



Unraveling the complex habitat use of white mullet *Mugil curema* of several environments from Neotropical Pacific and Atlantic waters

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Abstract

The white mullet *Mugil curema* is a widely distributed euryhaline species, whose migratory behavior is poorly understood. The objective of this work was to study large-scale habitat use of this species for the first time, considering several environments such as euryhaline and hypersaline lagoons, sea, and a river, distributed in the central Pacific (México) and Atlantic (Gulf of México, Caribbean Sea-Venezuela, and Northeast of Brazil). Otolith core-to-edge Sr/Ca ratios of 161 fish, determined by laser ablation inductively coupled plasma mass spectrometry, were used to study the salinity-habitat migration history. Fish from México (Tamiahua Lagoon, N=4; Alvarado Lagoon, N=2), Venezuela (N=1), and Brazil (N=10) (10.6% of the total) showed high Sr/Ca values at the beginning of the transects and were classified as marine migrants. Two specimens (Alvarado Lagoon and Balsas River, México) showed Sr/Ca values consistently below high salinity guide value (salinity<33.5). In the rest of the fish (88.1%), the Sr/Ca suggested a displacement from the estuary towards the sea or hypersaline environments, so they were classified as estuarine migrants. A change point analysis identified six individuals with a single stable Sr/Ca signature (SSS) through ontogeny (3 for Brazil, 1 for Venezuela, and 2 for Tamiahua Lagoon), suggesting limited displacement between environments with different salinity. The rest of the individuals showed between 2 and 10 SSSs (mean=4.07±1.85). The highest number of SSS (4.87±1.1) was found in Laguna Madre Fish (México) and the lowest in Brazil (3.27±1.70) (H=19.8, p=0.002). Otolith Sr/Ca time-series suggested a highly plastic migration behavior for *M. curema*, where the marine and estuarine migratory were the most common ones. This work revealed that the sustainable use of *M. curema* depends on the conservation of the corridors between river, estuary, mangrove and sea.

Keywords: catadromous, diadromous, LA-ICP-MS, life history, migration, mugilidae, otolith microchemistry,

1. Introduction

The study of life history in diadromous fish is crucial for the understanding habitat use and migration behavior, so as to generate proper conservation and management regulations for species and used areas (Beck et al., 2001; Jenkins et al., 2010; Wynne, Wilson & Limburg, 2015). Different methods have been applied to study movements and habitat selection of fish such as mark-recapture, and the chemical composition of calcified structures like otolith, spines and scales (Avigliano et al., 2017; Clarke, Telmer & Mark Shrimpton, 2007; Clément et al., 2014; Raabe & Gardner, 2013). In particular, the study of features in otoliths has facilitated not only the study of fish movements and migrations but also the stock identification of important commercial species (Biolé et al., 2019; Lemos et al., 2017; Soeth et al., 2019). Elemental deposition in the otolith is influenced by physiological and environmental factors, most particularly by the concentration of elements in the surrounding water (Avigliano, et al., 2019; Thomas & Swearer, 2019). The chemicals deposited represent a permanent record of the environmental conditions experienced by the fish at a particular time; therefore, they can be used to reconstruct their environmental migratory patterns and habitat use (Campana, et al., 2000). Different elemental ratios have been used to study displacements between environments with different salinities, being Sr/Ca the most used (Avigliano et al., 2017; Avigliano, Miller & Volpedo, 2018; Elsdon & Gillanders, 2005). Specially in diadromous fishes, the Sr/Ca ratio in otolith and water has been shown to be positively correlated to the salinity, thus having a strong association to marine environments (Avigliano, & Volpedo, 2013; Kraus & Secor, 2004; Tabouret et al., 2010). As a result, otolith Sr/Ca ratio has been used in a large number of studies worldwide to reveal movements of diadromous species through salinity gradients (Arai & Chino, 2017; Avigliano et al., 2018; Brown, & Severin, 2009; Chang, Lin, et al., 2004).

The white mullet *Mugil curema* Valenciennes 1836, is a widely distributed species from the Mugilidae family that inhabits the Atlantic Ocean (from Nova Scotia to Argentina in the east, from Gambia to the Congo in the west coast of Africa), and the eastern Pacific Ocean (from the Gulf of

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3 69 California to Northern Chile) (Crosetti et al. 2016; Froese and Pauly 2019). This euryhaline
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5 70 mugilidae species is thought to spawn offshore and its larvae to migrate from the sea to estuarine
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8 71 water, especially in mangrove habitat, until reaching sexual maturity (Barletta & Dantas, 2016; da
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10 72 Silva et al., 2018). Some authors have studied this species because of its important economic value as
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12 73 a commercial species and in aquaculture, from Brazil all through the Caribbean (Ibañez-Aguirre
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14 74 1996; Marin et al. 2003; Ibañez et al. 2012; Avigliano et al. 2015; Ibañez et al. 2017; Mai *et al.*,
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16 75 2018; Santana *et al.*, 2018). Santana *et al.* (2018) have found a positive relationship between otolith
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18 76 Sr/Ca and salinity for *M. curema* and have studied the life history of fish from a small estuary in
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20 77 northeast Brazil. Moreover, Ibañez et al. (2012) have revealed migratory patterns of *M. curema* in
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22 78 three sampling sites from the Mexican Atlantic coast using Sr/Ca, while Mai et al. (2018) have used
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24 79 Ba/Ca to study displacements in southeast Brazil. These studies reported some plasticity in the
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26 80 previously known life history migratory behavior of the species. However, they have focused on few
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28 81 specimens (Mai *et al.*, 2018, N=32; Ibañez *et al.*, 2012, N=40; Santana *et al.*, 2018, N=23) and local
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30 82 estuarine systems, and there is no previous evidence of the habitat use through ontogeny in
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32 83 hypersaline, Pacific ocean or Caribbean sea environments; therefore, migratory patterns are not fully
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34 84 understood. This information is necessary because the first step to generate conservation and
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36 85 management strategies at the ecoregional-scale is to know the use of the habitat and identify the sites
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38 86 that must be managed (Beck & Odaya, 2001).

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40 87 The objective of this study was to unravel for the first time the habitat use of *M. curema* from a wide
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42 88 range of environments and distribution in Pacific and Atlantic waters using otolith core-to-edge
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44 89 Sr/Ca time-series. For this purpose, otolith core-to-edge Sr/Ca time-series were analyzed in fish of
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46 90 euryaline and hypersaline lagoons, beaches, and a river from the Gulf of México (México),
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48 91 Caribbean Sea (Venezuela), neotropical Atlantic (Brazil) and neotropical Pacific (México) coasts.
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57 93 **2. Materials and methods**

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2.1. Sample collection and preparation

Adult *M. curema* (N=161, Table 1), were collected between November 2009 and July 2011 using trammel nets from the Gulf of México, Pacific coast, and Caribbean Sea, and gill nets from Southwestern Atlantic (Figure 1). Fish were caught from Mecoacan, Alvarado, Tamiahua and Laguna Madre lagoons (Gulf of México, México, Atlantic Ocean), Balsas River (México, Pacific Ocean), Nueva Esparta State (Margarita Island, Venezuela, Caribbean Sea), and Northeast coast of Brazil (Alagoas state, Atlantic Ocean) (Table 1). The Balsas River is the major river from south-central México and drains at an average height of 1,000 m.a.s.l. and covers a total area of 117,400 km². Laguna Madre is a long, shallow, and hypersaline lagoon (salinity up to 63.0), while Mecoacán and Alvarado are shallow coastal lagoon systems dominated by mangroves (Contreras & Castañeda, 2004). The Tamiahua Lagoon is a polyhaline water body surrounded by mangroves (Contreras & Castañeda, 2004). The Margarita Island and Brazil sampling sites correspond to sandy coastal beaches with great oceanic influence (Astor & Cárdenas, 1997). The features of the sampled environments are shown in Table 1.

