a screen printed gold electrode (SPAuE). In their  
400 work, the MIP was generated directly on the SPAuE via electropolymerisation of m-phenylenediamine (m-  
401 PD). DPV measurements performed using a [Fe(CN)<sub>6</sub>]<sup>3-</sup>/<sup>4-</sup> redox probe, allowed the determination ERY with  
402 an LOD of 0.1 nM.

403 Chloramphenicol (CAP), one of the oldest antibiotics, is widely used in veterinary medicine and to promote  
404 the growth of food-producing animals. This antibiotic was completely banned for use in food-producing  
405 animals in many European countries due to its potential serious adverse effects (Zhao et al., 2012). Despite  
406 legal bans, CAP is easily available and is still used because of economic interests (Ding et al., 2017; Shaheen  
407 et al., 2017). Zhao et al. (2012) reported the first electrochemical sensor for the determination of CAP in  
408 natural water samples, combining MIPs and the advantages of carbon nanotubes (CNTs) and gold  
409 nanoparticles (AuNPs). The functional monomer, diethylaminoethyl methacrylate (DMA), was added  
410 together with CAP to perform bulk polymerisation and the direct detection process was based on DPV using  
411 a glassy carbon electrode (GCE). The LOD of this sensor was 74 nM and the selectivity for CAP detection was  
412 demonstrated in the presence of penicillin-G, thiamphenicol and their analogue p-nitrophenol. Bulk  
413 polymerisation was also used to construct a novel sensor modified with chitosan and AuNPs (Ch-AuNPs) to  
414 detect ciprofloxacin (CIP), an antibiotic that has been detected in surface waters. Ch-AuNPs were used as the  
415 supporting material to polymerize MAA in the presence of CIP (Surya et al., 2020). The resulting MIP was  
416 drop casted onto the surface of a GCE and its capability to quantify the antibiotic in real samples was tested  
417 using mineral and tap water. Another antibiotic of the quinolone family that has also been detected in surface

418 waters is lomefloxacin (LFX). Li et al. (2020a) developed an effective MIP sensor for its determination in river  
 419 waters. They used a gold electrode modified with Fe-doped porous carbon (Fe-PC) and electropolymerisation  
 420 to produce the MIP film. Because of the hydrogen bonds and host-guest inclusion interactions, o-  
 421 phenylenediamine (o-PD) and  $\beta$ -cyclodextrin ( $\beta$ -CD) were chosen as functional monomers. This methodology  
 422 allowed high selectivity towards LFX, because of the use of the MIP prepared with bifunctional monomers,  
 423 and high sensitivity, which was attributed to the large specific surface area, rich porosity, and good catalytic  
 424 property of Fe-PC. An indirect detection approach and DPV were used for the analysis of LFX detection in  
 425 river waters, achieving an LOD of 0.2 nM.

426 **Table 1.** Electrochemical MIP sensors constructed on different sensing platforms for the detection of  
 427 pharmaceuticals in water samples.

Target	Functional monomer	Polymerisation method / Transducer	Electrochemical technique	Samples	Linear Range (nM)	LOD (nM)	Reference
Sulfamethoxazole	Py	Electropolymerisation, BDDE	SWV	Lake water	$1.0 \times 10^2 - 1.0 \times 10^5$	24 <sup>a</sup>	(Zhao <i>et al.</i> , 2015)
	MAA	Bulk polymerisation, SPCE, Fe <sub>3</sub> O <sub>4</sub> MNPs	EIS	Seawater	$0.1 - 1.0 \times 10^7$	$1.0 \times 10^{-3b}$	(Zamora-Gálvez <i>et al.</i> , 2016)
Sulfanilamide	Py	Electropolymerisation, PGE	DPV	Ground water	$50 - 1.1 \times 10^3$ $1.1 \times 10^3 - 4.8 \times 10^4$	20 <sup>c</sup>	(Tadi <i>et al.</i> , 2014)
Azithromycin	4-ABA	Electropolymerisation, SPCE	DPV	River water and tap water	$5.0 \times 10^2 - 1.0 \times 10^4$	80 <sup>d</sup>	(Rebelo <i>et al.</i> , 2020)
Erythromycin	m-PD	Electropolymerisation, SPAuE	DPV	Tap water	2 – 16	0.1 <sup>b</sup>	(Ayankojo <i>et al.</i> , 2020)
Chloramphenicol	DAM	Bulk polymerisation, MWCNTs, AuNPs, GCE	DPV	Seawater and reservoir water	$0.31 \times 10^3 - 3.1 \times 10^5$	74 <sup>a</sup>	(Zhao <i>et al.</i> , 2012)
Ciprofloxacin	MAA	Bulk polymerisation, Ch-AuNPs, GCE	DPV	Mineral and tap water	$1.0 \times 10^3 - 1.0 \times 10^5$	$2.1 \times 10^{2e}$	(Surya <i>et al.</i> , 2020)
Lomefloxacin	o-PD and $\beta$ -CD	Electropolymerisation, Au electrode, Fe-PC	DPV	River and lake water	$1 - 1.2 \times 10^2$	0.2 <sup>a</sup>	(Li <i>et al.</i> , 2020b)
Chlortetracycline	o-PD	Electropolymerisation, GO, GCE	DPV	Tap water and laboratory wastewater	$1.0 \times 10^4 - 5.0 \times 10^5$	Not mentioned	(Liu <i>et al.</i> , 2013)
Cefalexin	I3AA	Electropolymerisation, GCE, BDDE	DPV	River water	$10 - 1.0 \times 10^3$	3.2 and 4.9 <sup>b</sup>	(Feier <i>et al.</i> , 2019)
Mebendazole	MAA	Electropolymerisation, Fe-NCNF, GCE	DPV	Tap water and river water	$10 - 1.5 \times 10^3$	4 <sup>a</sup>	(Rao <i>et al.</i> , 2018)
17- $\beta$ -estradiol	MNA	Electropolymerisation, PtNPs, GCE	DPV	Hospital wastewater and tap water	$30 - 5.0 \times 10^4$	16 <sup>a</sup>	(Yuan <i>et al.</i> , 2011)
	PATP	Electropolymerisation, AuNPs, Au electrode	LSV	River water	$3.6 \times 10^{-6} - 3.6$	Not mentioned	(Florea <i>et al.</i> , 2015)
	An	Bulk polymerisation, Fe <sub>3</sub> O <sub>4</sub> MNPs, SPCE	SWV	River water	$50 - 1.0 \times 10^4$	20 <sup>a</sup>	(Lahcen <i>et al.</i> , 2017)

428 <sup>a</sup> LOD calculated as S/N = 3.

429 <sup>b</sup> LOD calculated as 3 times the standard deviation of the blank divided by the slope.

430 <sup>c</sup> LOD calculated as 3.3 times the standard deviation of the blank divided by the sensitivity.

431 <sup>d</sup> LOD calculated as 3 times the standard deviation of the intercept divided by the slope.

432 <sup>e</sup> LOD formula no specified.

433 4-ABA: 4-aminobenzoic acid;  $\beta$ -CD:  $\beta$ -cyclodextrin; An: aniline; AuNPs: gold nanoparticles; BDDE: boron doped diamond; Ch-AuNPs:  
434 chitosan gold nanoparticles; DAM: diethylaminoethyl methacrylate; DPV: differential pulse voltammetry; EIS: electrochemical  
435 impedance spectroscopic; Fe-NCNF: nitrogen-doped carbon nanosheet frameworks decorated with Fe; Fe-PC: Fe doped porous  
436 carbon; GCE: glassy carbon electrode; GO: graphene oxide; I3AA: indole-3-acetic acid; LSV: linear sweep voltammetry; m-PD: m-  
437 phenylenediamine; MAA: methacrylic acid; MNA: 6-mercaptopuronic acid; MNPs: magnetic nanoparticles; MWCNTs: multiwalled  
438 carbon nanotubes; o-PD: o-phenylenediamine; PATP: p-aminothiophenol; PGE: pencil graphite electrode; PtNPs: platinum  
439 nanoparticles; Py: pyrrole; SPAuE: screen printed gold electrode; SPCE: screen printed carbon electrode; SWV: square wave  
440 voltammetry.

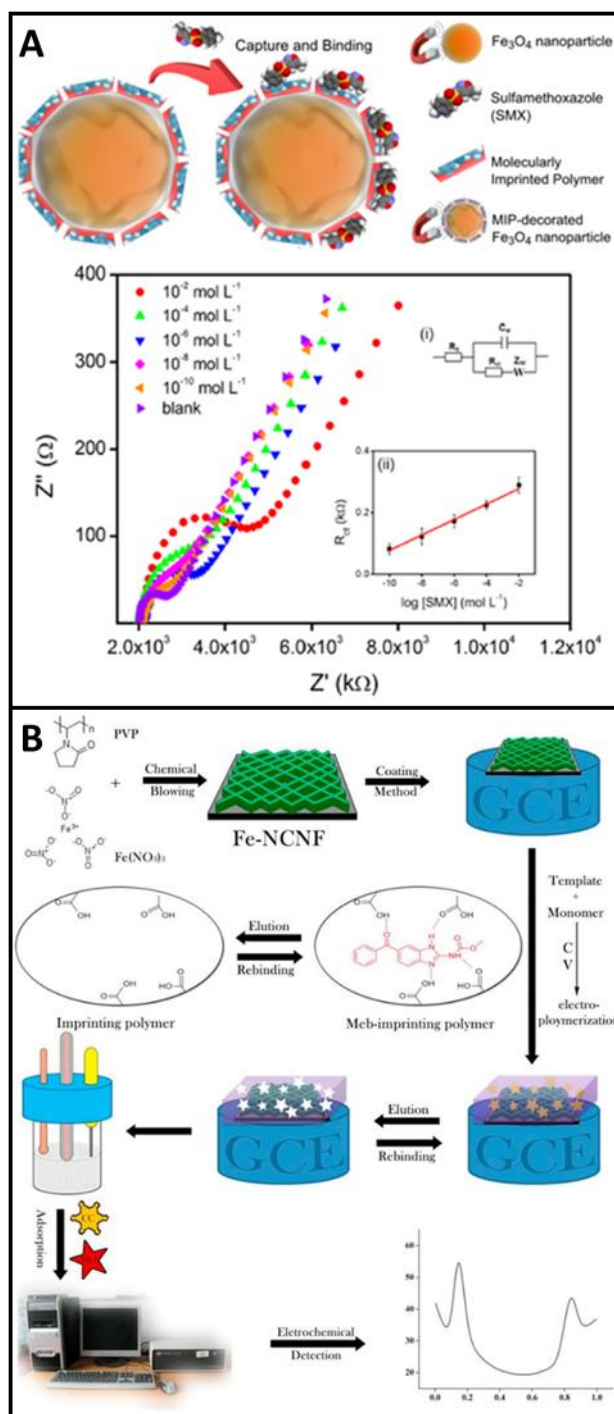
441

442 Tetracycline antibiotics are highly prescribed worldwide because of their favourable properties both for  
443 human and animal therapy (Borghi and Palma, 2014; Li et al., 2020b; Van et al., 2020). More than 70% of  
444 these antibiotics are released in their active forms into the environment (Daghrir and Drogui, 2013; Javid et  
445 al., 2016) and, due its presence in waters, several studies have shown direct effects on microbial community  
446 structures (Grenni et al., 2018). Liu et al. (2013) developed a sensor to detect chlortetracycline (CTC), a  
447 derivate of tetracycline, based on electropolymerisation of o-PD and controlled electrochemical reduction of  
448 graphene oxide (GO) on a GCE. DPV measurements evaluated the electrochemical performance of the MIP-  
449 sensor for the analysis of tap water and a spiked laboratory wastewater sample. Although a current  
450 amplification effect and a higher electroconductivity through the electrochemical reduction of GO was  
451 expected, the sensor did not show a high sensitivity and the authors acknowledge the need to expand the  
452 linear concentration range ( $1.0 \times 10^4 - 5.0 \times 10^5$  nM).

453 Cefalexin (CFX) is a broad-spectrum antibiotic that belongs to the class of cephalosporins (Rahim et al., 2019).  
454 Cephalosporins have been found in different aqueous matrices and river sediments in which cefalexin is the  
455 most frequently detected compound (Ribeiro et al., 2018). Its ubiquitous presence in sewage and wastewater  
456 is correlated with human and livestock populations. An electrochemical MIP sensor for its determination was  
457 recently described by Feier et al. (2019). Two different electrodes, GCE and BDDE, were used to assess the  
458 performances of the MIP sensors. Indole-3-acetic acid (I3AA) was chosen as the functional monomer, not  
459 only because it can be electropolymerised in aqueous solution, but also because of its functional groups that  
460 can form strong interactions with CFX. The indirect quantification of CFX was carried out by using  $[\text{Fe}(\text{CN})_6]^{3-}$   
461  $^{4-}$  and DPV. The LOD obtained with the BDDE was lower, but its precision was worse compared to the GCE.  
462 As mentioned above for the MIP sensor proposed for SMX (Zhao et al., 2015), BDDE provided very attractive  
463 advantages which was confirmed in this work: a more pronounced surface imprinting process and,  
464 consequently, a better sensitivity was achieved. However, GCE offered a better reproducibility and showed  
465 to be the most reliable transducer. This MIP sensor was easy to prepare and was successfully applied to river  
466 water analysis.

