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Cr(III)-Cr(VI) speciation

- Hexavalent chromium quantification by isotope dilution mass spectrometry in potentially
 contaminated soils from south Italy
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13 Abstract

Due to carcinogenicity of hexavalent chromium [Cr(VI)], its accurate quantification in Cr-14 contaminated soils is of paramount importance. The aim of this work was to quantify Cr(VI) by 15 species-specific IDMS in soil samples from two Italian case studies: A) farmland potentially 16 contaminated by pseudo-total Cr and Zn and heavy hydrocarbons due to past illegal burial of 17 tannery wastes; B) Solofrana valley where volcanic soils are potentially contaminated by pseudo-18 total Cr and Cu due to tannery activities. Hexavalent Cr extraction from soils was performed by 19 focused microwaves (5 min at 80 °C) using 50 mM EDTA, followed by the separation of Cr species 20 by IC and detection by ICP-MS. The Cr(VI) extracted from 20 soil samples of case study A ranged 21 from 0.15 to 11.18 µg g⁻¹, with 70 % of samples exceeding the Cr(VI) screening value set by Italian 22 Parliament for residential/urban soil to assess their potential contamination. Higher levels of Cr(VI) 23 (22.0-107.1 µg g⁻¹) were extracted from other 7 Cr-most-contaminated soil samples, which required 24

a pre-treatment with n-hexane to remove part of organic compounds from each sample, since these reducing agents made the quantification of Cr(VI) by IDMS more challenging because they caused an almost complete reduction of ${}^{50}Cr(VI)$ used for IDMS quantification. Hexavalent Cr extracted from soil samples of case study B ranged from 0.70 to 5.79 µg g⁻¹, with 42 % of samples exceeding the value set by Italian legislation. In both case studies, the Cr(VI) extracted from soil was significantly correlated to the pseudo-total Cr content.

- 31
- 32 Keywords: Cr(VI); IDMS; Cr speciation; soil contamination; tannery wastes; heavy hydrocarbons.
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34 1. INTRODUCTION

Chromium mainly exists in two stable oxidation states, trivalent chromium [Cr(III)] and hexavalent 35 chromium [Cr(VI)] (Jacobs and Testa, 2005; Lilli et al., 2015; Ščančar and Milačič, 2014; Shahid et 36 al., 2017). Trivalent Cr can be considered an essential nutrient for human health, being involved in 37 glucose, insulin and protein metabolism, while Cr(VI) is classified as Group A human carcinogen, 38 also provoking severe health problems on the respiratory tract, liver, kidney, gastrointestinal and 39 immune systems and possibly on the blood (IARC, 2012). Thus, several directives have been 40 adopted at the European and international levels to perform Cr speciation in the environmental 41 studies (Séby and Vacchina, 2018). 42

Chromium mobility and bioavailability in soil environments strongly depend on its chemical form 43 (Avudainayagam et al., 2003). Trivalent Cr is undoubtedly the most thermodynamically stable form 44 of Cr in soil, as it readily precipitates as chromium hydroxide [Cr(OH)₃] and iron-chromium 45 46 hydroxide [(Fe,Cr)(OH)₃] or becomes immobilised after sorption onto soil colloids (Leita et al., 2009). On the other hand, Cr(VI) is thermodynamically metastable in the pore solution and much 47 more mobile in soil, since Cr(VI) oxyanions (chromate CrO_4^{2-} , hydrogen chromate $HCrO_4^{-}$, and 48 dichromate $Cr_2O_7^{2-}$) are not adsorbed onto soil colloids under sub-neutral to alkaline conditions 49 (Becquer et al., 2003; Mpouras et al., 2017). At these soil pHs, Cr(VI) can be present in the form of 50 solid minerals in association with different cations characterised by a wide range of solubility: 51 barium, calcium, lead and zinc chromates are the most insoluble (Kotas and Stasicka, 2000; Pettine 52 and Capri, 2005), while alkaline and some alkaline-earth chromates are highly soluble. Being a 53 strong oxidant, Cr(VI) can be readily reduced to Cr(III) in the presence of various electron donors 54 such as soil organic matter (Bolan et al., 2003; Saha et al., 2011); however, with favourable pedo-55 climatic conditions, even in organic soils, Cr(VI) may persist for prolonged periods of time (Leita et 56 al., 2009). In most cases, the presence of Cr(VI) in the soil is due to anthropogenic sources, since it 57 is used in a number of industrial applications such as electroplating, tanning, industrial water 58 cooling, paper pulp production and petroleum refining (Choppala et al., 2013; Tchounwou et al., 59