The mullets were identified with the taxonomic keys of Harrison (2002) and Harrison et al. (2007), and using microsatellites markers (Pacheco-Almanzar, Loza-Estrada & Ibáñez, 2020). Fish were measured, and the *sagittae* otoliths were extracted. Right otoliths were weighted, decontaminated with 2% HNO₃, rinsed three times with ultrapure water (resistivity of 18 MΩ/cm), and embedded in crystal epoxy resin. Samples were sectioned transversely through the core using a Buehler Isomet low speed saw (Hong Kong, China). To reduce the effect of growth on the interpretations, samples were selected according to size and similar age (Table 1). Age was estimated using the annual growth ring count method, observing the sections of the otoliths immersed in ultrapure water. The age validation by the ring count in otoliths of *M. curema* from the Caribbean Sea was validated by Ibáñez-Aguirre & Gallardo-Cabello (1996). The selected samples were fixed to glass slides with

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3 118 epoxy resin, individually polished using a decreasing gradient of sandpaper (from 9 to 3 μm -grit),
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6 119 rinsed with ultrapure water, and sonicated for 10 min (Avigliano, et al., 2019).
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9 120 10 11 12 121 **2.2. Chemical analysis**

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14 122 The elements ^{43}Ca , and ^{88}Sr were quantified from core to edge in scan mode (Figure 2) by Laser
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16 123 Ablation Inductively Coupled Plasma Mass Spectrometry (LA-ICP-MS) using a 193 nm ArF
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19 124 Excimer laser (Photon Machines Analyte G2, USA) coupled to an ICP-QMS Agilent 7700 (Japan).
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22 125 Samples were measured during three analytical sessions using a laser-induced spot size of $\varnothing 40 \mu\text{m}$
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24 126 at 5 $\mu\text{m}/\text{s}$, at a laser fluence of 3.75 J/cm^2 and a repetition rate of 10 Hz. Helium was used as carrier
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26 127 gas (0.8 l/min) in the ablation cell, and argon was also added (0.9 l/min) before entering the ICP
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29 128 which was operated at 1600 W. The $^{238}\text{U}/^{232}\text{Th}$ (~ 1.2) and $^{232}\text{Th}^{16}\text{O}/^{232}\text{Th}$ ($< 0.4 \%$) ratios were
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31 129 monitored on the reference material NIST 612 (trace elements in glass, National Institute of
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33 130 Standards and Technology, NIST, USA) to check plasma robustness and oxide production rates,
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35
36 131 respectively. The standard reference materials NIST612, NIST610 (trace elements in silicate glass)
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38 132 and MACS-3 (trace elements in synthetic calcium carbonate, USGS, USA), were run in triplicate at
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40 133 the beginning and at the end of the each analytical session (in triplicate) and also every 10 transects
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43 134 for drift monitoring. To convert elemental signals to concentrations, NIST612 and Ca (38.3% weight,
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45 135 Yoshinaga *et al.*, 2000) were used as calibration and internal standards, respectively. Strontium
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47 136 recovery rates base on NIST610 and MACS-3 (Jochum et al., 2011; Pearce et al., 1997) ranged from
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50 137 101 to 105%. The relative standard deviation percentage of quadruplicate analysis of NIST610 and
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52 138 MACS-3 was below 3.5%. Strontium concentrations were expressed in relation to Ca (mmol/mol).
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58 140 **2.3. Data analysis and interpretation**

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3 141 Migratory patterns were described according to the use of environments with different salinities
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6 142 using the otolith Sr/Ca as a salinity proxy, as suggested by Ibáñez *et al.* (2012) and Santana *et al.*
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8 143 (2018). The positive relationships among *M. curema* otolith Sr/Ca and salinity were previously
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10 144 validated by Santana *et al.* (2018). Due to the lack of relationship between Ba/Ca and salinity found
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13 145 in pilot analysis, only the Sr/Ca ratio was used as a salinity proxy.

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16 146 To assess if the changes among Sr/Ca signatures through ontogeny were significant, Change-Point
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18 147 analyzes (CPA) were performed (Avigliano *et al.*, 2017, 2018). This analysis uses a combination of
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20 148 cumulative sum charts and bootstrapping to identify significant changes in stable Sr/Ca signatures
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23 149 (SSS) across chemical time-series. The analyses were performed using 95% confidence and 95%
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25 150 confidence levels. To assess spatial differences in the number of stable signatures experienced
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27 151 throughout life, SSS was compared between sampling sites using Kruskal Wallis test. Because
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30 152 potential differences in growth between individuals could be associated with the number of SSS thus
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32 153 affecting Kruskal Wallis' results, Spearman's correlation tests were performed to assess the
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34 154 relationship of SSS with fish size and age.

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37 155 Several authors have used transition Sr/Ca thresholds, based on experimental or field observations, to
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40 156 identify changes between freshwater, estuarine, and marine habitats (Avigliano *et al.*, 2017; Fowler
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42 157 *et al.*, 2016; Wang, 2014). Santana *et al.* (2018) have proposed transition thresholds for estuarine use
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44 158 based on the otolith Sr/Ca ratio of the early stages from wild adult *M. curema* (5×10^{-3} - 7.4×10^{-3}
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46 159 ppm/ppm, ~ 2.29 - 3.38 mmol/mol). However, the authors showed no direct evidence that assures the
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49 160 estuarine location in the early stage of these fish, and the estuarine salinities are not defined either. In
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51 161 this sense, it is not clear what the range of salinities represented by that threshold is. For this reason,
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53 162 these environmental transitional thresholds were not used. Instead, the mean Sr/Ca \pm twice standard
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56 163 deviation (mean \pm 2SD) of the otolith edge, which represents approximately the catch environment,
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58 164 from fish caught at Brazilian sea (salinity=33.5, N=32) was plotted with the core-to-edge Sr/Ca
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60 165 series as an approximate guide value for high salinity use (GHS). This approach was previously

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3 166 employed for several diadromous species such as *Genidens barbatus* (Avigliano et al., 2017), *Anguilla*
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5 167 *anguilla* (Tabouret et al., 2010), *A. mossambica* (Lin et al., 2014), among others.

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9 168 Fish were classified as marine migrant, estuarine migrant and estuarine resident according to Elliott
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11 169 *et al.* (2007). Use of the high salinity environments (salinity \geq 33.5, for example open sea and
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13 170 hypersaline lagoons) was considered as when the stable signatures of Sr/Ca were within or above the
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16 171 GHS. Therefore, marine migrants are fish that spawn at sea and often enter estuaries, estuarine
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18 172 migrant fish have larval stages in the estuary and often move to sea, and estuarine resident fish
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20 173 complete the entire life cycle within the estuarine environment (Elliott et al., 2007).

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23 174 To know the position of the rings along the ablation transect, the otolith sections were immersed in
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26 175 water and photographed after ablation analysis to observe the ablation line. The position of the rings
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28 176 was determined on the images using the Image-Pro Plus 4.5 software.

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31 177 Finally, core-to-edge Sr/Ca transects, GHS and CPA results were plotted together in order to
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33 178 facilitate the interpretation of fish movements through their life time.

39 180 **3. Results**

41 42 181 **3.1. Otolith edge**

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44 182 The mean Sr/Ca \pm SD based on otolith edge from fish caught at sea (N=32) was 4.01 \pm 0.32 mmol/mol,
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46 183 then the GHS (mean \pm 2SD) range was 3.46-4.75 mmol/mol. All Sr/Ca values of the otolith edge from
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48 184 fish caught at salinity 33.5 were included within that range.

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52 185 Core-to-edge Sr/Ca ratio ranged from 1.37 to 8.40 mmol/mol, where lowest levels were observed in
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54 186 Alvarado and Balsas River (e.g. Figure 3a and c), while the highest in Laguna Madre (e.g. Figure 4i
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56 187 and j).

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3 188 Only two specimens (Alvarado Lagoon and Balsas River) showed Sr/Ca values consistently below
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6 189 GHS (Figure 3a and b), representing the 5% of the total for these sampling sites. These individuals
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8 190 were considered estuarine resident (Figure 1).
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11 191 Around 10.6% of all individuals (N=17) showed Sr/Ca values above GHS at the beginning of the
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13 192 transects, being after both below and within (or even above) the GHS (Figure 3c-h) and so classified
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16 193 as marine migrants. These specimens were caught from Tamiahua (N=4), Alvarado (N=2), Margarita
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18 194 Island (N=1), and Brazil (N=10), representing the 20.0%, 10.5%, 9.5%, 18.5%, respectively, of total
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20 195 of each site.
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23 196 The rest of the specimens (N=141, 88.1% of the total) showed values below GHS at the beginning of
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25 197 the life (Figure 4), and after that, presented values within or above GHS range, suggesting a shift
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27 198 from relatively lower to higher salinities; therefore, they were classified as estuarine migrants. The
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29 199 spatial distribution of estuarine migrant individuals was 100% for Laguna Madre, 80% for Tamiahua,
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32 200 84.5% for Alvarado, 100% for Mecoacán, 95% for Balsas River, 90.5% for Margarita Island, and
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34 201 81.5% for Brazil (Figure 1). With these specimens, the individuals with relatively low chemical
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36 202 signatures in the otolith core reached 89.4% of the total sample (estuarine migrant plus estuarine
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39 203 resident).
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41 204 Regarding age, it was observed that the first movement between high (GHS) and moderate (e.g.
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43 205 estuary) salinity environments of estuarine and marine migrant fish occurred within the first year of
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46 206 life (except for one specimen, Figure 4b), after which no apparent migration patterns were observed
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48 207 in relation to age.
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50 208 According to the correlation analyzes, no significant relationship was found between the SSS
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52 209 ($r=0.07$, $p=0.7$) and age or total size ($r=0.2$, $p=0.6$). With a confidence interval of 95%, the CPA
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55 210 identified six individuals with a single stable Sr/Ca signature (e.g. Figure 3f) through ontogeny (N=3
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57 211 from Brazil, N=1 from Margarita Island, and N=2 from Tamiahua Lagoon). The rest of the
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59 212 individuals showed between 2 and 10 Sr/Ca stable signatures (Figure 3 and 4), with a global mean of
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3 213 4.07±1.85. According to the Kruskal Wallis test ($H=19.8$, $p=0.002$), the highest number of Sr/Ca
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6 214 stable signatures was found in Laguna Madre fish ($4.87±1.1$) and the lowest in Brazil ($3.27±1.70$)
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8 215 (Figure 5). Intermediate mean values were found in the rest of the catch stations (Figure 5).
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10 216 11 12 217 **4. Discussion**