### 467 3.1.2 Anthelmintic

468 Unlike the variety of MIP-sensors for antibiotic analysis, to the best of our acknowledge only one MIP-based  
469 sensor for the detection of anthelmintic pharmaceuticals has been reported. This sensor combined hybrid  
470 nanomaterials with a MIP prepared by electropolymerisation for the simultaneous determination of  
471 mebendazole (MB) and catechol (Rao et al., 2018). A carbon nanosheet was created through a chemical  
472 blowing process on a GCE (Fig. 3B) and after electrochemical polymerisation of MAA with MB, it was observed  
473 that, due to their thin shells, nitrogen-doped carbon nanosheet frameworks decorated with Fe (Fe-NCNF)  
474 played a significant role in absorbing more MB molecules. Consequently, its frame structure with a large  
475 specific surface area allowed a good electron-transfer for the oxidation of MB at 0.82 V, but also supported  
476 the strong adsorption of catechol, which was electrochemically oxidised at 0.15 V. An easy-to-prepare and  
477 efficient MB sensor was obtained in this work. Fe-NCNF was synthesized using a simple process and its  
478 combination with a MIP, synthesized through electropolymerisation, amplified the electrochemical response  
479 of the sensor.



480

481 **Figure 3.** A) Schematic representation of the MIP-based sensor for SMX detection based on MIP-modified  $\text{Fe}_3\text{O}_4$   
 482 MNPs and the Nyquist plots and calibration curve for the determination of SMX. Reproduced and adapted with  
 483 permission from Zamora-Gálvez et al. (2016). Copyright 2016 American Chemical Society. B) Preparation and analysis  
 484 procedures of the Fe-NCNF/MIP/GCE for detection of MB. Reproduced from Rao et al. (2018) with permission from  
 485 Elsevier.

### 486 3.1.3 Hormones

487 Other substances present in environmental waters at concentrations of toxicological and carcinogenic  
 488 concern are hormones, especially 17-  $\beta$ -estradiol (E2). E2 is commonly used in contraceptive pills and its  
 489 release in water has gained notable attention, mainly because it is considered the major contributor to  
 490 endocrine disruption of many species in the ecosystem (Salste et al., 2007). Additionally, it has been linked  
 491 with breast cancer in women and prostate cancer in man (Adeel et al., 2017). E2 was also added to the EU  
 492 Watch list of emerging aquatic contaminants (Decision, 2018/840/EU), highlighting the importance of

493 developing sensitive and accurate methods for its determination. In 2011, Yuan et al. (2011) reported an  
494 electrochemical MIP-based sensor for E2 quantification using a GCE modified with platinum nanoparticles  
495 (PtNPs). The MIP for E2 was constructed through the electropolymerisation of 6-mercaptopuric acid (MNA)  
496 and DPV was applied to directly detect the E2 binding to the MIP since it is an electroactive species. This  
497 sensor reached an LOD of 16 nM and was successfully applied to spiked hospital wastewater and tap water  
498 analysis. Later, Florea et al. (2015) developed another MIP sensor able to detect E2 via electropolymerisation  
499 of PATP. Assembled AuNPs on the surface of an Au electrode were used as electron wire for signal  
500 amplification, but in this work the response of the MIP sensor to E2 was measured indirectly by LSV. The  
501 combination of Fe<sub>3</sub>O<sub>4</sub> with molecularly imprinting again showed to be an excellent tool for the highly specific  
502 detection of E2 (Lahcen et al., 2017). Lahcen et al. (2017) developed a Fe<sub>3</sub>O<sub>4</sub>-MIP sensing system with high  
503 selectivity towards E2 for the analysis of river water samples. Following the traditional bulk polymerisation  
504 strategy, MIP-modified Fe<sub>3</sub>O<sub>4</sub> MNPs were obtained and then used to modify the working electrode surface  
505 of SPCE by drop-casting, leading to the enhancement of the direct oxidation current obtained by SWV. The  
506 developed sensor achieved an LOD of 20 nM.

## 507 **3.2. Pesticides**

508 Over the past 50 years, global sales of pesticides have increased because their importance in agricultural  
509 production. They inhibit and prevent the growth of harmful animals, insects, invasive plants, weeds, and  
510 fungi. However, repeated applications result in their accumulation in soils and they can be transported to the  
511 aquatic environment by surface runoff (Rousis et al., 2017). Considering their chemical properties and  
512 persistence, the biodiversity and the ecosystems' health is jeopardised since their toxic action is not restricted  
513 to target pests (Carvalho, 2017). Among all available pesticides, herbicides, insecticides, and fungicides are  
514 the most used types. Although electrochemical detection of pharmaceuticals in water samples using MIPs  
515 appears an unexplored area, many publications have been reported about their use for the trace analysis of  
516 pesticides, which are summarised in Table 2.

### 517 **3.2.1 Insecticides**

518 Organophosphates persist for days/weeks in the aquatic environment and studies showed that they are  
519 accumulated by crustaceans and fishes (Carvalho, 2017). These are the insecticides for which most  
520 electrochemical MIP sensors have been developed. As shown in Table 2 different strategies were adopted  
521 for the detection of chlorpyrifos (CPF) and the use of GCE is common to all of them. Xie et al. (2010b) reported  
522 a molecular self-assembly strategy for electropolymerisation of PATP on the surface of AuNPs-modified GCE  
523 by the formation of Au-S bonds. A linear range between  $5.0 \times 10^2$  nM and  $1.0 \times 10^4$  nM and an LOD of 330  
524 nM were obtained. This sensor showed good interclass selectivity and the authors tested its applicability in  
525 spiked tap water. A better LOD (4.08 nM), was attained via simple bulk polymerisation of MAA (Xu et al.,  
526 2017). A GCE was coated with a suspension of the resulting MIP and the binding of CPF was evaluated by DPV  
527 using [Fe(CN)<sub>6</sub>]<sup>3-/4-</sup> as redox probe. This simple sensor provided a good selectivity and its applicability was  
528 assessed in river water samples. However, an electrochemical MIP-sensor based on carbon nitride nanotubes  
529 (C<sub>3</sub>N<sub>4</sub> NTs) decorated with graphene quantum dots (GQDs) (Yola and Atar, 2017) provided a much lower LOD  
530 ( $2.0 \times 10^{-3}$  nM) in comparison with the previous works. As mentioned above, these nanomaterials have  
531 attracted great attention, and this work is one of the first reports that combines MIP with their excellent  
532 properties for application in wastewater samples. After the synthesis of the C<sub>3</sub>N<sub>4</sub> NTs@GQDs composite by  
533 hydrothermal treatment, a suspension of this nanohybrid material was dropped onto the GCE surface. The  
534 electropolymerisation was performed using Py as functional monomer creating electrostatic interactions and  
535 hydrogen bonds with CPF. The performance of the prepared MIP sensor was evaluated directly by SWV. High  
536 conductivity of the electrode surface was achieved, and it was demonstrated that the introduction of a finite  
537 bandgap into graphene improved its gapless nature, showing the potential role of GQDs in the development  
538 of electrochemical sensors.

539 In another example of the use of nanomaterials, electrochemical MIP sensors for the analysis of methyl-  
540 parathion (MP) were prepared using both electro- and bulk polymerisation. Wu et al. (2014) constructed a  
541 sensor based on AuNPs decorated with CNTs. After electrodeposition of functionalised AuNPs on a

542 MWCNTs/GCE surface, the electrode was immersed in a solution containing PATP. Since MP is electroactive,  
543 its recognition by the MIP could be directly monitored by LSV. The proposed sensor allowed a low LOD (0.30  
544 nM) and was successfully applied to the determination of MP in spiked distilled and tap water, and apple and  
545 cucumber samples with recoveries ranging from 95 to 106%.

546 Another quantitative method for MP was based on nitrogen-doped graphene sheets (N-GS) and a MIP  
547 synthesized by electropolymerisation of Ph (Xue et al., 2014). Although N-GS has been described as an  
548 excellent sensing material for molecular imprinting, the developed MIP-sensor showed a worse analytical  
549 performance than the one described above (Wu et al., 2014) (see Table 2).

550 The use of carbon paste-based electrodes (CPE) has some benefits that include easy surface modification and  
551 very low ohmic resistance (Toro et al., 2015; Vytras et al., 2009). These advantages were used to construct  
552 MIP sensors for parathion (Alizadeh, 2009) and diazinon (Motaharian et al., 2016), where the MIPs were  
553 synthesized by bulk polymerisation using MAA as functional monomer. The detection of these  
554 organophosphorus pesticides was performed by SWV and the authors demonstrated that the optimisation  
555 of the composition of the CPE is very important to improve the sensor's sensitivity.

556 The introduction of nanomaterials into CPEs seems to be crucial to achieve the best performance. In the past  
557 few years the rapid growth of research interest in metal organic frameworks (MOFs) linked via the self-  
558 assembly of transition metal ions/clusters and organic ligands has been observed (Xu et al., 2020b). Very  
559 recently, Xu et al. (2020b) based on its favourable characteristics like highly ordered structure and exposed  
560 sites, prepared a novel disposable carbon paste microelectrode (CPME) MIP sensor on zirconium (Zr) based  
561 MOF (UiO-66) to detect phosalone (PAS). UiO-66 was combined with Pt nanoparticles and the MIP imprinted  
562 with PAS was produced by the sol-gel method with 3-aminopropyltriethoxysilane (APTES) as the functional  
563 monomer. The resulting sensor exhibited an LOD of 0.078 nM and its feasibility was evaluated in lake water  
564 and soil samples.

565 Cypermethrin (CYP) belongs to the class of pyrethroid insecticides and its insecticidal action is more effective  
566 and less toxic compared to the organophosphates. So, this pesticide has been widely used through the world.  
567 Recently, a MIP-based sensor to determine CYP in spiked wastewater samples was reported (Atar and Yola,  
568 2018). The sensing phase of the sensor was constructed on the surface of core-shell type nanoparticles  
569 (Fe@AuNPs) incorporating two-dimensional hexagonal boron nitride (2D-hBN) nanosheets through  
570 polymerisation of Ph. The highly sensitive properties of the prepared nanocomposite allowed a superior  
571 ability of specifically binding CYP and an extraordinary LOD ( $3.0 \times 10^{-5}$  nM).

572 More recently, based on studies which showed a better performance of MIPs prepared with two or more  
573 functional monomers, Li et al. (2019d) produced a dual-monomer MIP for the analysis of CYP. Here, the  
574 preparation of the MIP sensor involved specific steps to produce a hybrid material by combining Ag and N  
575 co-doped zinc oxide (Ag-N@ZnO) with activated carbon (AC) (Fig. 4A). To produce an enlarged sensing  
576 surface and, consequently, amplify the signal, AC was chosen as a sensitising material on a GCE surface due  
577 to its low cost and easy availability. However, the authors reveal that the sole use of AC hinders the formation  
578 of a rigid layer on the electrode surface. This drawback was overcome by the addition of ZnO for better  
579 immobilisation. ZnO can easily be combined with other materials and doping can enhance its electrical  
580 conductivity. Therefore, as can be seen in Fig. 4A, Ag-N@ZnO was firstly produced by the sol-gel method and  
581 was then ultrasonically mixed with AC. After that, a suspension of the prepared solution was dropped on the  
582 GCE surface and the MIP was electropolymerised with dopamine (DA) and resorcinol (RC) as dual functional  
583 monomers. The electrochemical performance was evaluated by CV using  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  as redox probe and  
584 the sensor was applied not only to water samples, but also to a soil sample. Although this dual monomer MIP  
585 provided a novel strategy to enhance binding and affinity with target analytes, the LOD was not as low as the  
586 MIP sensor proposed by Atar and Yola (2018), in which only one functional monomer was used.

587 Electrochemiluminescence (ECL) combined with MIPs can improve the sensitivity and selectivity of the assay.  
588 The use of ECL has seen an exponential growth in electrochemical sensors. However, the use of ECL as  
589 detection technique for MIP sensors with applications in real water samples is still scarce. An example of this  
590 combination is the sensor developed by Xu et al. (2020a) to detect the insecticide cyfluthrin (CYF). They  
591 developed a MIP platform based on QDs as luminophore in the presence of  $\text{H}_2\text{O}_2$  as co-reactant. In the ECL  
592 process, both the luminophore and the co-reactant are oxidised or reduced at the electrode forming radical  
593 species. Then, an electrochemical reaction between their redox products occurs and radiation is emitted.  
594 Due to its high emission quantum yield, luminol is a classic ECL luminophore. However, an increase in the

595 demand of other alternatives has been observed because of limitations during its ECL process Li et al.  
 596 (2019a). So, in the work developed by Xu et al. (2020a), the combination of a MIP with ECL based on QDs  
 597 demonstrated high selectivity, good stability and controllability, where MWCNTs were utilised to improve  
 598 the electrocatalytic activity and minimise surface fouling on the GCE. To simplify the electrode preparation  
 599 process and improve the electrocatalytic activity, MIP-QDs were firstly synthesized by the sol-gel method  
 600 using APTES as functional monomer. Then, the resulting MIP-QDs were used to coat the surface of a GCE (Fig.  
 601 4B). After drying, MWCNTs prepared in a Nafion solution, which can facilitate the adhesion and the ionic  
 602 transportation across the electrode, were placed on the GCE surface. In the presence of CYF, the ECL and the  
 603 redox peak current intensity decreased, which was accompanied by the increase of the electron transfer  
 604 resistance. The LOD was 0.12 nM and good recoveries in seawater samples were obtained.