60 2012). The levels and forms of Cr(VI) in soils depend on several factors such as pH (HCrO₄⁻ and 61 $Cr_2O_7^{2-}$ at pH 0.7-6.5; and CrO₄²⁻ at pH 6.5-14.0), the redox potential and the presence of natural 62 oxidants (e.g., manganese oxide) or reducing agents [e.g., Fe(II), sulphide and organic matter] 63 (Banks et al., 2006; Choppala et al., 2018).

The determination of Cr(VI) in soil samples is often complicated, because quantitative extraction of the contaminant must be assured while avoiding Cr(III)-Cr(VI) redox interconversion reactions both during extraction and the subsequent chemical analysis (Pettine and Capri, 2005; Weibel et al., 2016). In addition to total Cr(VI) content, the easily-extractable and highly-mobile fractions of Cr(VI) are also determined, in soils where Cr contamination highly impacts on the surrounding environment (James et al., 1995; Zuliani et al., 2013).

Isotope dilution mass spectrometry (IDMS) is a calibration technique able to accurately quantify 70 Cr(VI) from soil samples. When multiple spiking is applied Cr(III)-Cr(VI) redox interconversions 71 72 during sample preparation can be corrected for (Fabregat-Cabello et al., 2012; Garcia Alonso and Rodriguez-Gonzalez, 2013; Guidotti et al., 2015; Rahman et al., 2005). The analytical procedure, 73 standardised by USEPA as method 6800 (USEPA, 2007) requires the addition of known amounts of 74 isotopically enriched Cr(III) and Cr(VI) to the samples before extraction. The selection of the most 75 proper extracting solution plays an important role on the correct quantification of the levels of 76 Cr(VI) in soils. The use of ethylenediaminetetraacetic acid (EDTA) as extracting solution, buffered 77 at pH 10.0 (Korolczuk and Grabarczyk, 2005), shows two main advantages over the well-78 recognised USEPA 3060A (USEPA, 1995) extraction procedure at pH 12.0: i) it complexes Cr(III) 79 preventing its oxidation to Cr(VI) throughout the analytical procedure; ii) it facilitates the extraction 80 of Cr(VI) from the solid sample by complexation of other metals (e.g., Ba, Pb, Sr, Zn) forming 81 insoluble chromates (Fabregat-Cabello et al., 2012; Guidotti et al., 2015). Therefore, since EDTA 82 was able to prevent Cr(III) oxidation to Cr(VI) during extraction, the use of a single isotopically-83 labelled Cr(VI)-spike provides an accurate quantification of Cr(VI) in soil by IDMS (Fabregat-84 Cabello et al., 2012). 85

In the present study single-spike IDMS in combination with EDTA at basic pH (10.0) was applied to quantify Cr(VI) in soil samples from two rural areas of south Italy: A) 27 soil samples from six hectares of farmland potentially contaminated by pseudo-total Cr (up to 10,046 μ g g⁻¹), Zn and heavy hydrocarbons (C>12), due to past illegal burial of industrial wastes; B) 12 soil samples from the Solofrana valley where volcanic soils are potentially contaminated by pseudo-total Cr (up to 594 μ g g⁻¹) and Cu due to tannery activities.