13 14 15 218 16 17 219 **4.1. Otolith chemistry as a salinity proxy**

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19 220 Otolith edge Sr/Ca ratio showed an increase in relation to salinity, as reported by Santana *et al.*
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21 (2018) for *M. curema* from northeast Brazil, and by several studies based on other mugilids such as
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24 222 *M. cephalus* caught in Taiwan (Chang, Iizuka & Tzeng, 2004; Chang, Lin, et al., 2004; Wang et al.,
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26 223 2010) and Australia (Fowler et al., 2016), and *M. liza* from Argentina and Brazil (Callicó Fortunato
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28 et al., 2017).
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32 225 On the other hand, Ba/Ca turned out to be a good indicator of salinity for some species (Elsdon &
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34 226 Gillanders, 2005), but this does not appear to be the case for mugilids in general, where the
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36 227 relationship between Ba/Ca and environment is not fully understood. Specifically, Wang (2014) and
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38 Wang et al. (2010) have reported high otolith Ba/Ca levels for freshwater *M. cephalus* from Taiwan
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41 229 (e.g. Tanshui River), suggesting that this ratio may be useful as a freshwater marker. In addition,
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43 230 Wang *et al.* (2011) and Fowler *et al.* (2016) have reported a weak relationship between otolith Ba/Ca
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45 231 and Sr/Ca in *M. cephalus*. Wang *et al.* (2011) have reported a poor linear relationship (regression
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47 analysis, $r^2 = 0.058$), while Fowler *et al.* (2016) have informed a better fit (exponential fit, $r < 0.6$), but
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49
50 233 have used a sampling size of 8,662 points. These analyses were based on all core-to-edge transect
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52 234 values, then, the significant results probably can be explained by the high number of points included
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54 in the analysis, rather than by a real relationship between both ratios. For *M. curema*, the use of
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57 236 otolith Ba/Ca ratio as a salinity proxy has not been directly validated. Mai et al. (2018) have
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59 237 classified *M. curema* from a relatively small estuary of southeastern Brazil as a marine migrant
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3 238 species by using otolith Ba/Ca. These authors stated that they used Ba/Ca because the Sr/Ca ratio
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6 239 fluctuated among individuals and a directional trend throughout the otolith transect was not clear
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8 240 (Ba/Ca patterns also looked abnormally variable). Moreover, they suggested that, in species that
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10 241 experience salinities from 20 to 35, the Sr/Ca ratio has limited ability to distinguish movements
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12 242 among habitats. For *M. curema* (this study, Ibáñez *et al.*, 2012, and Santana *et al.*, 2018), *M.*
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14 243 *cephalus* (e.g. Wang *et al.*, 2010; Wang, 2014; Fowler *et al.*, 2016) and *M. liza* (e.g. Callicó
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17 244 Fortunato *et al.*, 2017), Sr/Ca did not behave as described by Mai *et al.* (2018), suggesting that these
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19 245 observations could be due to analytical issues rather than intrinsic patterns of the species.
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22 246 Chang *et al.* (2004b) have suggested reference values for marine, freshwater and environmental use
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24 247 of *M. cephalus*, based on otolith Sr/Ca ranges (3×10^{-3} to 7×10^{-3} ppm/ppm for estuarine water use,
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26
27 248 and those above or below corresponded to seawater and freshwater use, respectively). Some authors
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29 249 (Ibáñez *et al.*, 2012; Mai *et al.*, 2018) have assumed that these thresholds are the same for other
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31 250 mugilids species like *M. curema*, which could lead to an error in the classifications of habitat use
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34 251 because the incorporation rate of trace elements into the otolith is species-dependent. Even the
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36 252 incorporation rate can vary within the same species, responding to genetic factors (Clarke, Conover
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38 253 & Thorrold, 2011). To give some examples on other euryhaline fishes, Sr/Ca thresholds (estuarine
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40 254 use) between 3.3–6.4 mmol/mol have been estimated for *M. cephalus* (Fowler *et al.*, 2016), 4.2–6.4
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43 255 mmol/mol for *Zenarchopterus dunckeri* (Kanai *et al.*, 2014), ~2.3–4.2 mmol/mol for *Osmerus*
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45 256 *mordax* (Bradbury, Campana & Bentzen, 2008), and 3.75–5.98 mmol/mol for *G. barbuis* (Avigliano
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48 257 *et al.*, 2017). To avoid adding confusion to the interpretations, a transition threshold was not used for
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50 258 *M. curema* because of the lack of validation in relation to the use of known salinities. Instead, a
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52 259 Sr/Ca guide value (3.46–4.75 mmol/mol) for high salinity environments (salinity=33.5) was used as
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55 260 an aid for the interpretations. The minimum Sr/Ca value found in the present study for use of the
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57 261 high salinity water (3.46 mmol/mol) was remarkably comparable to the maximum ratio (3.38
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59 262 mmol/mol, relative difference of 2.3%) for estuarine use estimated by Santana *et al.* (2018),
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3 263 suggesting that both guide values are reliable. Finally, as it happened in previous studies (Ibáñez et
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6 264 al., 2012; Santana et al., 2018), here it was not possible to distinguish the use of environments with
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8 265 salinities greater than the marine ones, because the Sr/Ca signatures compatible with salinities higher
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10 266 than 33.5 were classified within the same category. This could confuse ocean incursions with the use
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12
13 267 of other environments with marine-like salinity (mangroves, lagoons, among others) or even higher
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15 268 like hypersaline lagoons.

21 270 **4.2. Core Sr/Ca as a potential spawning area indicator**

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23 271 A typical concern in the interpretation of diadromous fish core-to-edge chemical time-series is to
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26 272 assume that the incorporation of elements into otolith only varies with salinity. However, this may
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28 273 not be the case for all species, especially in otolith core (Kalish, 1990; Liberoff et al., 2014; Volk et
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30 274 al., 2000), which would be reflected in a misinterpretation regarding the classification of spawning
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33 275 environments. In some species, the yolk-feeding could influence the Sr incorporation into the core,
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35 276 phenomenon known as “maternal effect” (Kalish, 1990). The maternal effect was observed mainly in
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37 277 salmonids (e.g. *Oncorhynchus mykiss* and *Salmo trutta*) (Kalish, 1990; Liberoff et al., 2014; Volk et
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40 278 al., 2000), which exhibit high egg diameter (>3 mm), long egg duration (30–170 days), and long
41
42 279 yolk-feeding period (> 2 weeks) (Lowe et al., 2012). In those cases, the chemistry of the larval
43
44 280 otolith core is influenced by the environment where the mother hydrated the eggs, and not by
45
46 281 spawning site. Nevertheless, unlike salmonids, *M. curema* has smaller egg diameter (0.46-0.9 mm,
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49 282 Marin et al., 2000), short incubation period prior to hatching (40 hs, Willam, 1957), and a short yolk-
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51 283 feeding periods of 3.5 days (Houde et al., 1976), which represent an otolith diameter of ~60 µm
52
53 284 (Radtke, 1984). In this regard, it was assumed that in this species there is no significant maternal
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56 285 effect at the binning of the Sr/Ca time-series, and if it exists could be imperceptible due to the size of
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58 286 the laser ablation spot used (40 µm).

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3 287 Historically, it was assumed that *M. curema* had a catadromous reproductive behavior, where
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6 288 generally spawns at sea, and the eggs and larvae are carried by the currents to coastal environments
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8 289 (Moore, 1974). In the last two decades, it was reported that the white mullet spawns in mangles and
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10 290 coastal lagoons, and could use brackish environments most of its life (Ibáñez et al., 2012; Marin et
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12 291 al., 2000, 2003; Santana et al., 2018). Here, when the beginning of the transects was analyzed, the
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14
15 292 10.6% of the fish showed a Sr/Ca signatures that overlapped with the GHS range, while the
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17 293 remaining specimens showed values were below GHS (89.4%; total sampling size, Nt=161),
18
19 294 suggesting that the spawning occurs mainly in relatively low or moderate salinity waters. These
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22 295 results suggest that the expected catadromous behavior is less frequent. This observation agreed with
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24 296 those reported by Ibáñez *et al.* (2012) and Santana *et al.* (2018), who found high proportions of core
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26 297 chemical signatures compatible with estuarine hatching for Tamiahua Lagoon (México) (80%,
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28
29 298 Nt=20) and Northeast Brazil (70%, Nt=23), respectively. On the contrary, Ibáñez *et al.* (2012) have
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31 299 found a high proportion (60%, Nt=10) of individuals from the Cazonos Estuary (México) with
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33 300 relatively high chemical signatures in the primordium, suggesting hatching in seawater. However,
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35 301 these estimates were based on a low number of samples, which added to the use of Sr/Ca thresholds
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38 302 of another species could have produced an overestimation of the proportions. In summary, the
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40 303 findings of this work suggest that low or intermediate salinity environments are essential for *M.*
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42 304 *curema* spawning and growth; therefore, they need special attention when generating conservation
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45 305 and management policies.