605 **Table 2.** Electrochemical MIP sensors constructed on different sensing platforms for the detection of  
 606 pesticides in water samples.

Target	Functional monomer	Polymerisation method / Transducer	Electrochemical techniques	Samples	Linear Range (nM)	LOD (nM)	Reference
Chlorpyrifos	PATP	Electropolymerisation, GCE, AuNPs	CV	Tap water	$5.0 \times 10^2 - 1.0 \times 10^4$	$3.3 \times 10^{2a}$	(Xie et al., 2010b)
	MAA	Bulk polymerisation, GCE	DPV	River water	$0.1 - 1.0 \times 10^4$	4.1 <sup>b</sup>	(Xu et al., 2017)
	Py	Electropolymerisation, GCE, C <sub>3</sub> N <sub>4</sub> NTs, GQDs	SWV	Industrial wastewater	$1.0 \times 10^{-2} - 1$	$2.0 \times 10^{-3c}$	(Yola and Atar, 2017)
Methyl-parathion	PATP	Electropolymerisation, GCE, AuNPs, CNTs	LSSV	Distilled water and tap water	0.38 – 4.2 4.2 – 42	0.30 <sup>a</sup>	(Wu et al., 2014)
	Ph	Electropolymerisation, SPAuE, N-GS	CV	River water	$3.8 \times 10^2 - 3.8 \times 10^4$	38 <sup>d</sup>	(Xue et al., 2014)
Parathion	MAA	Bulk polymerisation, CPE	SWV	Tap, river and lake water	$1.7 - 9.0 \times 10^2$	0.5 <sup>d</sup>	(Alizadeh, 2009)
Diazinon	MAA	Bulk polymerisation, CPE	SWV	Well water	$2.5 - 1.0 \times 10^2$ $1.0 \times 10^2 - 2.0 \times 10^3$	0.79 <sup>b</sup>	(Motahari an et al., 2016)
	MAA	Bulk polymerisation, CPE, MWNTs	SWV	Tap and river water	$0.5 - 1.0 \times 10^3$	0.13 <sup>b</sup>	(Khadem et al., 2017)
Phosalone	APTES	Sol-gel method, CPME, Pt-UiO-66	SWV	Lake water and soil	$0.50 - 2.0 \times 10^4$	0.078 <sup>b</sup>	(Xu et al., 2020b)
Cypermethrin	Ph	Electropolymerisation, GCE, Fe@AuNPs, 2D-hBN	DPV	Wastewater	$1.0 \times 10^{-3} - 10$	$3.0 \times 10^{-5d}$	(Atar and Yola, 2018)
	DA and RC	Electropolymerisation, GCE, Ag-N@ZnO, CHAC	EIS	Tap water and soil	$2.0 \times 10^{-4} - 8$	$6.7 \times 10^{-5e}$	(Li et al., 2019d)
Cyfluthrin	APTES	Sol-gel method, QDs, Nafion-MWCNTs	ECL	Seawater	$0.46 - 2.3 \times 10^2$	0.12 <sup>a</sup>	(Xu et al., 2020b)
Triazophos	PATP	Electropolymerisation, Au-electrode, luminol	ECL	Tap water, reservoir water and river water	$0.1 - 1.0 \times 10^3$	0.058 <sup>d</sup>	(Li et al., 2019a)
Glyphosate	Py	Electropolymerisation, Au-electrode	DPV	Tap water	$30 - 4.7 \times 10^3$	1.6 <sup>b</sup>	(Zhang et al., 2017)
	MAC	Bulk polymerisation, PGE, AuNPs, MWCNTs	DPASV	Soil	$49 - 4.7 \times 10^2$	2.1 <sup>f</sup>	(Prasad et al., 2014)

	PATP	Electropolymerisation, Au-electrode, AuNPs	LSV	Tap water	$5.9 \times 10^{-6} - 5.9$	$4.7 \times 10^{-6a}$	(Do et al., 2015)
<b>2,4-dichlorophenol</b>	MAA	Bulk polymerisation, GCE, microgel suspension, Ch, Nafion	DPV	Tap, river and drinking water	$5.0 \times 10^3 - 1.0 \times 10^5$	$1.6 \times 10^{3e}$	(Zhang et al., 2013)
	MAA	Bulk polymerisation, GCE, GO	DPV	Lake water	$4 - 1.0 \times 10^4$	$0.5^g$	(Liang et al., 2017)
	EDOT	Electropolymerisation, CFP	DPV	Lake, river and tap water	$0.21 - 3.0 \times 10^2$	$0.07^g$	(Maria G.C. et al., 2020)
<b>4-Chlorophenol</b>	o-PD	Electropolymerisation, GCE, ZnO NPs	SWV	Wastewater	$2.0 \times 10^2 - 1.7 \times 10^5$	$40^d$	(AL-Ammari et al., 2019)
<b>Atrazine</b>	Ph	Electropolymerisation, GCE, PtNPs, C <sub>3</sub> N <sub>4</sub> NTs	SWV	Wastewater	$1.0 \times 10^{-3} - 1.0 \times 10^{-1}$	$1.5 \times 10^{-4d}$	(Yola and Atar, 2017)
<b>2,4-Dichlorophenoxy acetic acid</b>	Py	Electropolymerisation, GCE	CV	Tap water	$1.0 \times 10^{-3} - 1.0 \times 10^4$	$8.3 \times 10^{2a}$	(Xie et al., 2010a)
<b>Paraquat</b>	Py	Electropolymerisation, PGE, EBB	DPV	Dam water	$5.0 \times 10^3 - 5.0 \times 10^4$	$2.2 \times 10^{2a}$	(Sayyahnesh et al., 2016)
<b>Diuron</b>	MAA	Bulk polymerisation, CPE, MWCNTs	SWV	River water	$52 - 1.3 \times 10^3$	$9.0^b$	(Wong et al., 2015)
<b>Chloridazon</b>	2-VP and MAA	Bulk polymerisation, CPE, MWCNTs	DPV	Ground, surface, seawater and drinking water	$5.0 \times 10^2 - 4.0 \times 10^5$	$62^d$	(Ghorbani et al., 2020)
<b>Hexazinone</b>	2-VP	Bulk polymerisation, CPE	DPV	River water	$0.019 - 0.11$	$2.6 \times 10^{-3d}$	(Toro et al., 2015)
<b>Dicloran</b>	MAA	Bulk polymerisation, CPE, MWCNTs	SWV	Tap water and river water	$1.0 \times 10^3 - 1$	$0.48^d$	(Khadem et al., 2016)
<b>Tributyltin</b>	APTES	Sol-gel method, SPCE, Fe <sub>3</sub> O <sub>4</sub> MNPs	EIS	Sea water	$5.0 \times 10^{-3} - 5.0 \times 10^3$	$5.4 \times 10^{-3g}$	(Zamora-Gálvez et al., 2017)

607

<sup>a</sup> LOD calculated as 3 times the standard deviation of the blank.

608

<sup>b</sup> LOD calculated as 3 times the standard deviation of the blank divided by the slope.

609

<sup>c</sup> LOD calculated as 3.3 times the standard deviation of the intercept divided by the slope.

610

<sup>d</sup> LOD formula no specified.

611

<sup>e</sup> LOD calculated as  $S/N = 3$ .

612

<sup>f</sup> LOD based on the minimal distinguishable signal for lower concentration of analyte.

613

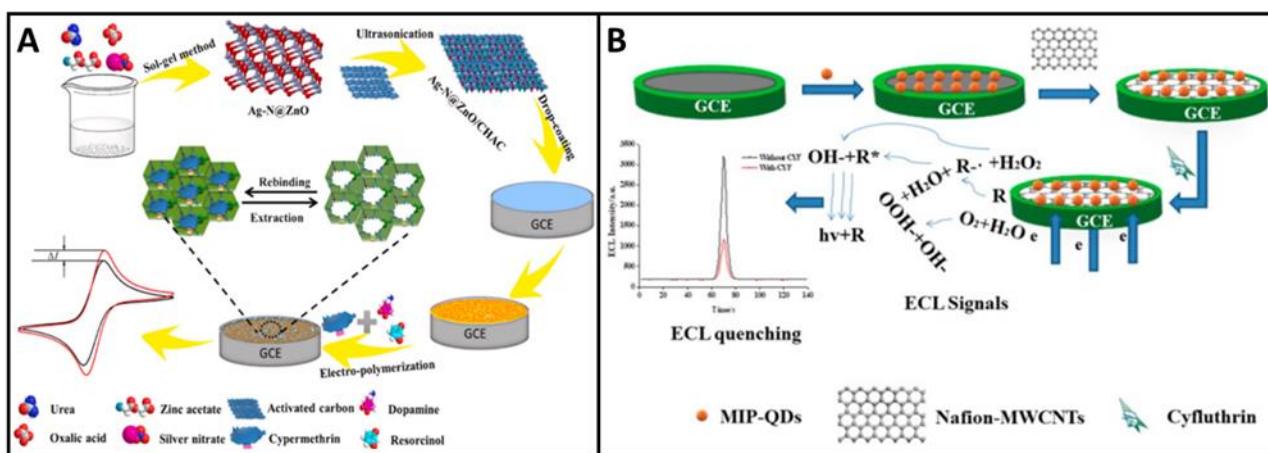
<sup>g</sup> LOD calculated as 3 times the standard deviation of the lowest concentration (or any concentration) divided by the slope.

614

2D-hBN: two dimensional hexagonal boron nitride; 2-VP: 2-vinylpyridine; APTES: (3-Aminopropyl)triethoxysilane; Ag-N@ZnO: silver and nitrogen co-doped zinc oxide; C<sub>3</sub>N<sub>4</sub> NTs: carbon nitride nanotubes; CFP: carbon fiber paper; Ch: chitosan; CHAC: activated carbon prepared from coconut husk; CMPE: carbon paste microelectrode; CNTs: carbon nanotubes; CPE: carbon paste electrode; CV: cyclic voltammetry; DA: dopamine; DPASV: differential pulse anodic stripping voltammetry; EBB: eriochrome blue-black B; ECL: electrochemiluminescence; EDOT: 3,4-ethylenedioxythiophene; Fe@AuNPs: core-shell type nanoparticles; GQDs: graphene quantum dots; LSSV: linear stripping sweep voltammetry; MAC: N-methacryloyl-L-cysteine; N-Gs: nitrogen doped graphene sheets; Ph: phenol; Pt-UiO-66: Zr-based metal-organic framework catalyst; QDs: quantum dots; RC: resorcinol; ZnO NPs: zinc oxide nanoparticles.

621





622

623

624

625

626

627

### 3.2.2 Herbicides

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647

648

649

650

651

652

653

654

655

656

657

658

659

**Figure 4.** A) Schematic representation of the fabrication and characterisation of a dual MIP monomer based on Ag–N@ZnO/CHAC/GCE. Reproduced from Li et al. (2019d) with permission from Elsevier. B) Schematic representation of a MIP ECL sensor to detect CYF. Reproduced from Xu et al. (2020a).

Glyphosate (Gly) is among the mostly widely used herbicides by farmers during the past 40 years. Although not proven, Gly has been associated with cancer in humans and a strong debate about its potential harmfulness has been generated (Silva et al., 2018). Due to its persistence in seawater (Mercurio et al., 2014), the need to identify trace levels of Gly in drinking water is also urgent. To the best of our knowledge, the most recent electrochemical MIP-sensor used for Gly detection in water samples was reported by Zhang et al. (2017). In this work, a simple electropolymerisation procedure, using Py as functional monomer, on an Au electrode was performed. The sensor presented good binding kinetics to Gly and showed good stability, selectivity, and sensitivity, and an LOD of 1.60 nM. The analytical signal was based on the use of the  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  redox probe and DPV. This MIP sensor exhibited a higher sensitivity when compared with the MIP sensor constructed by Prasad et al. (2014), where N-methacryloyl-L-cysteine (MAC) molecules were used as monomer in the bulk polymerisation process. However, the best sensitivity to detect GLY in water samples was obtained by the construction of an electrochemical sensor based on MIP-MOF films formed on Au surfaces through electropolymerisation of PATP functionalised with AuNPs (Do et al., 2015). A low LOD of  $4.7 \times 10^{-6}$  nM was achieved and the sensing capacity of the sensor was evaluated in tap water.

All other published MIP-sensors to detect herbicides with applications in water samples were based on the modification of GCE (AL-Ammari et al., 2019; Liang et al., 2017; Xie et al., 2010a; Yola and Atar, 2017; Zhang et al., 2013), PGE (Sayyahmanesh et al., 2016) or CPE (Ghorbani et al., 2020; Khadem et al., 2017; Toro et al., 2015; Wong et al., 2015). Additionally, a newly developed carbon fibre paper used as working electrode was also reported (Maria et al., 2020). In order to enhance the selectivity and precision, various nanomaterials were combined in these sensors.