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93 2. EXPERIMENTAL SECTION

94 2.1. Case studies, soil sampling and processing

95 2.1.1. Case study A - Farmland contaminated by illegally burial of tannery wastes

Case study A is a $60,000 \text{ m}^2$ farmland located in the Campania Plain (Province of Naples, 96 Campania, south Italy). This area is currently confiscated by the Italian Judiciary (Caporale et al., 97 98 2018), due to a past illegal burial of industrial wastes of leather-tanning origin. From the geomorphological and geological points of view, case study A is located in a lowland environment, 99 the Piana Campana graben, on the north of the Phlegrean Fields and NW of the Somma-Vesuvius 100 complex, which are the main active volcanoes of the region. Due to the proximity to those volcanic 101 centres, a preferential accumulation of pyroclastic deposits is found in these areas (Di Vito et al., 102 2013; Orsi et al., 2004) and then the soil parent material is mostly volcanic ash and pumice. 103 According to the soil maps of Di Gennaro et al. (2002) and World Reference Base classification 104 (FAO, 1998), the soils in the surroundings of case study A were classified as Pachi-Vitric Andosols. 105 At the end of 2015, a soil contamination monitoring campaign, aimed to address the level and 106 spatial variability of contamination, was carried out in case study A. Soil samples were collected at 107 3 depths: 0-20 cm (d1), 30-60 cm (d2) or 70-90 cm (d3), from georeferenced points according to a 108 sampling grid of 20x20 m. Also, soil samples were collected from pedological profiles and trenches 109 (pt) opened in the field and well describing the complex distribution of the buried waste materials 110 (Caporale et al., 2018). Prior to analysis, soil samples were firstly air-dried, then 2-mm sieved and 111

lastly pulverised (particles $< 177 \mu m - 80$ Mesh) by PM 200 ball mill (Reutch). On the basis of 112 these analyses, site A was found to be potentially contaminated by pseudo-total Cr (up to 10,046 µg 113 g^{-1}), Zn (up to 1,846 µg g^{-1}) and heavy hydrocarbons (C>12, up to 2,150 µg g^{-1}), mostly 114 concentrated in the first metre of depth. Since the contaminant pseudo-total concentrations exceed 115 the screening values set by Italian Parliament for residential/urban soil (150 μ g g⁻¹ for Cr and Zn; 50 116 $\mu g g^{-1}$ for heavy hydrocarbons), according to Italian legislation (Italian-Parliament, 2006) the site is 117 classified as potentially contaminated by pseudo-total Cr and Zn and heavy hydrocarbons and then 118 in need of investigation to assess the factual risk for surrounding environment and human health. 119 Pseudo-total Cr was quantified by aqua regia digestion (ISO 11466, 1995) and Inductively Coupled 120 Plasma (ICP-MS) analysis at Acme Analytical Laboratories Ltd (Vancouver, Canada), while heavy 121 hydrocarbons were analysed by UNI EN ISO 16703 (2011) at Chelab Srl (Resana, TV, Italy). The 122 contents in soil organic carbon (C) and organic matter ($OM = C \times 1.724$) were determined by wet 123 124 digestion using the Walkley-Black procedure (Walkley and Black, 1937). The quantification of Cr(VI) by IDMS was carried out at the University of Oviedo (Spain) on selected soil samples 125 (n=27), chosen according to the overall level of contamination (pseudo-total Cr and heavy 126 hydrocarbons) and the spatial distribution in the field. 127

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129 2.1.2. Case study B - Farmland contaminated by floods with Cr-enriched river sediments

Case study B is located in the Solofrana river valley (Province of Avellino, Campania, south Italy), 130 where volcanic soils with moderate to high andic properties are potentially contaminated by 131 pseudo-total Cr and Cu due to tannery activities. Chromium contamination results from the long use 132 of contaminated river waters for irrigation (prohibited since 1990) and frequent floods after intense 133 rainfall, spreading Cr-enriched sediments on surrounding surfaces (Adamo et al., 2003; Adamo et 134 al., 2006; D'Ascoli et al., 2006; Zampella et al., 2010). However, there is still no evidence that 135 Cr(VI) occurs in these soils. The quantification of Cr(VI) by IDMS was carried out at the 136 University of Oviedo in 12 air-dried, 2-mm sieved and pulverised (particles $< 177 \mu m - 80$ Mesh) 137

soil samples, collected from topsoil (0-20 cm) of areas affected by flooding (70 to 550 m far from
the Solofrana riverbeds) in a past soil contamination monitoring campaign, object of several
scientific studies (Adamo et al., 2003; Adamo et al., 2006; D'Ascoli et al., 2006; Zampella et al.,
2010) and adequately refrigerated and stored over time.