49 307 **4.3. Core-to-edge Sr/Ca time-series**

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52 308 Three other studies have assessed the ontogenetic migration of *M. curema* using chemical time series
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54 309 in otoliths (Ibáñez et al., 2012; Mai et al., 2018; Santana et al., 2018); nevertheless, the present work
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56 310 has been the most exhaustive in relation to the samples size and study areas so far.
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3 311 The results presented in this study showed a high variability in the Sr/Ca patterns throughout the
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6 312 ontogeny, evidencing a complex habitat use. Interestingly, despite one exception, all fish classified as
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8 313 migratory showed changes between signatures consistent with estuarine and marine (or high salinity)
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10 314 use within the first year of life. This suggests that there is high mobility in the early stages, perhaps
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12 315 associated with the search for nursery areas such those with mangroves or coastal lagoons.
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15 316 Considering all core-to-edge Sr/Ca time-series, the results obtained recorded a wide variation in
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17 317 otolith chemistry in the different locations, being estuarine and marine migrant the most common
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19 318 patterns, as showed by Ibáñez *et al.* (2012). The Sr/Ca patterns suggested that a few fish spent their
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21
22 319 entire life in low or intermediate salinity environments (estuarine resident). Some specimens even
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24 320 recorded Sr/Ca values below 2 mmol/mol (Figure 3a, c and g), which could be associated with the
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26 321 use of freshwater. Santana *et al.* (2018) have suggested that values below 2.29 mmol/mol could be
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29 322 associated with the use of freshwater; however, they have not caught specimens in freshwater to test
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31 323 this value. The freshwater environment use is supported by previous observations, which reported
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33 324 migrations 700 km upstream in rivers from the Gulf of México (Rodiles-Hernández, González-Díaz
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35 325 & Chan-Sala, 2005). In addition, six fish from Brazil, Margarita Island, and Tamiahua Lagoon
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38 326 showed a single stable Sr/Ca signature through ontogeny, which suggests a relatively low mobility
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40 327 between environments with different salinity, highlighting plasticity of the species. Finally, several
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42 328 fish showed relatively high chemical signatures (>7 mmol/mol), suggesting the use of hypersaline
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45 329 environments. These specimens mainly correspond to the hypersaline lagoon Laguna Madre (Figure
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47 330 4i and j), which can reach salinities greater than 60 (Table 1). The use of this environments have also
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49 331 been reported for other sites such the Saloum Delta (Senegal), whose salinities reach 55 (Le Loc'h *et*
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52 332 *al.*, 2015).

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54 333 The number of changes in Sr/Ca stable signatures was highly variable for all sites (Figure 5).
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56 334 Nevertheless, Laguna Madre fish showed the highest values, suggesting that they experienced more
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58 335 salinity changes. Laguna Madre has the widest salinity range of all the sampled sites (Table 1),
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3 336 which allows exposure to extreme salinities. On the contrary, Brazilian fish, which were caught at
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6 337 the sea, seem to have been exposed to fewer changes in salinity, being consistent with a more
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8 338 homogeneous environment in relation to the estuaries or lagoons allocated to the ocean.
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14 340 **5. Final remarks, conservation and management**

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16 341 This work assessed for the first time the habitat use of *M. curema* in a wide range of distribution in
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19 342 the Neotropical Pacific and Atlantic waters. Core-to-edge Sr/Ca time-series suggested a marine and
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21 343 estuarine migratory behavior, rather than catadromous. The apparent number of changes in salinity
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23 344 was highly variable between specimens and studied environments. Finally, high migratory plasticity
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26 345 was observed in most of the studied sites, with the estuary being the most used environment. Due to
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28 346 the complexity in the migration patterns, the conservation and management of this important fishing
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30 347 and cultural resource will also need comprehensive and complex management strategies (Jenkins et
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33 348 al., 2010). This finding suggests that resource management and conservation strategies should focus
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35 349 on various coastal environments, but mainly on those with intermediate and low salinity, such as
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37 350 estuaries and mangroves, which are largely used as spawning, nursery and growth areas. In this
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39 351 sense, the maintenance of corridors between river, estuary, mangrove and sea is needed to preserve
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41
42 352 this migratory fish.

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7. References

- Arai, T. & Chino, N. (2017). Influence of water salinity on the strontium:calcium ratios in otoliths of the giant mottled eel, *Anguilla marmorata*. *Environmental Biology of Fishes*, 100(3), 281–286. <https://doi.org/10.1007/s10641-016-0569-7>
- Astor, Y. & Cárdenas, J. (1997). *Informe de la sexta campaña del proyecto de evaluación pesquera VECEP (Venezuela, Colombia, Ecuador y Perú)*. Caracas: Programa de Cooperación Técnica para la Pesca UE-VECEP ALA 92/43, UE-VECEP.
- Avigliano, E., Maichak de Carvalho, B., Miller, N., Córdoba Gironde, S., Tombari, A., Limburg, K. et al. (2019). Fin spines chemistry as a non-lethal alternative to otoliths for habitat and stock discrimination: comparison between structures for an endangered catfish species. *Marine Ecology Progress Series*, 614, 147–157. <https://doi.org/10.3354/meps12895>
- Avigliano, E. & Volpedo, A.V. (2013). Use of otolith strontium:calcium ratio as an indicator of seasonal displacements of the silverside (*Odontesthes bonariensis*) in a freshwater-marine environment. *Marine and Freshwater Research*, 64(8), 746–751. <https://doi.org/10.1071/MF12165>
- Avigliano, E., Callicó-Fortunato, R., Buitrago, J. & Volpedo, A.V. (2015). Is otolith microchemistry (Sr:Ca and Ba:Ca ratios) useful to identify *Mugil curema* populations in the Southeastern Caribbean Sea? *Brazilian Journal of Biology*, 75(4), 45–51.
- Avigliano, E., Leisen, M., Romero, R., Carvalho, B., Velasco, G., Vianna, M. et al. (2017). Fluvio-marine travelers from South America: Cyclic amphidromy and freshwater residency, typical behaviors in *Genidens barbatus* inferred by otolith chemistry. *Fisheries Research*, 193(April), 184–194. Elsevier. <https://doi.org/10.1016/j.fishres.2017.04.011>
- Avigliano, E., Miller, N. & Volpedo, A.V. (2018). Silversides (*Odontesthes bonariensis*) reside within freshwater and estuarine habitats, not marine environments. *Estuarine, Coastal and Shelf Science*, 205, 123–130. Elsevier Ltd. <https://doi.org/10.1016/j.ecss.2018.03.014>
- Barletta, M. & Dantas, D.V. (2016). Biogeography and distribution of Mugilidae in the Americas. *Biology, Ecology and Culture of Grey Mulletts (Mugilidae)*, 42.
- Beck, M.W., Heck, K.L., Able, K.W., Childers, D.L., Eggleston, D.B., Gillanders, B.M. et al. (2001). The identification, conservation, and management of estuarine and marine nurseries for fish and invertebrates. *BioScience*, 51(8), 633. [https://doi.org/10.1641/0006-3568\(2001\)051\[0633:TICAMO\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2001)051[0633:TICAMO]2.0.CO;2)
- Beck, M.W. & Odaya, M. (2001). Ecoregional planning in marine environments: Identifying priority sites for conservation in the northern Gulf of Mexico. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 11, 235–242. <https://doi.org/10.1002/aqc.449>
- Biolé, F.G., Thompson, G.A., Vargas, C. V., Leisen, M., Barra, F., Volpedo, A.V. et al. (2019). Fish stocks of *Urophycis brasiliensis* revealed by otolith fingerprint and shape in the Southwestern Atlantic Ocean. *Estuarine, Coastal and Shelf Science*, 229, 106406. <https://doi.org/10.1016/j.ecss.2019.106406>
- Bradbury, I.R., Campana, S.E. & Bentzen, P. (2008). Otolith elemental composition and adult tagging reveal spawning site fidelity and estuarine dependency in rainbow smelt. *Marine Ecology Progress Series*, 368, 255–268. <https://doi.org/10.3354/meps07583>
- Brown, R.J. & Severin, K.P. (2009). Otolith chemistry analyses indicate that water Sr:Ca is the primary factor influencing otolith Sr:Ca for freshwater and diadromous fish but not for marine fish. *Canadian Journal of Fisheries and Aquatic Sciences*, 66(10), 1790–1808. <https://doi.org/10.1139/F09-112>
- Callicó Fortunato, R., González-Castro, M., Reguera Galán, A., García Alonso, I., Kunert, C., Benedito Durà, V. et al. (2017). Identification of potential fish stocks and lifetime movement patterns of *Mugil liza* Valenciennes 1836 in the Southwestern Atlantic Ocean. *Fisheries Research*, 193(April), 164–172. Elsevier. <https://doi.org/10.1016/j.fishres.2017.04.005>