As can be seen in Table 2, the LOD for 2,4-dichlorophenol (2,4-DCP), a typical chlorophenol that is widely employed in the synthesis of herbicides and insecticides, was much lower in the presence of GO (Liang et al., 2017). Moreover, Liang et al. (2017), using the same functional monomer (MAA), proposed a very simple methodology for the fabrication of a 2,4-DCP-MIP electrochemical sensor in comparison with Zhang et al. (2013). Besides using MAA as functional monomer, Zhang et al. (2013) also used a co-monomer, chlorohemin, to introduce chemically active sites into the MIP as well as a combination of chitosan (Ch) and Nafion for immobilisation of the MIP and to increase conductivity, respectively. This assembly probably blocked the mass transport to the electrode surface which resulted in a worse LOD. More recently, one of the first reports using a poly(3,4-ethylenedioxythiophene) (PEDOT) based MIP on carbon fibre paper, and the first applied to water samples, was published by Maria et al. (2020). To detect 2,4-DCP, carbon fibre paper was chosen as working electrode because of its porosity, offering abundant reaction sites. This novelty and the use of PEDOT as conductive polymer not only improved the LOD (0.07 nM), but also the stability and reproducibility. Other

660 approach was presented by AL-Ammari et al. (2019), in this case, to explore the detection of 4-chlorophenol  
661 (4-CP). The MIP was prepared by electropolymerisation of o-PD with multifunctional nanomaterials, zinc  
662 oxide nanoparticles (ZnO NPs) and graphene platelets, on a GCE. Compared with other techniques applied  
663 for the detection of 4-CP, this MIP, which was tested in real wastewater samples, showed higher sensitivity.  
664 Another excellent approach to improve the LOD was based on combining Pt NPs and C<sub>3</sub>N<sub>4</sub> NTs, which were  
665 obtained by hydrothermal treatment and allowed minimised waste formation. Yola and Atar (2017)  
666 employed these nanomaterials to modify a GCE surface and, through electropolymerisation of Ph, they  
667 created a sensor that showed an LOD of  $1.5 \times 10^{-4}$  nM for the analysis of atrazine (ATR). PATP was also  
668 investigated for the determination of 2,4-dichlorophenoxy acetic acid (2,4-D) (Xie et al., 2010a). In this work,  
669 a simple procedure which involved electropolymerisation on the surface of a GCE, was developed. The 2,4-D  
670 molecules were removed from the polymeric matrix without solvent extraction, which is in accordance with  
671 “green chemistry”. However, this simple sensor did not show a great LOD.  
672 Due to the favourable features of Py in aqueous media, it was chosen again as functional monomer for the  
673 recognition of paraquat in dam water (Sayyahmanesh et al., 2016). The electropolymerisation of the MIP  
674 occurred on top of a PGE in the presence of a functional doping ion (eriochrome blue-black B (EBB)). EBB is  
675 an anionic complexing agent, which was used to maintain the electroneutrality during the reduction of Py  
676 and, consequently, to conserve its electroactivity. In order to enhance the electrochemical signal of diuron,  
677 a CPE modified with a MIP and MWCNTs was proposed by Wong et al. (2015). The MIP was synthesized via  
678 bulk polymerisation. This polymerisation technique was also adopted by (Ghorbani et al., 2020) for the  
679 determination of chloridazon (CLZ) in surface, ground and drinking water. This MIP sensor was constructed  
680 based on the multiple interactions formed between CLZ and two functional monomers, 2-vinylpyridine (2-VP)  
681 and MAA. In the same way, but without the use of nanomaterials, Toro et al., 2015 developed a selective  
682 sensor for hexazinone (HXZ) using computational simulations to evaluate the interaction of HXZ with  
683 functional monomers in the prepolymerisation mixture. MAA was selected as the most suitable monomer  
684 and this computational study allowed the production of a more effective MIP for HXZ with an excellent LOD  
685 of  $2.6 \times 10^{-3}$  nM.

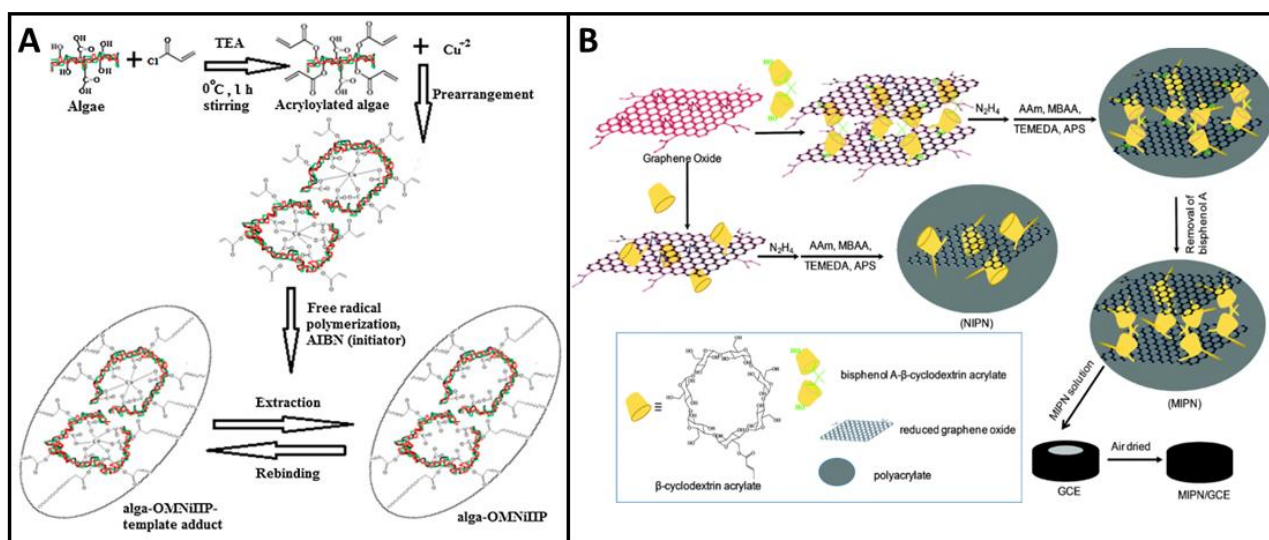
### 686 3.2.3 Fungicides and biocides

687 Compared with insecticides and herbicides, the number of publications on the analysis of fungicides is much  
688 lower (see Table 2). An electrochemical MIP-sensor for the determination of dicloran in tap water and river  
689 water was developed by Khadem et al. (2016). In this work, the MIP produced by bulk polymerisation was  
690 deposited on top of a CNTs-modified CPE. The sensor showed very high recognition ability in comparison  
691 with a NIP electrode with a linear range between 1 and  $1.0 \times 10^3$  nM and an LOD of 0.48 nM.  
692 Besides this, a MIP sensor for tributyltin (TBT), a biocide used in antifoulant paints to prevent the growth of  
693 marine organisms on the hulls of large ships, was also described (Zamora-Galvez et al., 2017). Fe<sub>3</sub>O<sub>4</sub> MNPs  
694 were used to modify the MIP prepared by the sol-gel process using APTES as monomer. Using an SPCE as  
695 transducer and EIS as detection technique, a low LOD was achieved ( $5.4 \times 10^{-3}$  nM).

### 696 3.3 Heavy metals

697 In developing countries, the increase of industrial activity and urbanisation has led to the proliferation of  
698 heavy metals in the soil and water sources (Ali et al., 2019a). The pollution of the environment with heavy  
699 metals has a particular impact on ecotoxicology because of their long persistence, non-degradability,  
700 bioaccumulation and biomagnification in the food chain, which causes serious harm not only to wildlife, but  
701 also to human health (Hong et al., 2020; Kumar et al., 2019). The MIP technology has attracted considerable  
702 attention to assess and monitor the concentration of heavy metals (Liu et al., 2019; Yaroshenko et al., 2020).  
703 In the field of separation and sensing, several works involving the adsorption of heavy metals using MIPs can  
704 be found in the literature (Sharma and Kandasubramanian, 2020). However, as shown in Table 3, only a small  
705 number of publications concerning electrochemical MIP-based sensors to detect metal ions with applications  
706 in water and soil samples were reported. Electrochemical MIP sensors for the analysis of beryllium(II) (Be<sup>2+</sup>)  
707 (Li et al., 2015), cobalt(II) (Co<sup>2+</sup>) (Li et al., 2019), copper(II) (Cu<sup>2+</sup>) (Di Masi et al., 2020; Prasad and Fatma, 2016;  
708 Prasad and Singh, 2016) and zinc(II) (Zn<sup>2+</sup>) (Shirzadmehr et al., 2016) were developed following different

709 strategies to facilitate metal ion imprinting and improve the selectivity of the sensor. The combination of MIP  
 710 and ECL was reported for ultra-trace  $\text{Be}^{2+}$  detection in water samples (Li et al., 2015). The sensor was  
 711 fabricated using  $\text{Be}^{2+}$  and 4-(2-pyridylazo)-resorcinol (PAR), a metallochromic indicator, to form a complex  
 712 which was copolymerised with o-PD. CV characterisation of the prepared MIP sensor revealed that the  
 713 imprinted cavities could only recognise the  $\text{Be}^{2+}$ -PAR complex and not the metal ion and the ligand, showing  
 714 an efficient imprinting process. ECL was indirectly measured by the luminol- $\text{H}_2\text{O}_2$  interaction and a low LOD  
 715 (0.024 nM) was achieved. The same strategy for the detection of nanomolar levels of  $\text{Co}^{2+}$  was used by Li et  
 716 al. (2019c). In their work,  $\text{Co}^{2+}$  and bovine serum albumin (BSA) were introduced as the template molecules  
 717 and an ECL sensor was designed. Another attractive and different concept, based on biosorption, was  
 718 described to detect  $\text{Cu}^{2+}$  in soil and water samples (Prasad and Fatma, 2016; Prasad and Singh, 2016). As  
 719 certain algae show enormous potential for biosorption of heavy metals, Prasad and Fatma (2016) used a blue  
 720 green-algae (*Aulospira* sp.) modified PGE for the development of one MoNomer Ion Imprinted Polymer  
 721 (OMNiMIP) (Fig. 5A). In this procedure, the functionalised algae were simultaneously used as crosslinker and  
 722 functional monomer. The use of carboxylic groups in the OMNiMIP sensor facilitated the metal ion imprinting  
 723 and a highly sensitive method with precise results was obtained. The absence of an extra crosslinker allowed  
 724 the LOD to be several times lower than the one obtained by Prasad and Singh (2016). In this work, the same  
 725 algae were employed as a natural ligand to form a “complex-metal” with N-methacryloylglutamic acid  
 726 (NMGA) as the monomer for the analysis of  $\text{Cu}^{2+}$ . The algae were considered as cheap substitutes of MWCNTs  
 727 to impart electroconductivity to the film. Using differential pulse anodic stripping voltammetry (DPASV), an  
 728 extremely sensitive technique used for the detection of heavy metal ions, Prasad and Singh (2016) developed  
 729 a OMNiMIP without the need to select a crosslinker and, consequently, without optimisation of the  
 730 monomer-crosslinker ratio.



731  
 732 **Figure 5.** A) Illustration of the preparation of an alga-OMNiMIP for electrochemical detection of  $\text{Cu}^{2+}$ . Reproduced  
 733 from Prasad and Fatma (2016) with permission from Elsevier. B) Schematic illustration of the synthesis procedure of a  
 734 MIP sensor for BPA analysis. Reproduced from Ali et al. (2019b) with permission from the Centre National de la  
 735 Recherche Scientifique (CNRS) and The Royal Society of Chemistry.

736

737

738

739

740

741 **Table 3.** Electrochemical MIP sensors constructed in different sensing platforms for detection of heavy metals  
 742 in water samples.

Target	Functional monomer	Polymerisation method / Transducer	Electrochemical techniques	Samples	Linear Range (nM)	LOD (nM)	Reference
<b>Beryllium(II) (Be<sup>2+</sup>)</b>	o-PD	Electropolymerisation, SPAuE, luminol-H <sub>2</sub> O <sub>2</sub>	ECL	Rain, bottled and well water	0.07 – 8.0	0.024 <sup>a</sup>	(Li <i>et al.</i> , 2015)
<b>Cobalt(II) (Co<sup>2+</sup>)</b>	o-AP	Electropolymerisation, Au electrode, BSA, MWCNTs, Cu, C-Dots	ECL	Surface water, industrial sewage and agricultural soil	1 – 1.0×10 <sup>2</sup>	0.31 <sup>a</sup>	(Li <i>et al.</i> , 2019c)
<b>Copper(II) (Cu<sup>2+</sup>)</b>	Acryloylated-algae	Bulk polymerisation, PGE	DPASV	Lake water and soil	0.13 – 1.2×10 <sup>2</sup>	0.028 <sup>b</sup>	(Prasad and Fatma, 2016)
	NMGA	Bulk polymerisation, PGE	DPASV	Lake water and soil	0.16 – 98	0.060 <sup>b</sup>	(Prasad and Singh, 2016)
	o-PD	Electropolymerisation, SPpTE	DPV	Drinking water	0.95 – 2.4×10 <sup>2</sup>	2.7 <sup>c</sup>	(Di Masi <i>et al.</i> , 2020)
<b>Zinc(II) (Zn<sup>2+</sup>)</b>	MAA	Bulk polymerisation, CPE, GO, AgNPs, [BMP]Tf2N (IL)	Potentiometry	River water, industrial wastewater	3.9 – 1.0×10 <sup>10</sup>	0.030 <sup>d</sup>	(Shirzadmehr <i>et al.</i> , 2016)

743 <sup>a</sup> LOD calculated as 3 times the standard deviation of the intercept divided by the slope.