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143 2.2. Preliminary investigations: standard Cr net oxidation test and Cr(VI) quantification by 144 colorimetric-based techniques in soil samples from case study A

The standard Cr net oxidation test (Bartlett and James, 1996) was performed on selected soil 145 samples (n=10) from case study A, in a pseudo-total Cr concentration range of 34-4,487 μ g g⁻¹. The 146 analytical procedure is based on the contact between the soil and a solution of Cr(III) (CrCl₃ as 147 source) and the subsequent measurement of the possible formed-Cr(VI) by colorimetric reaction of 148 Cr(VI) with diphenylcarbazide and spectrophotometric detection at 540 nm. The number of 149 150 potentially oxidisable moles of Cr(III) to Cr(VI) by soil manganese oxides present in the soil was calculated by dividing the number of moles of Mn from easily reducible manganese oxides by 1.5, 151 being 1.5 the molar ratio between Mn and Cr(III) in the oxidation reaction (Chon et al., 2008). The 152 moles of Mn from easily reducible manganese oxides were determined by analysing the extract of 153 the second step of the three-stage sequential extraction protocol proposed by the Measurements and 154 Testing Programme - MAT (formerly BCR) of the European Commission (Rauret et al., 1999). 155

The extraction of total Cr(VI) was carried out in a few representative samples according to USEPA 156 3060A method (USEPA, 1995), which provides an extraction in a strongly basic environment (pH 157 12.0) at 90 °C for 1 h. The determination of Cr(VI) in the extract was carried out according to the 158 USEPA 7196A method (USEPA, 1992), which provides a specific colorimetric reaction between 159 Cr(VI) and diphenylcarbazide in an acidic environment and a spectrophotometric detection at 540 160 nm of the colored complex Cr(III)-difenylcarbazone. A Lambda 25, Perkin Elmer (Waltham, MA, 161 USA) was employed for this purpose. Some modifications to the official analytical methods were 162 made to minimise Cr(VI) reduction to Cr(III) during extraction: 1) extraction at ambient 163

temperature and 2) purification of the extract from organic matter in basic environment by
aluminum sulphate (Ministerial-Decree, 1999). The determination of Cr(VI) in the extracts from
USEPA 3060A was carried out even by USEPA 7199 method (USEPA, 1996), which provides a
phase of purification of the extract from the organic matter with the aid of an ion chromatograph
Dionex DX-120 (Sunnyvale, CA, USA), equipped with guard column (Dionex IonPac NG1) and an
analytical column (Dionex IonPack AS7).

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171 **2.3. Instrumentation**

The microwave-assisted extraction of Cr(VI) from the soil samples was carried out by an Explorer 172 Hybrid focused microwave system (CEM, Mathews, NC). Focused microwaves were applied to the 173 samples contained in 10-mL disposal glass vials. The system was equipped with temperature and 174 pressure feedback control and magnetic stirring, so that the temperature and pressure of the vial was 175 176 controlled throughout the extraction step. The chromatographic separation of the trivalent and hexavalent Cr species was performed on a Dionex DX-120 Liquid Chromatography (LC) system 177 (Sunnyvale, CA, USA) using a Dionex Ion pack AS9-HC anion-exchange column (PEEK, 178 4×250mm×9µm). The detection of the Cr(III) and Cr(VI) species was carried out by an Agilent 179 7500c quadrupole ICP-MS instrument (Agilent Technologies, Tokyo, Japan) equipped with an 180 octapole collision cell. Helium was used as the collision gas (Fabregat-Cabello et al., 2012; Guidotti 181 et al., 2015) to minimise polyatomic interferences (e.g. ${}^{35}Cl^{16}O^{1}H^{+}$, ${}^{35}Cl^{17}O^{+}$ and ${}^{40}Ar^{12}C^{+}$ for ${}^{52}Cr^{+}$). 182 The Cr(VI) extraction solution and the mobile phase for LC-ICP-MS analyses were prepared by 183 dissolving 50 and 5 mmol, respectively, of ethylenediaminetetraacetic acid disodium salt dihydrate 184 (Na₂-EDTA, Sigma, St. Louis, MO) in 1 L of ultrapure water at pH 10.0, adjusted with NH₄OH (30 185 %) (Merck, Darmstadt, Germany). 186