- 1
2
3 420 Campana, S.E., Chouinard, G.A., Hanson, J.M., Fréchet, A. & Bratney, J. (2000). Otolith elemental
4 421 fingerprints as biological tracers of fish stocks. *Fisheries Research*, 46(1–3), 343–357.
5 422 [https://doi.org/10.1016/S0165-7836\(00\)00158-2](https://doi.org/10.1016/S0165-7836(00)00158-2)
6 423
7 423 Chang, C.W., Iizuka, Y. & Tzeng, W.N. (2004). Migratory environmental history of the grey mullet
8 424 *Mugil cephalus* as revealed by otolith Sr:Ca ratios. *Marine Ecology Progress Series*, 269, 277–
9 425 288. <https://doi.org/10.3354/meps269277>
10 426
11 427 Chang, C.W., Lin, S.H., Iizuka, Y. & Tzeng, W.N. (2004). Relationship between Sr:Ca ratios in
12 428 Otoliths of grey mullet *Mugil cephalus* and ambient salinity: Validation, mechanisms, and
13 429 applications. *Zoological Studies*, 62, 1809–1813.
14 429
15 430 Clarke, A.D., Telmer, K.H. & Mark Shrimpton, J. (2007). Elemental analysis of otoliths, fin rays and
16 431 scales: A comparison of bony structures to provide population and life-history information for
17 432 the Arctic grayling (*Thymallus arcticus*). *Ecology of Freshwater Fish*, 16(3), 354–361.
18 433 <https://doi.org/10.1111/j.1600-0633.2007.00232.x>
19 434
20 434 Clarke, L.M., Conover, D.O. & Thorrold, S.R. (2011). Population differences in otolith chemistry
21 435 have a genetic basis in menidia menidia. *Canadian Journal of Fisheries and Aquatic Sciences*,
22 436 68(1), 105–114. <https://doi.org/10.1139/F10-147>
23 437
24 438 Clément, M., Chiasson, A.G., Veinott, G. & Cairns, D.K. (2014). What otolith microchemistry and
25 439 stable isotope analysis reveal and conceal about anguillid eel movements across salinity
26 440 boundaries. *Oecologia*, 175, 1143–1153. <https://doi.org/10.1007/s00442-014-2969-8>
27 441
28 441 CNA. (2000). *Lineamientos estratégicos para el desarrollo hidráulico de las regiones hidrológicas*
29 442 *pertenecientes a la región administrativa IV Balsas. Comisión Nacional de Agua/Water*
30 443 *National Commission. México.*
31 444
32 445 Contreras, F. & Castañeda, O. (2004). Coastal lagoons and estuaries of the Gulf of Mexico: toward
33 446 the establishment of ecological indicators. In M. Caso, I. Pisanty & E. Ezcurra (Hrsg.),
34 447 *Diagnóstico Ambiental del Golfo de México* (1. Auflage, S. 628). ,
35 448 SEMARNAT/INE/INECOL/HARP Institute.
36 449
37 449 Crosetti, D., Blaber, S., González-Castro, M. & Ghasemzadeh, J. (2016). Morphology and
38 450 morphometry based taxonomy of Mugilidae. *Biology, Ecology and Culture of Grey Mulletts*
39 451 (*Mugilidae*). <https://doi.org/10.1201/b19927-2>
40 452
41 452 Elliott, M., Whitfield, A.K., Potter, I.C., Blaber, S.J.M., Cyrus, D.P., Nordlie, F.G. et al. (2007). The
42 453 guild approach to categorizing estuarine fish assemblages: a global review. *Fish and Fisheries*,
43 454 8(3), 241–268.
44 455
45 455 Elsdon, T.S. & Gillanders, B.M. (2005). Alternative life-history patterns of estuarine fish: barium in
46 456 otoliths elucidates freshwater residency. *Canadian Journal of Fisheries and Aquatic Sciences*.
47 457 <https://doi.org/10.1139/f05-029>
48 458
49 459 Fowler, A.M., Smith, S., Booth, D.J. & Stewart, J. (2016). Partial migration of grey mullet (*Mugil*
50 460 *cephalus*) on Australia's east coast revealed by otolith chemistry. *Marine Environmental*
51 461 *Research*, 119, 238–244. Elsevier Ltd. <https://doi.org/10.1016/j.marenvres.2016.06.010>
52 462
53 462 Froese, R. & Pauly, D. (2019). Fish Base. World Wide Web electronic publication.
54 463 www.fishbase.org.
55 464
56 464 Harrison, I. (2002). Mugilidae. *FAO Species identification guide for fisheries purposes: the living*
57 465 *marine resources of the western Central Atlantic* (S. 1071–1085). Rome.
58 466
59 466 Harrison, I.J., Nirchio, M., Oliveira, C., Ron, E. & Gaviria, J. (2007). A new species of mullet
60 467 (Teleostei: Mugilidae) from Venezuela, with a discussion on the taxonomy of *Mugil*
61 468 *gaimardianus*. *Journal of Fish Biology*, 71, 76–97. <https://doi.org/10.1111/j.1095-8649.2007.01520.x>
62 469
63 469 Houde, E.D., Berkeley, S.A., Klinovsky, J.J. & Schekter, R.C. (1976). Culture of larvae of the white
64 470 mullet, *Mugil curema* Valenciennes. *Aquaculture*, 8, 365–370. [https://doi.org/10.1016/0044-8486\(76\)90118-6](https://doi.org/10.1016/0044-8486(76)90118-6)
65 471
66 471 Ibañez-Aguirre, A.L. & Gallardo-Cabello, M. (1996). Age determination of the grey Mullet *Mugil*