744 <sup>b</sup> LOD calculated as 3 times the standard deviation of the blank divided by the slope.

745 <sup>c</sup> LOD calculated as 3 times the standard error from the weighted regression plot divided by the slope.

746 <sup>d</sup> LOD formula no specified.

747 AgNPs: silver nanoparticles; [BMP]Tf2N (IL): ionic liquid 1-butyl-1-methylpyrrolidinium bis(trifluoromethylsulfonyl)imide; BSA: bovine  
 748 serum albumin; C-Dots: carbon dots; NMGA: N-methacryloylglutamic acid; o-AP: o-aminophenol; SPpTE: screen printed platinum  
 749 electrode.

750

### 751 3.4 Other contaminants

752 Besides pharmaceuticals, pesticides and heavy metals, the environment is also charged with other chemicals  
 753 that may harm human health. Many products of daily use, such as industrial explosives, sub-products of  
 754 industrial syntheses, ingredients of cosmetics, brominated flame retardants or even amino acids have not  
 755 surprisingly been detected in environmental matrices. As can be observed in Table 4, a substantial amount  
 756 of work has been done to develop novel sensors to determine the presence of these unwanted compounds  
 757 in different environmental water samples. In this section, emphasis will only be given to new design strategies  
 758 of MIPs.

759 Since 2011, bisphenol A (BPA) has attracted widespread attention as contaminant. It is mainly used to  
 760 produce epoxy resins and polycarbonate plastics and is found in bottles, food containers and toys. BPA is  
 761 considered an endocrine disruptor and human exposure leads to a variety of health issues (Ali *et al.*, 2019b).  
 762 Various electrochemical strategies have been used for the selective determination of BPA in different types  
 763 of waters (Ali *et al.*, 2019b; Chen *et al.*, 2014b; Dadkhah *et al.*, 2016; Tan *et al.*, 2016; Wang *et al.*, 2011; Zhu  
 764 *et al.*, 2014) (see Table 4). There is no consensus on the technique used for its molecular imprinting, but the  
 765 presence of nanomaterials was essential for the adequate performance of the sensors. Recently, Ali *et al.*  
 766 (2019b) reported a very robust MIP based on a nanocomposite of polyacrylate, reduced GO (rGO) and  $\beta$ -  
 767 cyclodextrin that were covalently linked forming a 3D network (Fig. 5B).  $\beta$ -cyclodextrin is known to form an  
 768 inclusion complex with BPA by means of host-guest complexation, while polyacrylate allows to form porous  
 769 3D networks. The first step of the synthesis consisted of the reaction between GO and  $\beta$ -cyclodextrin in the  
 770 presence of BPA; the hydroxyl groups of  $\beta$ -cyclodextrin strongly react with the epoxide groups of GO and a  
 771 stable interaction is obtained. Then, radical polymerisation was performed using two functional monomers,  
 772 acrylamide (AA) and N,N'-methylenebis-acrylamide (MBAA). The electrochemical detection of BPA was  
 773 performed using GCE as transducer, reaching an interesting LOD (8 nM). In this work the combination of rGO

774 and  $\beta$ -cyclodextrin in molecular imprinting was explored for the first time, providing promising insights for  
 775 the design of more selective MIPs.  
 776 The same LOD was observed with a low-cost imprinted sensor based on laser scribed graphene (LSG)  
 777 technology (Beduk et al., 2020). This device was used as transducer to detect BPA after electropolymerisation  
 778 of Py. Dadkhah et al. (2016) and Wang et al. (2011) developed strategies to prepare MIPs on the surface of  
 779 GO and hexagonally structured mesoporous silica, respectively, to enhance the electrochemical signal. In  
 780 both works, amine functionalisation of GO and SiO<sub>2</sub> NPs was achieved by a simple procedure using APTES to  
 781 improve the recognition ability of the sensors. After functionalisation, the targets were immobilised on its  
 782 surface and not only many homogenous imprinting sites were formed, but also an easy access of the analyte  
 783 molecules to the imprinting sites was reported. At the same time, the best LOD found for BPA was 3 nM using  
 784 GO in the modification of the electrochemical MIP sensor. Recently, ECL again showed its potential to achieve  
 785 a higher sensitivity (Zhang et al., 2020). These authors developed a MIP sensor based on a GCE modified with  
 786 Fe<sub>3</sub>O<sub>4</sub> nanocrystals (Fe<sub>3</sub>O<sub>4</sub>-NCs), which increased the ECL response signal of luminol. The proposed MIP  
 787 sensor displayed an excellent analytical performance and a low LOD ( $1.8 \times 10^{-3}$  nM) was observed.  
 788 Another compound that is widely used in epoxy and polycarbonate resins is tetrabromobisphenol A (TBBPA),  
 789 which is the most common brominated flame retardant on the market (Zhou et al., 2016). TBBPA has also  
 790 been linked with endocrine disruption. For its electrochemical detection, Chen et al. (2014a) presented a MIP  
 791 based on nickel nanoparticle-modified GO and showed that the combination of more than one  
 792 nanostructured material increased the effective surface that can be an attractive route for the preparation  
 793 of MIP-based sensors with adequate sensitivities.

794 **Table 4.** Electrochemical MIP sensors constructed on different sensing platforms for the detection of various  
 795 contaminants in water samples.

Target	Functional monomer	Polymerisation method / Transducer	Electrochemical techniques	Samples	Linear range (nM)	LOD (nM)	Reference
<b>2,4,6-Trinitrotoluene</b>	MAA	Bulk polymerisation, CPE	SWV	Tap water, ground water and soil	5 – $1.0 \times 10^3$	1.5 <sup>a</sup> 5 <sup>a</sup>	(Alizadeh <i>et al.</i> , 2010)
	MAA	Bulk polymerisation, CPE, Fe <sub>3</sub> O <sub>4</sub> MNPs	SWV	Tap and sea water	$1.0 - 1.3 \times 10^1$	0.5 <sup>a</sup>	(Alizadeh, 2014)
<b>Bisphenol A</b>	MMA	Miniemulsion polymerisation, CPE, Fe <sub>3</sub> O <sub>4</sub> MNPs, CTAB	EIS	Drinking bottle and lake water	$6.0 \times 10^2 - 1.0 \times 10^5$	$1.0 \times 10^{2a}$	(Zhu <i>et al.</i> , 2014)
	4-VP	Bulk polymerisation, CPE, MWCNTs	DPV	River, tap and pure water	$80 - 1.0 \times 10^5$	22 <sup>b</sup>	(Chen <i>et al.</i> , 2014b)
	AA and MBAA	Bulk polymerisation, GCE, $\beta$ -CD, GO	DPV	Lake, tap and drinking water	$20 - 1.0 \times 10^3$	8 <sup>c</sup>	(Haydar Ali <i>et al.</i> , 2019)
	Py	Electropolymerisation, GCE, GQDs	DPV	Tap and sea water	$1.0 \times 10^2 - 5.0 \times 10^4$	40 <sup>a</sup>	(Tan <i>et al.</i> , 2016)
	Py	Electropolymerisation, LSG electrode	DPV	Mineral and tap water	$50 - 2.0 \times 10^4$	8 <sup>a</sup>	(Beduk <i>et al.</i> , 2020)
	APTES	Sol-gel method, GCE, GO	DPV	Mineral water	$6 - 1.0 \times 10^2$ $2.0 \times 10^2 - 2.0 \times 10^4$	3 <sup>b</sup>	(Dadkhah <i>et al.</i> , 2016)
	APTES	Sol-gel method, CPE, SiO <sub>2</sub> NPs	CV	River water	$1.0 \times 10^2 - 5.0 \times 10^5$	32 <sup>c</sup>	(Wang <i>et al.</i> , 2011)
APTES	Sol-gel method, GCE, Fe <sub>3</sub> O <sub>4</sub> -NCs	ECL	Seawater	$8.8 \times 10^{-3} - 2.2 \times 10^4$	$1.8 \times 10^{-3d}$	(Zhang <i>et al.</i> , 2020)	

<b>Tetrabromobisphenol A</b>	Py	Electropolymerisation, GCE, GO, NiNPs	DPV	Tap water, rain and lake water	0.5 – 1.0×10 <sup>4</sup>	0.13 <sup>a</sup>	(Chen <i>et al.</i> , 2014a)
	APTES	Sol-gel method, CPE, Fe <sub>3</sub> O <sub>4</sub> MNPs	DPV	Tap water, rain and pool water	5.0 – 2.0×10 <sup>3</sup>	0.77 <sup>b</sup>	(Zhou <i>et al.</i> , 2016)
<b>Tetrabromobisphenol S</b>	MAA and PATP	Bulk polymerisation, CPE, AuNPs	DPV	Tap, lake and drinking water	0.1 – 10	0.029 <sup>b</sup>	(Sarpong <i>et al.</i> , 2020)
<b>Sodium Lauryl sulfate</b>	2-ATP	Electropolymerisation, SPAuE	DVP	Wastewater and river water	0.1 – 3.5	6.3×10 <sup>-4b</sup>	(Motia <i>et al.</i> , 2018)
<b>3-Methylindole</b>	o-PD	Electropolymerisation, GCE	CV	Tap and lake water	10 – 1.2×10 <sup>3</sup>	4 <sup>d</sup>	(Yu <i>et al.</i> , 2019)
<b>4-nonylphenol</b>	Py	Electropolymerisation, GCE, GO	DPV	Rain and lake water	0.45 – 45	0.016 <sup>a</sup>	(Chen <i>et al.</i> , 2013)
<b>Para-nitrophenol</b>	MAA	Bulk polymerisation, CPE	DPV	River and tap water	8 – 5.0×10 <sup>3</sup>	3 <sup>c</sup>	(Alizadeha <i>et al.</i> , 2009)
<b>N-Nitrosodimethylamine</b>	Py	Electropolymerisation, GCE, SWCNTs	EIS	Bottled drinking water and tap water	1.4×10 <sup>2</sup> – 3.1×10 <sup>3</sup>	11.5 <sup>d</sup>	(Cetó <i>et al.</i> , 2016)
<b>L-cystein</b>	MAA	Bulk polymerisation, CPE	DPV	Tap water	20 – 1.8×10 <sup>2</sup>	9.6 <sup>a</sup>	(Aswini <i>et al.</i> , 2014)
<b>Triclosan</b>	AA	Bulk polymerisation, SPAuE, PVC-COOH	DPV	Wastewater and mineral water	3.5×10 <sup>-4</sup> – 3.5×10 <sup>2</sup>	7.9×10 <sup>-4d</sup>	(Motia <i>et al.</i> , 2019)
<b>Methyl green dye</b>	AA	Bulk polymerisation, CPE, Fe <sub>3</sub> O <sub>4</sub>	SWAdASV	River water and industrial wastewater	99 – 1.8×10 <sup>3</sup>	10 <sup>c</sup>	(Khan <i>et al.</i> , 2019)
<b>Diphenylamine</b>	([VC4mim][PF6] (IL))	Bulk polymerisation, GCE, RGO, Fe <sub>3</sub> O <sub>4</sub>	DPV	Lake water	1.0×10 <sup>2</sup> – 1.8×10 <sup>3</sup>	50 <sup>a</sup>	(Liu <i>et al.</i> , 2018)

796 <sup>a</sup> LOD calculated as S/N = 3.

797 <sup>b</sup> LOD calculated as 3 times the standard deviation of the blank divided by the slope.

798 <sup>c</sup> LOD formula no specified.

799 <sup>d</sup> LOD calculated as 3 times the standard deviation of the blank.

800 2-ATP: 2-aminothiophenol; 4-VP: 4-vinylpyridine; AA: acrylamide; CTAB: Cetyltrimethylammonium bromide; MBAA: N,N'-methylenebis-acrylamide; NCs: nanocrystals; NiNPs: nickel nanoparticles; LSG: laser scribed graphene; PVC-COOH: carboxylic polyvinyl chloride; RGO: reduced Graphene oxide; SiO<sub>2</sub> NPs: silica nanoparticles; SWAdAS: square-wave adsorptive anodic stripping; 802 SWCNTs: single walled carbon nanotubes; ([VC4mim][PF6] (IL): ionic liquid 1-vinyl-3-butylimidazolium hexafluorophosphate. 803

804

805 Incorporation of ionic liquids (ILs) in the MIP framework has also been the focus of many studies. By exploring 806 their good electrocatalytic activity, adsorption capacity and multiple interactions with targets, there are some 807 works adopting polymerisable ILs as functional monomers and crosslinkers to prepare MIPs (Ding *et al.*, 808 2020). Liu *et al.* (2018) introduced a new IL composite using IL 1-vinyl-3-butylimidazolium 809 hexafluorophosphate ([VC4mim][PF6]) as functional monomer, IL 1,4-butanediyl-3,3'-bis-1-vinylimidazolium 810 dihexafluorophosphate ([V2C4(mim)2][(PF6)2]) as crosslinker and GO and Fe<sub>3</sub>O<sub>4</sub> as support, to detect 811 diphenylamine (DPA) in water samples. In their work the authors showed that the proposed MIP based on 812 such IL as crosslinker had a higher electrochemical response than that of the traditional ethylene glycol 813 dimethacrylate. Although, ILs have been considered alternative green solvents, it is important to note that 814 ILs are quite expensive and their recycling is difficult.