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188 2.4. Reagents and Materials

The ⁵⁰Cr-enriched Cr(VI) spike solution was provided by ISC-Science (Oviedo, Spain). Table 1 189 shows the isotopic composition (% abundance) and the concentration of ⁵⁰Cr-enriched Cr(VI), 190 quantified by reverse IDMS analysis. For this purpose, a natural abundance Cr(VI) standard was 191 prepared dissolving at pH 10.0 potassium dichromate (K₂Cr₂O₇) certified by the German Federal 192 Institute for Materials Research and Testing (BAM Berlin). The standard reference materials SRM 193 2700 (hexavalent chromium in contaminated soil, low level: $14.9\pm1.2 \text{ ug g}^{-1}$) and SRM 2701 (soil 194 containing chromite ore processing residue, high level: $551.2\pm34.5 \ \mu g \ g^{-1}$), purchased from NIST 195 (National Institute of Standards and Technology, USA), were analysed for quality control purposes. 196 The quantitative extraction of Cr(VI) from both reference materials required a two-cycle of 197 microwave-assisted extraction at 90 °C for 5 min, as reported Fabregat-Cabello et al. (2012). 198 Unfortunately, these latter microwave-assisted extraction conditions were found not applicable to 199 real soil samples from a contaminated area of northern Italy, because they caused the complete 200 201 reduction of Cr(VI) (Guidotti et al., 2015). Thus, as assessed by Guidotti et al. (2015), in real soils the milder microwave-assisted extraction at 80 °C for 5 min represents the best compromise 202 solution to assure both accurate Cr(VI) quantification and minimisation of the reduction of the 203 Cr(VI) during extraction. Accordingly, we applied in this work the extraction conditions proposed 204 by Guidotti et al. (2015) to avoid Cr(VI) reduction in our soil samples, since they were set-up by the 205 same lab equipments and chemicals employed in this work. 206

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208 **2.5** Sample preparation procedures for the analysis of the soil samples

The single-spike IDMS procedure requires the addition of known amount of isotopically-labelled Cr(VI) to the samples to reach isotopic equilibration (Guidotti et al., 2015). A total of 0.10 g of the soil samples was directly weighed into a 10-mL microwave glass vessel. Then, 5 mL of 50 μ mol mL⁻¹ Na₂-EDTA (pH 10.0) was added, and the sample was lately spiked with 0.10 g of a 3.4 or 34 μ g g⁻¹ ⁵⁰Cr-enriched Cr(VI) spike solution [according to the expected natural Cr(VI) concentration in the soil sample]. The amounts of sample and ⁵⁰Cr-enriched Cr(VI) solution were accurately

weighted by an analytical balance with a precision of 0.01 mg. The ratio of concentrations between natural Cr(VI) in soil sample and 50 Cr-enriched Cr(VI) always ranged from 0.1 to 10 to ensure accurate and precise results (Fabregat-Cabello et al., 2012).

The focused microwave assisted extraction was carried out at 80 °C for 5 min. After extraction, the 218 samples were centrifuged at 5000 rpm for 5 min. An aliquot of 1 mL of the extract was diluted 1:10 219 with ultrapure water. Finally 100 µL of the diluted extract were injected into the LC-ICP-MS 220 system. The use of a 1:10 dilution of the Na₂-EDTA extracting solution as chromatographic mobile 221 phase avoided the need for neutralisation or acidification of the sample. Also, the centrifugation 222 step avoided the need of sample filtering allowing a quicker sample preparation and a lower risk of 223 contamination and/or interconversion of the Cr species (Fabregat-Cabello et al., 2012; Guidotti et 224 al., 2015). 225

A pre-treatment with n-hexane was applied in the analysis of 7 Cr-most-contaminated soil samples from case study A. Briefly, 0.1 g of each soil sample were pre-treated with 5 mL of n-hexane and an mechanically shaken for 15 min in an horizontal shaker at 150 rpm. Then the sample was centrifuged at 5000 rpm for 5 min. Finally the hexane was removed from the soil sample before microwave-assisted extraction (at 80 °C for 5 min).