- 1
2
3 470 *cephafus* L. and the white Mullet *Mugil curema* V. (Pisces: Mugilidae) in Tamiahua Lagoon,
4 471 Veracruz. *Ciencias Marinas*, 22(3), 329–345. <https://doi.org/10.7773/cm.v22i3.861>
5 472 Ibáñez, A.L., Chang, C.W., Hsu, C.C., Wang, C.H., Iizuka, Y. & Tzeng, W.N. (2012). Diversity of
6 473 migratory environmental history of the mullets *Mugil cephalus* and *M. curema* in Mexican
7 474 coastal waters as indicated by otolith Sr: Ca ratios. *Ciencias Marinas*, 38(1A), 73–87.
8 475 Ibáñez, A.L., Hernández-Fraga, K. & Alvarez-Hernández, S. (2017). Discrimination analysis of
9 476 phenotypic stocks comparing fish otolith and scale shapes. *Fisheries Research*, 185, 6–13.
10 477 <https://doi.org/10.1016/j.fishres.2016.09.025>
11 478 Jenkins, A.P., Jupiter, S.D., Qauqau, I. & Atherton, J. (2010). The importance of ecosystem-based
12 479 management for conserving aquatic migratory pathways on tropical high islands: A case study
13 480 from Fiji. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 20, 224–238.
14 481 <https://doi.org/10.1002/aqc.1086>
15 482 Jochum, K.P., Weis, U., Stoll, B., Kuzmin, D., Yang, Q., Raczek, I. et al. (2011). Determination of
16 483 reference values for NIST SRM 610-617 glasses following ISO guidelines. *Geostandards and*
17 484 *Geoanalytical Research*, 35(4), 397–429. <https://doi.org/10.1111/j.1751-908X.2011.00120.x>
18 485 Kalish, J. (1990). Use of otolith microchemistry to distinguish the progeny of sympatric anadromous
19 486 and non-anadromous salmonids. *Fishery Bulletin*, 88(657– 666).
20 487 Kanai, T., Nanjo, K., Yamane, K., Amano, Y., Kohno, H., Watanabe, Y. et al. (2014). Utilization
21 488 patterns of estuarine and marine habitats by the halfbeak *Zenarchopterus dunckeri* at Iriomote
22 489 Island, southern Japan, evaluated from otolith microchemistry. *Fisheries Science*, 80(6), 1231–
23 490 1239. <https://doi.org/10.1007/s12562-014-0797-8>
24 491 Kraus, R.T. & Secor, D.H. (2004). Incorporation of strontium into otoliths of an estuarine fish.
25 492 *Journal of Experimental Marine Biology and Ecology*, 302(1), 85–106.
26 493 <https://doi.org/10.1016/j.jembe.2003.10.004>
27 494 Lemos, V.M., Monteiro-Neto, C., Cabral, H. & Vieira, J.P. (2017). Stock identification of tainha
28 495 (*Mugil liza*) by analyzing stable carbon and oxygen isotopes in otoliths. *Fishery Bulletin*,
29 496 115(2), 201–205. <https://doi.org/10.7755/FB.115.2.7>
30 497 Liberoff, A.L., Miller, J.A., Riva-Rossi, C.M., Hidalgo, F.J., Fogel, M.L., Pascual, M.A. et al.
31 498 (2014). Transgenerational effects of anadromy on juvenile growth traits in an introduced
32 499 population of rainbow trout (*Oncorhynchus mykiss*). *Canadian Journal of Fisheries and Aquatic*
33 500 *Sciences*, 71(November 2013), 398–407. <https://doi.org/10.1139/cjfas-2013-0466>
34 501 Lin, Y.J., Jessop, B.M., Weyl, O.L.F., Iizuka, Y., Lin, S.H. & Tzeng, W.N. (2014). Migratory history
35 502 of African longfinned eel *Anguilla mossambica* from Maningory River, Madagascar: discovery
36 503 of a unique pattern in otolith Sr:Ca ratios. *Environmental Biology of Fishes*, 98(1), 457–468.
37 504 <https://doi.org/10.1007/s10641-014-0275-2>
38 505 Le Loc'h, F., Durand, J.D., Diop, K. & Panfili, J. (2015). Spatio-temporal isotopic signatures ($\delta^{13}\text{C}$
39 506 and $\delta^{15}\text{N}$) reveal that two sympatric West African mullet species do not feed on the same basal
40 507 production sources. *Journal of Fish Biology*, 83, 1444–1453. <https://doi.org/10.1111/jfb.12650>
41 508 Lowe, M.R., Ludsins, S.A., Fryer, B.J., Wright, R.A., DeVries, D.R. & Farmer, T.M. (2012).
42 509 Response to 'Comment on' Otolith Microchemistry Reveals Substantial Use of Freshwater by
43 510 Southern Flounder in the Northern Gulf of Mexico by Pedro Morais. *Estuaries and Coasts*, 35,
44 511 907–910. <https://doi.org/10.1007/s12237-012-9493-z>
45 512 Mai, A.C.G., dos Santos, M.L., Lemos, V.M. & Vieira, J.P. (2018). Discrimination of habitat use
46 513 between two sympatric species of mullets, *Mugil curema* and *Mugil liza* (Mugiliformes:
47 514 Mugilidae) in the rio Tramandaí Estuary, determined by otolith chemistry. *Neotropical*
48 515 *Ichthyology*, 16(2), e170045. <https://doi.org/10.1590/1982-0224-20170045>
49 516 Marin, E., J., B. & Dodson, J.J. (2000). Age, growth and fecundity of the silver mullet, *Mugil curema*
50 517 (Pisces: Mugilidae), in coastal areas of Northeastern Venezuela. *Revista de Biología Tropical*,
51 518 48, 389–398.
52 519 Marin, E., J., B., Quintero, A., Bussière, D. & Dodson, J.J. (2003). Reproduction and recruitment of

- 1
2
3 520 white mullet (*Mugil curema*) to a tropical lagoon (Margarita Island, Venezuela) as revealed by
4 521 otolith microstructure. *Fishery Bulletin*, 101, 809–821.
- 5 522 Moore, R.H. (1974). General ecology, distribution and relative abundance of *Mugil cephalus* and
6 523 *Mugil curema* on the south Texas coast. *Contrib. Mar. Sci.*, 18, 241–245.
- 7 524 Pacheco-Almanzar, E., Loza-Estrada, N. & Ibáñez, A.L. (2020). Do the fish scales shape of *Mugil*
8 525 *curema* reflect the genetic structure using microsatellites markers and the Mexican Marine
9 526 Ecoregions Classification? *Frontiers in Marine Science*.
10 527 <https://doi.org/10.3389/fmars.2020.00166>
- 11 528 Passos, C.V.B., Fabr e, N.N., Malhado, A.C.M., Batista, V.S. & Ladle, R.J. (2016). Estuarization
12 529 increases functional diversity of demersal fish assemblages in tropical coastal ecosystems.
13 530 *Journal of fish biology*, 89(1), 847–862. <https://doi.org/10.1111/jfb.13029>
- 14 531 Pearce, N.J.G., Perkins, W.T., Westgate, J.A., Gorton, M.P., Jackson, S.E., Neal, C.R. et al. (1997).
15 532 A compilation of new and published major and trace element data for NIST SRM 610 and NIST
16 533 SRM 612 glass reference materials. *Geostandards Newsletter*, 21(1), 115–144.
17 534 <https://doi.org/10.1111/j.1751-908X.1997.tb00538.x>
- 18 535 Raabe, J. & Gardner, B. (2013). A spatial capture–recapture model to estimate fish survival and
19 536 location from linear continuous monitoring arrays. *Canadian Journal of Fisheries and Aquatic*
20 537 *Sciences*, 130(September 2013), 120–130. <https://doi.org/10.1139/cjfas-2013-0198>
- 21 538 Radtke, R.L. (1984). Formation and structural composition of larval striped mullet otoliths.
22 539 *Transactions of the American Fisheries Society*, 113(2), 186–191. [https://doi.org/10.1577/1548-8659\(1984\)113<186:fascol>2.0.co;2](https://doi.org/10.1577/1548-8659(1984)113<186:fascol>2.0.co;2)
- 23 540 Rodiles-Hernández, R., González-Díaz, A.A. & Chan-Sala, C. (2005). Lista de Peces Continentales
24 541 de Chiapas, México. *Hidrobiológica*, 15, 245–253.
- 25 542 Santana, F.M., Morize, E., Labonne, M., Lessa, R. & Clavier, J. (2018). Connectivity between the
26 543 marine coast and estuary for white mullet (*Mugil curema*) in northeastern Brazil revealed by
27 544 otolith Sr:Ca ratio. *Estuarine, Coastal and Shelf Science*, 215, 124–131.
28 545 <https://doi.org/10.1016/j.ecss.2018.09.032>
- 29 546 da Silva, V.E.L., Teixeira, E.C., Batista, V.S. & Fabr e, N.N. (2018). Spatial distribution of juvenile
30 547 fish species in nursery grounds of a tropical coastal area of the south-western Atlantic. *Acta*
31 548 *Ichthyologica et Piscatoria*, 48(1), 245–252. <https://doi.org/10.3750/AIEP/02299>
- 32 549 Soeth, M., Spach, H.L., Daros, F.A., Adelir-Alves, J., de Almeida, A.C.O. & Correia, A.T. (2019).
33 550 Stock structure of Atlantic spadefish *Chaetodipterus faber* from Southwest Atlantic Ocean
34 551 inferred from otolith elemental and shape signatures. *Fisheries Research*, 211, 81–90.
35 552 <https://doi.org/10.1016/j.fishres.2018.11.003>
- 36 553 Tabouret, H., Bareille, G., Claverie, F., P echeyran, C., Prouzet, P. & Donard, O.F.X. (2010).
37 554 Simultaneous use of strontium:calcium and barium:calcium ratios in otoliths as markers of
38 555 habitat: Application to the European eel (*Anguilla anguilla*) in the Adour basin, South West
39 556 France. *Marine Environmental Research*, 70(1), 35–45.
40 557 <https://doi.org/10.1016/j.marenvres.2010.02.006>
- 41 558 Thomas, O.R.B. & Swearer, S.E. (2019). Otolith Biochemistry—A Review. *Reviews in Fisheries*
42 559 *Science and Aquaculture*, 27(4), 458–489. <https://doi.org/10.1080/23308249.2019.1627285>
- 43 560 Volk, E.C., Blakley, A., Schroder, S.L. & Kuehner, S.M. (2000). Otolith chemistry reflects migratory
44 561 characteristics of Pacific salmonids: Using otolith core chemistry to distinguish maternal
45 562 associations with sea and freshwaters. *Fisheries Research*, 46, 251–266.
46 563 [https://doi.org/10.1016/S0165-7836\(00\)00150-8](https://doi.org/10.1016/S0165-7836(00)00150-8)
- 47 564 Wang, C.H. (2014). Otolith elemental ratios of flathead mullet *Mugil cephalus* in Taiwanese waters
48 565 reveal variable patterns of habitat use. *Estuarine, Coastal and Shelf Science*, 151(5), 124–130.
49 566 <https://doi.org/10.1016/j.ecss.2014.08.024>
- 50 567 Wang, C.H., Hsu, C.C., Chang, C.W., You, C.F. & Tzeng, W.N. (2010). The migratory
51 568 environmental history of freshwater resident flathead mullet *Mugil cephalus* L. in the Tanshui
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River, northern Taiwan. *Zoological Studies*, 49, 504–514.