815 Another strategy in surface imprinting was shown by Motia et al. (2019) through the functionalisation of an  
816 SPAuE with a carboxylic polyvinyl chloride (PVC–COOH) layer for the detection of triclosan (TCS), which is an  
817 antibacterial agent incorporated in many products of daily use. After linking TCS with PVC, strong hydrogen  
818 bonds between the polar groups of TCS and –COOH groups of the PVC were established.

819

#### 820 **4. Conclusions and perspectives**

821 Environmental water quality monitoring is essential to ensure the safety of the aquatic environment and,  
822 consequently, human and animal health. Moreover, the analysis of contaminants in the environment is  
823 essential to know their pathways, fate and effects, but also to determine the WWTP's removal efficiency.

824 In this review, the considered studies demonstrated a significant increase in the development of MIP-based  
825 electrochemical sensors to detect contaminants in water samples. MIPs are polyvalent receptors, which are  
826 easily synthesized, modified and manipulated and, more importantly, they are compatible with water. The  
827 MIP's sensitivity and affinity are usually improved by their combination with nanomaterials. This has  
828 stimulated researchers to design and create new and innovate MIP sensors with the ability to recognise the  
829 target molecules at trace levels. However, although MIPs have proven their potential as recognition  
830 elements, the need of extremely low LODs remains an open challenge. This is the reason why many MIP  
831 sensors for determination of pharmaceuticals are not applied to water analysis but to serum and/or urine  
832 (for clinical control) or pharmaceutical formulations (for quality control).

833 Regarding the imprinting process, there is no specific approach for a particular class of molecules. So, the  
834 synthesis process and functional monomer that provide the best MIP for the target molecule have to be  
835 obtained by experimental studies. In this context, computational studies, reported only in few publications,  
836 are increasingly used since they have proven to be valuable for selecting suitable functional monomers.  
837 Besides the fact that computational modelling allows a better understanding of the monomer-template  
838 interactions, it provides important guidelines and, therefore, leads to an overall more environmentally  
839 friendly process for MIP construction. Accordingly, more efforts should be made to exploit theoretical  
840 approaches to pre-screen the preparation conditions, since good selectivities and LODs are achieved.  
841 Regarding the transducers, carbon electrodes are undoubtedly the most widely reported sensing platform  
842 for the determination of contaminants. Regarding the detection strategy, the highest sensitivity is usually  
843 attained by indirect electrochemical sensing, although this strategy can have some drawbacks, such as false  
844 results due to the presence of masked interferents.

845 Although MIPs are highly selective, this can also be a disadvantage when the determination of several  
846 contaminants is required. Thus, the development of MIPs capable to recognise more than one contaminant  
847 (for example, a family of contaminants with similar molecular structures) could be useful for some  
848 applications. Another approach could be the development of multiplex MIP-based sensors, although  
849 nowadays they are scarce for environmental applications. Moreover, portable electrochemical sensors show  
850 another difficulty for their application in environmental analysis: analytes have to be extracted from solid  
851 samples (such as soils) to allow analysis. Liquid samples, mainly water, are easier to analyse (since they are  
852 aqueous); however, sometimes they present a complex matrix that could interfere in the analysis.

853 Therefore, continuous studies within multidisciplinary teams must be performed to develop highly sensitive  
854 and accurate methods, aiming to implement MIPs in the analysis of contaminants in environmental waters.  
855 The construction of portable MIP sensors, especially by using screen printed electrodes, could be very useful  
856 in field applications. However, the commercialisation of these sensors remains a challenge, since, as for other  
857 kind of sensors (biological sensors and sensors for food applications), the knowledge transfer from  
858 laboratories to the market is difficult.

#### 859 **Acknowledgements**

860 The authors are grateful for the financial support from the Fundação para a Ciência e a Tecnologia (FCT) / the  
861 Ministério da Ciência, Tecnologia e Ensino Superior (MCTES) through national funds (Portugal)  
862 (UIDB/50006/2020 and UIDP/50006/2020). The authors would also like to thank to the EU and FCT / UEFISCDI



863 / FORMAS for funding in the frame of the collaborative international consortium REWATER, financed under  
864 the ERA-NET Cofund WaterWorks 2015 Call. This ERA-NET is an integral part of the 2016 Joint Activities  
865 developed by the Water Challenges for a Changing World Joint Programme Initiative (Water JPI). This work  
866 was also supported by the project Farmasense (39957), funded by Sistema de Incentivos à Investigação e  
867 Desenvolvimento Tecnológico de Portugal 2020, through the Programa Operacional do Norte (NORTE 2020)  
868 and the Fundo Europeu de Desenvolvimento Regional (FEDER). Isabel Seguro received her research Grant  
869 through this project. E. Costa-Rama thanks the Government of Principado de Asturias and Marie Curie-  
870 Cofund Actions for the post-doctoral grant “Clarín-Cofund” (ACA17-20). Patrícia Rebelo is grateful to FCT for  
871 her PhD grant (SFRH/BD/132384/2017). João Pacheco is grateful to FCT for his postdoc grant  
872 (SFRH/BPD/101419/2014), financed by POPH-QREN-Tipologia 4.1-Formação Avançada, funded by Fundo  
873 Social Europeu and MCTES.

## 874 **References**

- 875 Adeel, M., Song, X., Wang, Y., Francis, D., Yang, Y., 2017. *Environ. Int.* 99, 107–119.
- 876 Adumitrăchioaie, A., Tertiș, M., Cernat, A., Săndulescu, R., Cristea, C., 2018. *Int. J. Electrochem. Sci.* 13,  
877 2556–2576.
- 878 Afonso-Olivares, C., Sosa-Ferrera, Z., Santana-Rodríguez, J.J., 2017.. *Sci. Total Environ.* 599–600, 934–943.
- 879 Ahmad, O.S., Bedwell, T.S., Esen, C., Garcia-cruz, A., Piletsky, A., Le, L., 2019. *Trends Biotechnol.* 37, 294–  
880 309.
- 881 AL-Ammari, R.H., Ganash, A.A., Salam, M.A., 2019. *Synth. Met.* 254, 141–152.
- 882 Ali, Hazrat, Khan, E., Ilahi, I., 2019. *J. Chem.* 6730305.
- 883 Ali, Haydar, Mukhopadhyay, S., Jana, N.R., 2019. *New J. Chem.* 43, 1536–1543.
- 884 Alizadeh, T., 2014. *Biosens. Bioelectron.* 61, 532–540.
- 885 Alizadeh, T., 2009. *Electroanalysis* 21, 1490–1498.
- 886 Alizadeh, T., Zare, M., Ganjali, M.R., Norouzi, P., Tavana, B., 2010. *Biosens. Bioelectron.* 25, 1166–1172.
- 887 Alizadeha, T., Ganjali, M.R., Norouzi, P., Zare, M., Zeraatkar, A., 2009. *Talanta* 79, 1197–1203.
- 888 Ansari, S., 2017. *TrAC - Trends Anal. Chem.* 90, 89–106.
- 889 Ansari, S., Karimi, M., 2017a. *TrAC - Trends Anal. Chem.* 89, 146–162.
- 890 Ansari, S., Karimi, M., 2017b. *Talanta* 164, 612–625.
- 891 Archer, E., Petrie, B., Kasprzyk-Hordern, B., Wolfaardt, G.M., 2017. *Chemosphere* 174, 437–446.
- 892 Arismendi, D., Becerra-Herrera, M., Cerrato, I., Richter, P., 2019. *Talanta* 201, 480–489.
- 893 Arrigo, P., Baroni, D., 2020. *Sensors in Water Pollutants Monitoring: Role of Material Advanced Functional*  
894 *Materials and Sensors.* pp. 213–231.
- 895 Aswini, K.K., Mohan, A.M.V., Biju, V.M., 2014. *Mater. Sci. Eng. C* 37, 321–326.
- 896 Atar, N., Yola, M.L., 2018. *J. Electrochem. Soc.* 165, H255–H262.
- 897 Ayankojo, A.G., Reut, J., Ciocan, V., Öpik, A., Syritski, V., 2020. *Talanta* 209, 120502.
- 898 Ayukekbong, J.A., Ntemgwa, M., Atabe, A.N., 2017. *TAntimicrob. Resist. Infect. Control* 6, 1–8.
- 899 Beduk, T., Ait-Lahcen, A., Tashkandi, N., Salama, K.N., 2020. *Sensors Actuators B Chem.* 314, 128026.
- 900 Belbruno, J.J., 2019. *Chem. Rev.* 119, 94–119.
- 901 Beldean-Galea, M.S., Vial, J., Thiébaud, D., Coman, M.V., 2020. *Environ. Sci. Pollut. Res.* 9535–9546.
- 902 Beluomini, M.A., da Silva, J.L., de Sá, A.C., Buffon, E., Pereira, T.C., Stradiotto, N.R., 2019. *J. Electroanal.*  
903 *Chem.* 840, 343–366.
- 904 Benedetti, B., Majone, M., Cavaliere, C., Montone, C.M., Fatone, F., Frison, N., Laganà, A., Capriotti, A.L.,  
905 2020. *Microchem. J.* 155, 104732.
- 906 Bilal, M., Adeel, M., Rasheed, T., Zhao, Y., Iqbal, H.M.N., 2019. *Environ. Int.* 124, 336–353.
- 907 Björleinius, B., Tysklind, M., Lindberg, R.H., Haglund, P., Fick, J., Ripszám, M., 2018. *PSci. Total Environ.* 633,  
908 1496–1509.
- 909 Borghi, A.A., Palma, M.S.A., 2014. *Brazilian J. Pharm. Sci.* 50, 25–40.
- 910 Boxall, A.B.A., 2004. *EMBO Rep.* 5, 1110–1116.
- 911 Boyle, A., Geniès, E.M., Lapkowski, M., 1989. *Synth. Met.* 28, 769–774.
- 912 Boysen, R.I., 2019. *J. Sep. Sci.* 42, 51–71.
- 913 Cai, Y., He, X., Cui, P.L., Liu, J., Li, Z. Bin, Jia, B.J., Zhang, T., Wang, J.P., Yuan, W.Z., 2019. *Food Chem.* 280,



914 103–109.

915 Carvalho, F.P., 2017. *Food Energy Secur.* 6, 48–60.

916 Cetó, X., Saint, C.P., Chow, C.W.K., Voelcker, N.H., Prieto-Simón, B., 2016. *ESensors Actuators, B Chem.* 237,  
917 613–620.

918 Chakraborty, I., Sathe, S.M., Khuman, C.N., Ghangrekar, M.M., 2020. *Mater. Sci. Energy Technol.* 3, 104–  
919 115.

920 Chapman, P.M., 2007. *Environ. Int.* 33, 492–501.

921 Chen, H., Zhang, Z., Cai, R., Rao, W., Long, F., 2014. *Electrochim. Acta* 117, 385–392.  
922 <https://doi.org/10.1016/j.electacta.2013.11.185>

923 Chen, H.J., Zhang, Z.H., Cai, R., Chen, X., Liu, Y.N., Rao, W., Yao, S.Z., 2013. *Talanta* 115, 222–227.

924 Chen, L., Wang, X., Lu, W., Wu, X., Li, J., 2016. *Chem. Soc. Rev.* 45, 2137–2211.

925 Chen, L., Xu, S., Li, J., 2011. *Chem Soc Rev* 40, 2922–2942.

926 Chen, Z., Tang, C., Zeng, Y., Liu, H., Yin, Z., Li, L., 2014. *Anal. Lett.* 47, 996–1014.

927 Cieplak, M., Kutner, W., 2016. *A Trends Biotechnol.* 34, 922–941.

928 Crapnell, R.D., Hudson, A., Foster, C.W., Eersels, K., van Grinsven, B., Cleij, T.J., Banks, C.E., Peeters, M.,  
929 2019. *Sensors (Switzerland)* 19.

930 Dadkhah, S., Ziaei, E., Mehdinia, A., Baradaran Kayyal, T., Jabbari, A., 2016. *Microchim. Acta* 183, 1933–  
931 1941.

932 Daghbir, R., Drogui, P., 2013. *Environ. Chem. Lett.* 11, 209–227.

933 Daughton, C.G., 2016. *Sci. Total Environ.* 562, 391–426.

934 Deore, B., Chen, Z., Nagaoka, T., 1999. *Anal. Sci.* 15, 827–828.

935 Des Azevedo, S., Lakshmi, D., Chianella, I., Whitcombe, M.J., Karim, K., Ivanova-Mitseva, P.K.,  
936 Subrahmanyam, S., Piletsky, S.A., 2013. *Ind. Eng. Chem. Res.* 52, 13917–13923.