Wang, C.H., Hsu, C.C., Tzeng, W.N., You, C.F. & Chang, C.W. (2011). Origin of the mass mortality of the flathead grey mullet (*Mugil cephalus*) in the Tanshui River, northern Taiwan, as indicated by otolith elemental signatures. *Marine Pollution Bulletin*, 62,) 1809–1813.

<https://doi.org/10.1016/j.marpolbul.2011.05.011>

Willam, A. (1957). Early development, spawning, growth, and occurrence of the silver mullet (*Mugil curema*) along the south atlantic coast of the united states. *Fishery Bulletin*, 57, 397–414.

Wynne, L.M.P., Wilson, K.A. & Limburg, K.E. (2015). Retrospective examination of habitat use by blueback herring (*Alosa aestivalis*) using otolith microchemical methods. *Canadian Journal of Fisheries and Aquatic Science*, 72, 1073–1086. <https://doi.org/10.1139/cjfas-2014-0206>

Yoshinaga, J., Nakama, A., Morita, M. & Edmonds, J.S. (2000). Fish otolith reference material for quality assurance of chemical analyses. *Marine Chemistry*, 69(1–2), 91–97.

[https://doi.org/10.1016/S0304-4203\(99\)00098-5](https://doi.org/10.1016/S0304-4203(99)00098-5)

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585 Table 1. Characterization of sampling locations.

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Location	Coast	N	Total length mean±std (cm)	Age (year)	Salinity range (ppt)	Depth mean (m)	Temperature range (°C)	pH range	Habitat type	Reference
Laguna Madre, México	Gulf of México	20	30.2±1.5	2-4	33.5-63.0	1.1	14.6-28.0	7.9-9.4	Hypersaline lagoon	Contreras & Castañeda, 2004
Tamiahua, México	Gulf of México	19	29.9±2.1	2-4	14.4-37.5	2.5	23.0-34.0	7.5-9.4	Polyhaline lagoon surrounded by mangrove	Contreras & Castañeda, 2004
Alvarado, México	Gulf of México	19	29.4±1.6	2-4	0.1-39.0	2.0	22.5-33.0	7.0-9.0	Coastal lagoon system dominated by mangrove	Contreras & Castañeda, 2004
Mecoacán, México	Gulf of México	17	29.7±2.4	2-3	0.5-29.0	1.2	26.2-29.0	-	Coastal lagoon system dominated by mangrove	Contreras & Castañeda, 2004
Balsas, México	Pacific	19	32.7± 2.15	2-4	0.0-34.5	5	24.0-26.0	-	Freshwater river	CNA, 2000
Margarita Island, Venezuela	Caribbean Sea	11	26.7±2.1	2-3	35.8-36.6	<5	23.5-24.5	-	Beach	Astor & Cárdenas, 1997
North Alagoas, Brasil	Atlantic Sea	56	29.1±2.0	2-4	29-35	20	28.03±0.38	-	Costal sea and low estuary	Passos et al., 2016

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589 Figure 1: *Mugil curema* sampling sites. Charts show the classification of the fish based on otolith
Sr/Ca ontogenetic profiles.

591 Figure 2: *Mugil curema* otolith sectioned at the core level showing the core-to-edge-laser ablation
transects. The red line represents the Sr/Ca ratio and the arrows indicate the *annuli* position.

593 Figure 3: Otolith core-to-edge Sr/Ca profiles of *Mugil curema*. Solid horizontal lines illustrate stable
signatures identified using change-point analysis. The light blue horizontal bar suggests high
salinity environment use (~33.5). TL= total length (cm).

596 Figure 4: Otolith core-to-edge Sr/Ca profiles of *Mugil curema*. Solid horizontal lines illustrate stable
signatures identified using change-point analysis. The light blue horizontal bar suggests high
salinity environment use (~33.5). TL= total length (cm).

599 Figure 5: Box plot based on the number of significant changes in the Sr/Ca signatures through
ontogeny. Different letters show significant differences between sampling sites (Kruskal Wallis
test, $p < 0.05$).

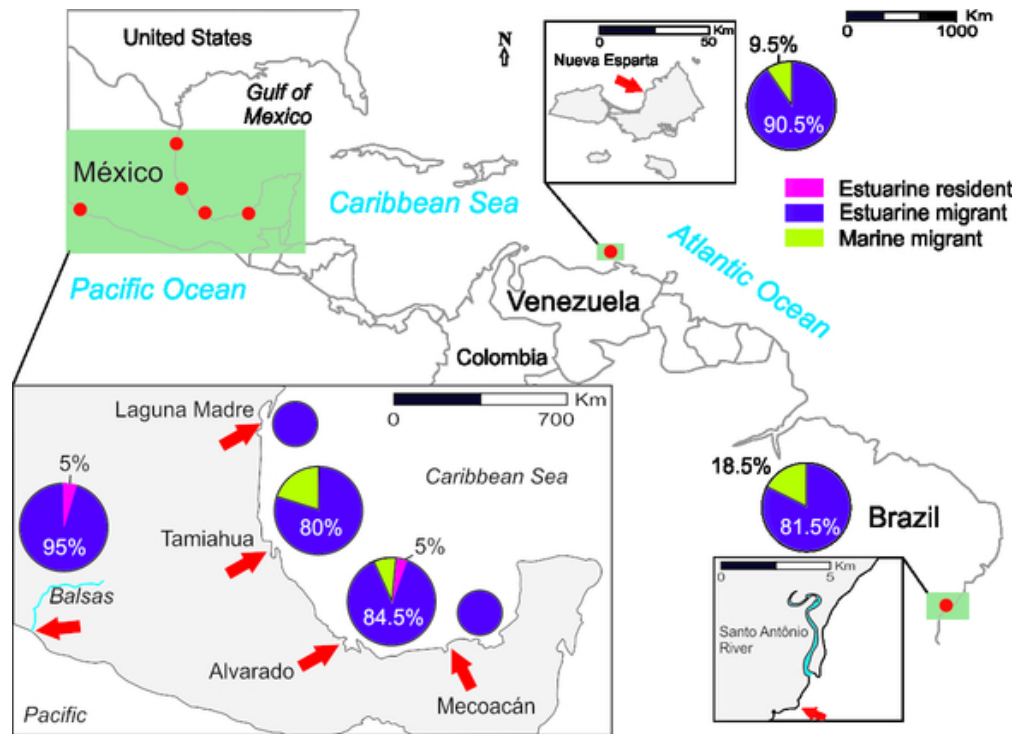


Figure 1: *Mugil curema* sampling sites. Charts show the classification of the fish based on otolith Sr/Ca ontogenetic profiles.

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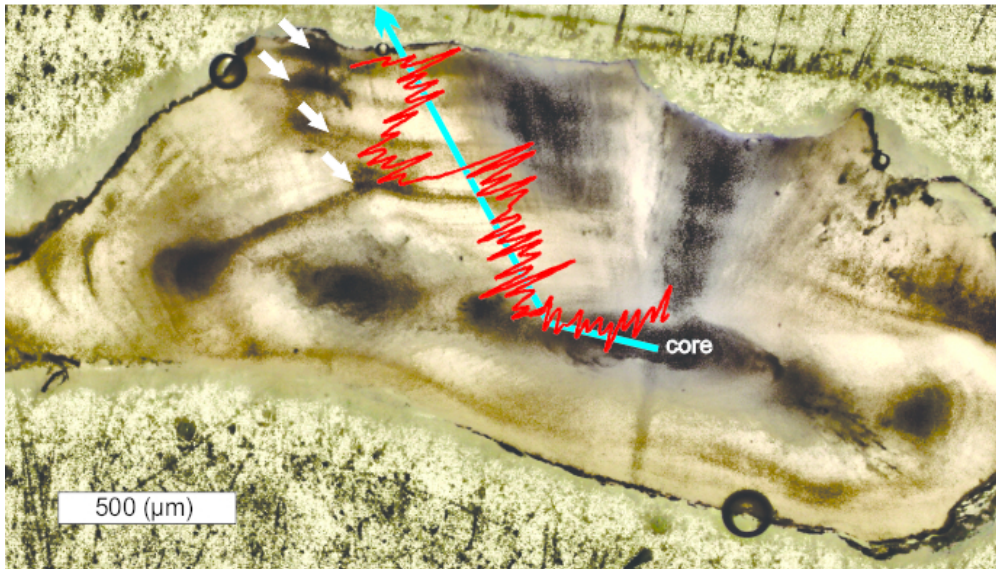


Figure 2: Mugil curema otolith sectioned at the core level showing the core-to-edge-laser ablation transects. The red line represents the Sr/Ca ratio and the arrows indicate the annuli position.