937 Di Masi, S., Pennetta, A., Guerreiro, A., Canfarotta, F., De Benedetto, G.E., Malitesta, C., 2020. *Sensors*  
938 *Actuators, B Chem.* 307, 127648.

939 Ding, S., Lyu, Z., Niu, X., Zhou, Y., Liu, D., Falahati, M., Du, D., Lin, Y., 2020. *Biosens. Bioelectron.* 149,  
940 111830.

941 Ding, Y., Zhang, X., Yin, H., Meng, Q., Zhao, Y., Liu, L., Wu, Z., Xu, H., 2017. *Sensors* 17.

942 Do, M.H., Florea, A., Farre, C., Bonhomme, A., Bessueille, F., Vocanson, F., Tran-Thi, N.T., Jaffrezic-Renault,  
943 N., 2015. *Int. J. Environ. Anal. Chem.* 95, 1489–1501.

944 Dong, S., Sun, Z., Lu, Z., 1988. *Analyst* 113, 1525–1528.

945 Dulio, V., Bavel, B. van, Brorström-Lundén, E., Harmsen, J., Hollender, J., Schlabach, M., Slobodnik, J.,  
946 Thomas, K., Koschorreck, J., 2018. *Environ. Sci. Eur.* 30.

947 Ertürk, G., Mattiasson, B., 2017. *Sensors (Switzerland)* 17, 1–17.

948 Evgenidou, E.N., Konstantinou, I.K., Lambropoulou, D.A., 2015. *Sci. Total Environ.* 505, 905–926.

949 Farooq, S., Nie, J., Cheng, Y., Yan, Z., Li, J., Bacha, S.A.S., Mushtaq, A., Zhang, H., 2018. *Analyst* 143, 3971–  
950 3989.

951 Feier, B., Blidar, A., Pusta, A., Carciuc, P., Cristea, C., 2019. *Biosensors* 9, 31.

952 Fekadu, S., Alemayehu, E., Dewil, R., Van der Bruggen, B., 2019. *Sci. Total Environ.* 654, 324–337.

953 Figueiredo, L., Erny, G.L., Santos, L., Alves, A., 2016. *Talanta* 146, 754–765.

954 Florea, A., Cristea, C., Vocanson, F., Săndulescu, R., Jaffrezic-Renault, N., 2015. *Electrochem. commun.* 59,  
955 36–39.

956 Futra, D., Heng, L.Y., Jaapar, M.Z., Ulianas, A., Saeedfar, K., Ling, T.L., 2016. *Anal. Methods* 8, 1381–1389.

957 García-Espinoza, J.D., Mijaylova Nacheva, P., 2019. *Environ. Sci. Pollut. Res.*

958 Geissen, V., Mol, H., Klumpp, E., Umlauf, G., Nadal, M., van der Ploeg, M., van de Zee, S.E.A.T.M., Ritsema,  
959 C.J., 2015. *Int. Soil Water Conserv. Res.* 3, 57–65.

960 Ghorbani, A., Ganjali, M.R., Ojani, R., Raoof, J., 2020. *Int. J. Electrochem. Sci.* 15, 2913–2922.

961 Grenni, P., Ancona, V., Barra Caracciolo, A., 2018. *Microchem. J.* 136, 25–39.

962 Gui, R., Guo, H., Jin, H., 2019. *Nanoscale Adv.* 1, 3325–3363.

963 Gui, R., Jin, H., Guo, H., Wang, Z., 2018. *Biosens. Bioelectron.* 100, 56–70.

964 Han, Q., Shen, X., Zhu, W., Zhu, C., Zhou, X., Jiang, H., 2016. *Biosens. Bioelectron.* 79, 180–186.

965 Han, S., Su, L., Zhai, M., Ma, L., Liu, S., Teng, Y., 2019. *J. Mater. Sci.* 54, 3331–3341.

966 Holdgate, M.W., 1979. *A perspective of environmental pollution.* Cambridge University Press.

967 Hong, Y.J., Hong, Y.J., Liao, W., Liao, W., Yan, Z.F., Bai, Y.C., Feng, C.L., Xu, Z.X., Xu, D.Y., 2020. *PJ. Chem.*  
968 2020, 1–12.  
969 <https://www.norman-network.net/?q=node/19> (Last accessed November 2019).  
970 Huang, S., Xu, J., Zheng, J., Zhu, F., Xie, L., Ouyang, G., 2018. Synthesis and application of magnetic  
971 molecularly imprinted polymers in sample preparation *Anal. Bioanal. Chem* 3991–4014.  
972 Hutchins, R.S., Bachas, L.G., 1995. *Anal. Chem.* 67, 1654–1660.  
973 Javid, A., Mesdaghinia, A., Nasser, S., Mahvi, A.H., Alimohammadi, M., Gharibi, H., 2016. *J. Environ. Heal.*  
974 *Sci. Eng.* 14, 1–5.  
975 Kamba, P.F., Kaggwa, B., Munanura, E.I., Okurut, T., Kitutu, F.E., 2017. *Pan Afr. Med. J.* 27, 2–5.  
976 Khadem, M., Faridbod, F., Norouzi, P., Foroushani, A.R., Ganjali, M.R., Shahtaheri, S.J., 2016. *J. Iran. Chem.*  
977 *Soc.* 13, 2077–2084.  
978 Khadem, M., Faridbod, F., Norouzi, P., Rahimi Foroushani, A., Ganjali, M.R., Shahtaheri, S.J., Yarahmadi, R.,  
979 2017. *Electroanalysis* 29, 708–715.  
980 Khan, S., Wong, A., Zanon, M.V.B., Sotomayor, M.D.P.T., 2019. *Mater. Sci. Eng. C* 103, 109825.  
981 Khanmohammadi, A., Jalili Ghazizadeh, A., Hashemi, P., Afkhami, A., Arduini, F., Bagheri, H., 2020. *J. Iran.*  
982 *Chem. Soc.* 17, 2429–2447.  
983 Kroon, F.J., Berry, K.L.E., Brinkman, D.L., Kookana, R., Leusch, F.D.L., Melvin, S.D., Neale, P.A., Negri, A.P.,  
984 Puotinen, M., Tsang, J.J., van de Merwe, J.P., Williams, M., 2020. *SSci. Total Environ.* 719, 135140.  
985 Kuhn, J., Aylaz, G., Sari, E., Marco, M., Yiu, H.H.P., Duman, M., 2020. *S J. Hazard. Mater.* 387, 121709.  
986 Kumar, V., Parihar, R.D., Sharma, A., Bakshi, P., Singh Sidhu, G.P., Bali, A.S., Karaouzas, I., Bhardwaj, R.,  
987 Thukral, A.K., Gyasi-Agyei, Y., Rodrigo-Comino, J., 2019. *Chemosphere* 236, 124364.  
988 Lahcen, A.A., Amine, A., 2019. *Electroanalysis* 31, 188–201.  
989 Lahcen, A.A., Baleg, A.A., Baker, P., Iwuoha, E., Amine, A., 2017. *Sensors Actuators, B Chem.* 241, 698–705.  
990 Lapworth, D.J., Baran, N., Stuart, M.E., Ward, R.S., 2012. *Environ. Pollut.* 163, 287–303.  
991 Li, C., Zeng, C., Chen, Z., Jiang, Y., Yao, H., Yang, Y., Wong, W.T., 2020. *J. Hazard. Mater.* 384, 121498.  
992 Li, H., Wang, Y., Zha, H., Dai, P., Xie, C., 2019. *Arab. J. Sci. Eng.* 44, 145–152.  
993 Li, J., Huang, X., Ma, J., Wei, S., Zhang, H., 2020. *Ionics* 26, 4183–4192  
994 Li, J., Ma, F., Wei, X., Fu, C., Pan, H., 2015. *Anal. Chim. Acta* 871, 51–56.  
995 Li, R., Feng, Y., Pan, G., Liu, L., 2019. *Sensors* 19.  
996 Li, S., Ge, Y., Piletsky, S.A., Lunec, J., 2012. *Molecularly imprinted sensors: overview and applications*, 1st  
997 editio. ed. Elsevier.  
998 Li, S., Li, J., Ma, X., Pang, C., Yin, G., Luo, J., 2019. *Biosens. Bioelectron.* 139, 111321.  
999 Li, Y., Xu, W., Zhao, X., Huang, Y., Kang, J., Qi, Q., Zhong, C., 2018. *Analyst* 143, 5094–5102.  
1000 Li, Y., Zhang, L., Dang, Y., Chen, Z., Zhang, R., Li, Yingchun, Ye, B.C., 2019. *Biosens. Bioelectron.* 127, 207–  
1001 214. h  
1002 Liang, M., Xian, Y., Wang, B., Hou, X., Wang, L., Guo, X., Wu, Y., Dong, H., 2020. *Environ. Pollut.* 263,  
1003 114389.  
1004 Liang, Y., Yu, L., Yang, R., Li, X., Qu, L., Li, J., 2017. *Sensors Actuators, B Chem.* 240, 1330–1335.  
1005 Liu, G., Huang, X., Li, L., Xu, X., Zhang, Y., Lv, J., Xu, D., 2019. *Nanomaterials* 9, 1–19.  
1006 Liu, L., Zhu, X., Zeng, Y., Wang, H., Lu, Y., Zhang, J., Yin, Z., Chen, Z., Yang, Y., Li, L., 2018. *Polymers.* 10, 1–13.  
1007 Liu, Y., Zhu, L., Luo, Z., Tang, H., 2013. *Sensors Actuators, B Chem.* 185, 438–444.  
1008 Lopes, F., Pacheco, J.G., Rebelo, P., Delerue-Matos, C., 2017. *Sensors Actuators, B Chem.* 243, 745–752.  
1009 Luliński, P., 2017. *Mater. Sci. Eng. C* 76, 1344–1353.  
1010 Luo, Y., Guo, W., Ngo, H.H., Nghiem, L.D., Hai, F.I., Zhang, J., Liang, S., Wang, X.C., 2014. *Sci. Total Environ.*  
1011 473–474, 619–641.  
1012 Luong, J.H.T., Male, K.B., Glennon, J.D., 2009. *Analyst* 134, 1965–1979.  
1013 Madikizela, L., Tavengwa, N., Pakade, V., 2018. *Recent Res. Polym.*  
1014 Madikizela, L.M., Tavengwa, N.T., Chimuka, L., 2018. *J. Pharm. Biomed. Anal.* 147, 624–633.  
1015 Maduraiveeran, G., Sasidharan, M., Ganesan, V., 2018. *Biosens. Bioelectron.* 103, 113–129.  
1016 Malitesta, C., Losito, I., Zamboni, P.G., 1999. *Anal. Chem.* 71, 1366–1370.  
1017 Manisalidis, I., Stavropoulou, E., Stavropoulos, A., Bezirtzoglou, E., 2020. *Front. Public Heal.* 8, 1–13.  
1018 Maria G.C., A., K B, A., Rison, S., Varghese, A., George, L., 2020. *Synth. Met.* 261, 116309.  
1019 Mayes, A.G., Whitcombe, M.J., 2005. *Adv. Drug Deliv. Rev.* 57, 1742–1778.

1020 Mercurio, P., Flores, F., Mueller, J.F., Carter, S., Negri, A.P., 2014. *Mar. Pollut. Bull.* 85, 385–390.

1021 Merlo, F., Speltini, A., Maraschi, F., Sturini, M., Profumo, A., 2020. *Arab. J. Chem.* 13, 4673–4680. h

1022 Miege, C., Choubert, J.M., Ribeiro, L., Eusèbe, M., Coquery, M., 2009. *Environ. Pollut.* 157, 1721–1726.

1023 Moein, M.M., Abdel-Rehim, A., Abdel-Rehim, M., 2019. *Recent Applications of Molecularly Imprinted.*

1024 *Molecules* 24, 1–12.

1025 Mokhtari, P., Ghaedi, M., 2019. *Eur. Polym. J.* 118, 614–618.

1026 Motaharian, A., Motaharian, F., Abnous, K., Hosseini, M.R.M., Hassanzadeh-Khayyat, M., 2016. *Anal.*

1027 *Bioanal. Chem.* 408, 6769–6779.

1028 Motia, S., Albert, I., Popescu, L.M., Mioara, R., Bouchikhi, B., El, N., 2018. *J. Electroanal. Chem.* 823, 553–

1029 562.

1030 Motia, S., Tudor, I.A., Ribeiro, P.A., Raposo, M., Bouchikhi, B., El Bari, N., 2019. *Sci. Total Environ.* 664, 647–

1031 658.

1032 Mujahid, A., Lieberzeit, P.A., Dickert, F.L., 2010. *Materials.* 3, 2196–2217.

1033 Mzukisi, L., Tawanda, N., Tutu, H., Chimuka, L., Madikizela, L.M., Tavengwa, N.T., Tutu, H., Chimuka, L.,

1034 2018. *Anal. Chem.* 17, 14–22. h

1035 Naidu, R., Arias Espana, V.A., Liu, Y., Jit, J., 2016. *Chemosphere* 154, 350–357.

1036 Namieśnik, J., 2000. *Crit. Rev. Anal. Chem.* 30, 221–269.

1037 Nsibandé, S.A., Forbes, P.B.C., 2019. *Luminiscence* 480–488.

1038 Özcan, A., Topçuoğulları, D., 2017. *Sensors Actuators, B Chem.* 250, 85–90.