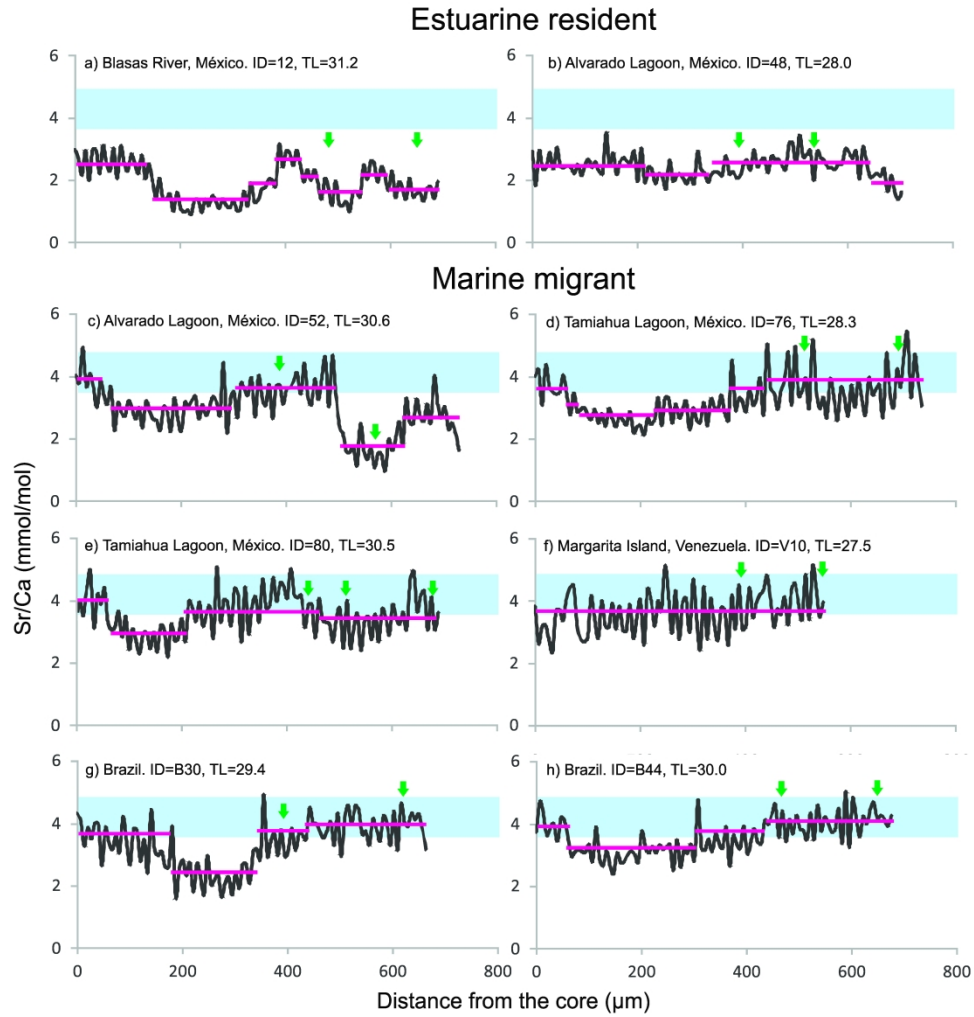


Figure 3: Otolith core-to-edge Sr/Ca profiles of *Mugil curema*. Solid horizontal lines illustrate stable signatures identified using change-point analysis. The light blue horizontal bar suggests high salinity environment use (~ 33.5). TL= total length (cm).

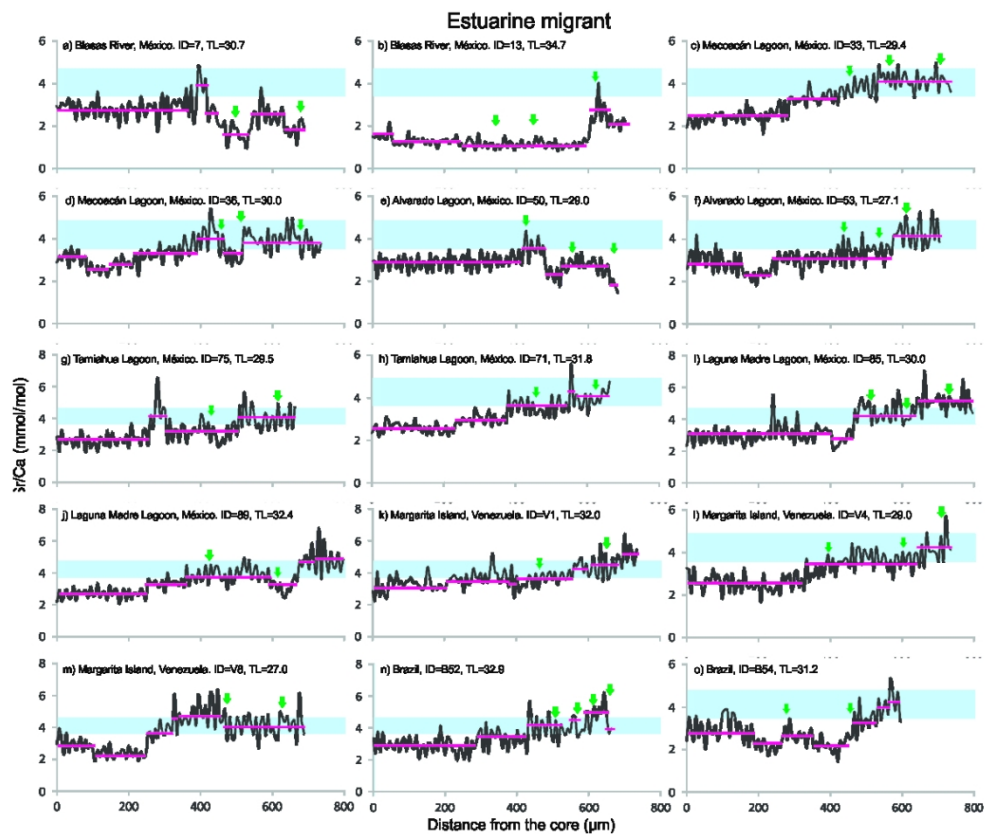


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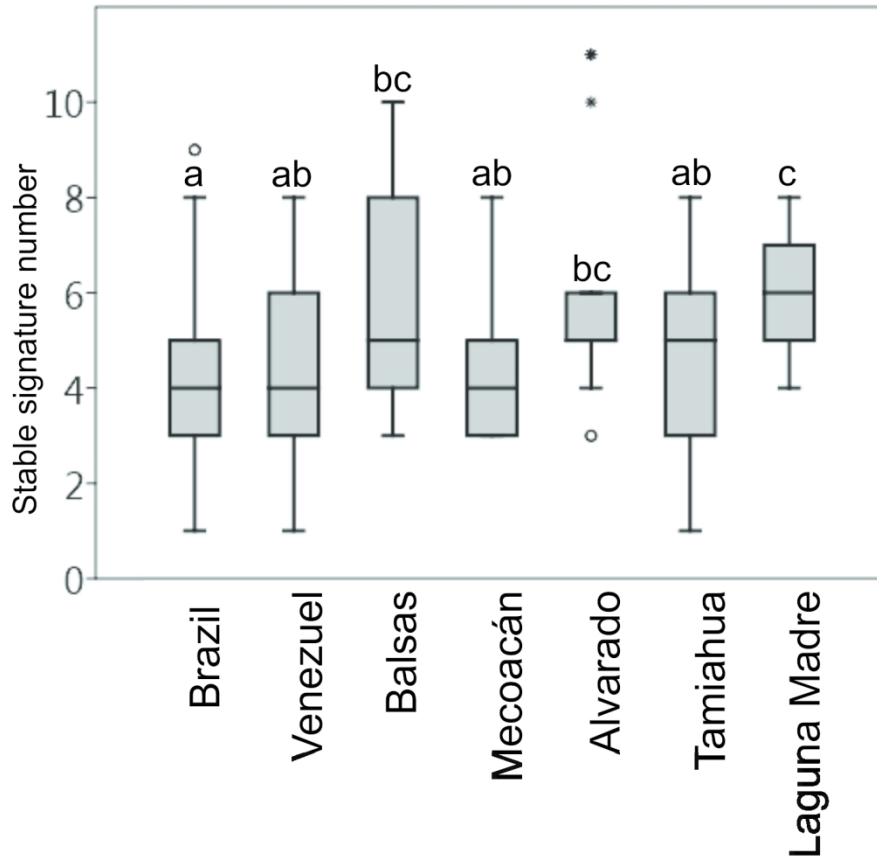


Figure 5: Box plot based on the number of significant changes in the Sr/Ca signatures through ontogeny. Different letters show significant differences between sampling sites (Kruskal Wallis test, p < 0.05).