1039 Pacheco, J.G., Rebelo, P., Freitas, M., Nouws, H.P.A., Delerue-Matos, C., 2018. *Sensors Actuators, B Chem.*

1040 273, 1008–1014.

1041 Paíga, P., Correia, M., Fernandes, M.J., Silva, A., Carvalho, M., Vieira, J., Jorge, S., Silva, J.G., Freire, C.,

1042 Delerue-Matos, C., 2019. *Sci. Total Environ.* 648, 582–600.

1043 Paíga, P., Santos, L.H.M.L.M., Ramos, S., Jorge, S., Silva, J.G., Delerue-Matos, C., 2016. *Sci. Total Environ.*

1044 573, 164–177.

1045 Patel, M., Kumar, R., Kishor, K., Mlsna, T., Pittman, C.U., Mohan, D., 2019. *Chem. Rev.* 119, 3510–3673.

1046 Petrie, B., Barden, R., Kasprzyk-Hordern, B., 2014. *Water Res.* 72, 3–27.

1047 Pohanka, M., 2017. *Int. J. Electrochem. Sci.* 12, 8082–8094.

1048 Prasad, B.B., Fatma, S., 2016. *Sensors Actuators, B Chem.* 229, 655–663.

1049 Prasad, B.B., Jauhari, D., Tiwari, M.P., 2014. *Biosens. Bioelectron.* 59, 81–88.

1050 Prasad, B.B., Singh, K., 2016. *Electrochim. Acta* 187, 193–203.

1051 Rahim, S., Bhayo, A.M., Shah, M.R., Malik, M.I., 2019. *Microchem. J.* 149, 104048.

1052 Ramírez-Malule, H., Quiñones-Murillo, D.H., Manotas-Duque, D., 2020. *Emerg. Contam.* 6, 179–193.

1053 Rao, H., Liu, X., Ding, F., Wan, Y., Zhao, X., Liang, R., Zou, P., Wang, Y., Wang, X., Zhao, Q., 2018. *Chem. Eng.*

1054 *J.* 338, 478–487.

1055 Rasheed, T., Bilal, M., Nabeel, F., Adeel, M., Iqbal, H.M.N., 2019. *Environ. Int.* 122, 52–66.

1056 Rebelo, P., Pacheco, J.G., Cordeiro, M.N.D.S., Melo, A., Delerue-Matos, C., 2020. *Anal. Methods* 12.

1057 Ribeiro, A.R., Sures, B., Schmidt, T.C., 2018. *Environ. Pollut.* 241, 1153–1166.

1058 Richardson, S.D., Ternes, T.A., 2014. *Anal. Chem.* 86, 2813–2848.

1059 Rivera-Jaimes, J.A., López de Alda, M., Aceña, J., Melgoza-Alemán, R.M., Postigo, C., Barceló, D., 2018. *Sci.*

1060 *Total Environ.* 613–614, 1263–1274.

1061 Rousis, N.I., Bade, R., Bijlsma, L., Zuccato, E., Sancho, J. V., Hernandez, F., Castiglioni, S., 2017. *Environ. Res.*

1062 156, 31–38.

1063 Sadki, S., Schottland, P., Brodie, N., Sabouraud, G., 2000. *Chem. Soc. Rev.* 29, 283–293.

1064 Sadriu, I., Bouden, S., Nicolle, J., Podvorica, F.I., Bertagna, V., Berho, C., Amalric, L., Vautrin-UI, C., 2020.

1065 *Talanta* 207, 120222.

1066 Salste, L., Leskinen, P., Virta, M., Kronberg, L., 2007. *Sci. Total Environ.* 378, 343–351.

1067 Sarafraz-Yazdi, A., Amiri, A., Rounaghi, G., Eshtiagh-Hosseini, H., 2012. *Anal. Chim. Acta* 720, 134–141.

1068 Sarpong, K.A., Zhang, K., Luan, Y., Cao, Y., Xu, W., 2020. *Microchem. J.* 154, 104526.

1069 Sayyahmanesh, M., Asgari, S., Meibodi, A.S.E., Ahooyi, T.M., 2016. *arXiv:1604.07853*.

1070 Shaheen, A., Munawar, A., Tahir, M.A., Khan, W.S., Lieberzeit, P.A., Bajwa, S.Z., 2017. *J. Hazard. Mater.* 342,

1071 96–106.

1072 Sharma, G., Kandasubramanian, B., 2020. *J. Chem. Eng. Data* 65, 396–418.

1073 Sharma, P.S., Pietrzyk-Le, A., D'Souza, F., Kutner, W., 2012. *Anal. Bioanal. Chem.* 402, 3177–3204.

1074 Shi, T., Tan, L., Fu, H., Wang, J., 2019. *Mar. Pollut. Bull.* 146, 591–597.

1075 Shirzadmehr, A., Rezaei, M., Bagheri, H., Khoshshafar, H., 2016. *Anal. Chem.* 96, 929–944.

1076 Silva, V., Montanarella, L., Jones, A., Fernández-Ugalde, O., Mol, H.G.J., Ritsema, C.J., Geissen, V., 2018. *Sci. Total Environ.* 621, 1352–1359.

1077 Sobiech, M., Bujak, P., Lulinski, P., Pron, A., 2019. *Nanoscale* 11, 12030–12074.

1078 Spietelun, A., Marcinkowski, Ł., de la Guardia, M., Namieśnik, J., 2013. *J. Chromatogr. A* 1321, 1–13.

1079 Sui, Q., Cao, X., Lu, S., Zhao, W., Qiu, Z., Yu, G., 2015. *Emerg. Contam.* 1, 14–24.

1080 Surya, S.G., Khatoun, S., Ait Lahcen, A., Nguyen, A.T.H., Dzantiev, B.B., Tarannum, N., Salama, K.N., 2020. *RSC Adv.* 10, 12823–12832.

1081 Tadi, K.K., Motghare, R. V., Ganesh, V., 2014. *Electroanalysis* 26, 2328–2336.

1082 Tan, F., Cong, L., Li, X., Zhao, Q., Zhao, H., Quan, X., Chen, J., 2016. *Sensors Actuators, B Chem.* 233, 599–606.

1083 Tao, Q., Li, B., Li, Q., Han, X., Jiang, Y., Jupa, R., Wang, C., Li, T., 2019. *Sci. Total Environ.* 659, 1448–1456.

1084 Toro, M.J.U., Marestoni, L.D., Del Pilar Taboada Sotomayor, M., 2015. *Sensors Actuators, B Chem.* 208, 299–306. h

1085 Uzun, L., Turner, A.P.F., 2016. *Biosens. Bioelectron.* 76, 131–144.

1086 Van, T.T.H., Yidana, Z., Smooker, P.M., Coloe, P.J., 2020. *AJ. Glob. Antimicrob. Resist.* 20, 170–177.

1087 Vinokurov, I.A., 1992. *Sensors Actuators B. Chem.* 10, 31–35.

1088 Vytřas, K., Švancara, I., Metelka, R., 2009. *J. Serbian Chem. Soc.* 74, 1021–1033.

1089 Wackerlig, J., Schirhagl, R., 2016. *Anal. Chem.* 88, 250 - 261

1090 Wang, X., Ding, H., Yu, X., Shi, X., Sun, A., Li, D., Zhao, J., 2019. *Talanta* 197, 98–104.

1091 Wang, Y., Yang, Y., Xu, L., Zhang, J., 2011. *Bisphenol Electrochim. Acta* 56, 2105–2109.

1092 Wong, A., Foguel, M.V., Khan, S., Oliveira, F.M. De, Tarley, C.R.T., Sotomayor, M.D.P.T., 2015. *Electrochim. Acta* 182, 122–130.

1093 Wu, B., Hou, L., Du, M., Zhang, T., Xue, Z., Lu, X., 2014. *RSC Adv.* 4, 53701–53710.

1094 Wulff, G., Sarhan, A., 1972. *TAngew. Chem. Int. Ed. Engl.* 11, 341–343.

1095 Xiao, D., Jiang, Y., Bi, Y., 2018. *Microchim Acta* 185

1096 Xiao, L., Zhang, Z., Wu, C., Han, L., Zhang, H., 2017. *Food Chem.* 221, 82–86.

1097 Xie, C., Gao, S., Guo, Q., Xu, K., 2010a. *Microchim. Acta* 169, 145–152.

1098 Xie, C., Li, H., Li, S., Wu, J., Zhang, Z., 2010b. *Anal. Chem.* 82, 241–249.

1099 Xie, L., Xiao, N., Li, L., Xie, X., Li, Y., 2020. *Int. J. Mol. Sci.* 21, 4139.

1100 Xu, J., Zhang, R., Liu, C., Sun, A., Chen, J., Zhang, Z., Shi, X., 2020. *Sensors* 20, 884.

1101 Xu, L., Li, J., Zhang, J., Sun, J., Gan, T., Liu, Y., 2020. *Analyst.*

1102 Xu, W., Wang, Q., Huang, W., Yang, W., 2017. *J. Sep. Sci.* 40, 4839–4846.

1103 Xue, X., Wei, Q., Wu, D., Li, H., Zhang, Y., Feng, R., Du, B., 2014. *Electrochim. Acta* 116, 366–371.

1104 Yang, S., Wang, Y., Jiang, Y., Li, S., Liu, W., 2016. *Polymers (Basel).* 8.

1105 Yang, Y., Fang, G., Liu, G., Pan, M., Wang, X., Kong, L., He, X., Wang, S., 2013. *EBiosens. Bioelectron.* 47, 475–481.

1106 Yang, Y., Ok, Y.S., Kim, K.H., Kwon, E.E., Tsang, Y.F., 2017. *Sci. Total Environ.* 596–597, 303–320.

1107 Yaroshenko, I., Kirsanov, D., Marjanovic, M., Lieberzeit, P.A., Korostynska, O., Mason, A., Frau, I., Legin, A., 2020. *Sensors* 20, 1–23.

1108 Ye, L., 2013. *Molecular Imprinting: Principles and Applications of Micro- and Nanostructure Polymers.* CRC Press: Boca Raton, FL, USA.

1109 Yola, M.L., Atar, N., 2017. *J. Electrochem. Soc.* 164, B223–B229.

1110 Yu, R., Zhou, H., Li, M., Song, Q., 2019. *J. Electroanal. Chem.* 832, 129–136.

1111 Yuan, L., Zhang, J., Zhou, P., Chen, J., Wang, R., Wen, T., Li, Y., Zhou, X., Jiang, H., 2011. *Biosens. Bioelectron.* 29, 29–33.

1112 Zaidi, S.A., 2017. *Biomater. Sci.* 5, 388–402.

1113 Zamora-Gálvez, A., Ait-Lahcen, A., Mercante, L.A., Morales-Narváez, E., Amine, A., Merkoçi, A., 2016. *Anal. Chem.* 88, 3578–3584.

1114 Zamora-Gálvez, A., Mayorga-Matinez, C.C., Parolo, C., Pons, J., Merkoçi, A., 2017. *Electrochem. commun.* 82, 6–11.

1115

- 1126 Zhang, B., Fan, X., Zhao, D., 2019. *Polymers (Basel)*. 11, 1–21.
- 1127 Zhang, C., She, Y., Li, T., Zhao, F., Jin, M., Guo, Y., Zheng, L., Wang, S., Jin, F., Shao, H., Liu, H., Wang, J., 2017.  
1128 *Anal. Bioanal. Chem.* 409, 7133–7144.
- 1129 Zhang, J., Lei, J., Ju, H., Wang, C., 2013. *Anal. Chim. Acta* 786, 16–21.
- 1130 Zhang, R., Zhang, G., Zheng, Q., Tang, J., Chen, Y., Xu, W., Zou, Y., Chen, X., 2012. *Ecotoxicol. Environ. Saf.*  
1131 80, 208–215.
- 1132 Zhang, R.R., Zhan, J., Xu, J.J., Chai, J.Y., Zhang, Z.M., Sun, A.L., Chen, J., Shi, X.Z., 2020. *Sensors Actuators, B*  
1133 *Chem.* 317, 128237.
- 1134 Zhao, H., Chen, Y., Tian, J., Yu, H., Quan, X., 2012. *J. Electrochem. Soc.* 159, J231–J236.
- 1135 Zhao, Y., Yuan, F., Quan, X., Yu, H., Chen, S., Zhao, H., Liu, Z., Hilal, N., 2015. *Anal. Methods* 7, 2693–2698.
- 1136 Zhou, T., Ding, L., Che, G., Jiang, W., Sang, L., 2019. *Trends Anal. Chem.* 114, 11–28.
- 1137 Zhou, T., Feng, Y., Zhou, L., Tao, Y., Luo, D., Jing, T., Shen, X., Zhou, Y., Mei, S., 2016. *Sensors Actuators, B*  
1138 *Chem.* 236, 153–162.
- 1139 Zhou, T., Halder, A., 2018. *Biosensors*
- 1140 Zhu, L., Cao, Y., Cao, G., 2014. *Biosens. Bioelectron.* 54, 258–261.
- 1141 Zulkifli, S.N., Rahim, H.A., Lau, W.J., 2018. *Sensors Actuators, B Chem.* 255, 2657–2689.
- 1142