The reduction (%) of the 50 Cr-enriched Cr(VI) spike after microwave-assisted extraction was even calculated by IDMS, analysing Cr(III)-EDTA⁻ and Cr(VI) peak areas by linear multiple regression, similarly to the approach followed by Pinel-Raffaitin et al. (2007), which considered the signals (peak areas) obtained for each tin isotope as a linear combination of the different contributing sources. The uncertainty on the amount of Cr(VI) extracted by each sample was calculated by the Kragten spreadsheet approach.

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238 **2.6. Statistical analyses**

Box-plots, descriptive statistics and Pearson's correlation analysis (2-tails) were performed by the software IBM SPSS Statistics 20 (Chicago, IL, USA). Statistical differences were assumed at p < 0.05.

242

243 **3. RESULTS AND DISCUSSION**

244 3.1. Cr(VI) quantification in soil samples from case study A

245 *3.1.1. Preliminary investigations*

The potential of the soil from case study A to oxidise Cr(III) to Cr(VI), assessed on 10 samples by 246 the standard Cr net oxidation test (Bartlett and James, 1996), was found to be very low (max: 0.4 247 µmol per 2 g of tested soil). According to the test guidelines and Italian regulations on the 248 application of sewage sludge, a soil can be considered at risk of Cr(VI) formation when the amount 249 of the formed-Cr(VI) obtained by the standard Cr net oxidation test is higher than 1 µmol per 2 g of 250 tested soil. The oxidation of Cr(III) to Cr(VI) in a soil can be essentially attributed to the presence 251 of easily reducible manganese oxides (Palmer and Puls, 1994). The number of potentially 252 oxidisable moles of Cr(III) by the manganese oxides present in the soil was calculated and ranged 253 from 13 to 31 μ g g⁻¹. However, it should be stated that these calculated values might not represent 254 the real condition in the soil, since the reduced manganese oxides can be re-oxidised and therefore 255 become new reducing substrates. Also the new formed-Cr(VI) can be reduced by other soil 256 components (Bartlett, 1991). In any case, some authors find a significant correlation in soil between 257 the amounts of easily reducible manganese oxides and the soil capacity to oxidise Cr(III) (Chon et 258 al., 2008). In our soil samples we did not observe this correlation (data not shown). 259

USEPA 3060A and 7196A methods are widely used for the determination of Cr(VI) in soil samples (Weibel et al., 2016), however, they were not able to provide reliable Cr(VI) concentrations in soil samples from case study A. In fact, the recovery of the Cr(VI) spike added to the soil was low (about 50 %) (Table 2), even when the Cr(VI) spike was added to the extractant solution during the procedure and not to the soil (data not shown). This was probably due to the reduction of Cr(VI) to

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Cr(III) induced by the organic matter at the moment of extract acidification. The modification of the 265 official procedure by cold extraction or by the purification of the extract from organic matter by 266 means of aluminium sulphate led to an improvement in the recovery of the Cr(VI) spike in the 267 samples previously analysed with the official procedure, but not always reach satisfactory results 268 and this was especially true when the organic matter content of the samples increases (Table 2). 269 Also, when applying cold extraction the risk of an incomplete Cr(VI) extraction from the soil 270 increases. Even when USEPA 7199 was applied after USEPA 3060A extraction the recovery of the 271 spike was unsatisfactory in most cases (spike reduction between 28 % and 96 %) (Table 2). The 272 extent of spike reduction appeared higher when the soil organic matter content increased, 273 reinforcing the hypothesis that organic compounds of the soil from case study A were the main 274 electron donors readily reducing Cr(VI) to Cr(III). However, despite the underestimation of the real 275 levels of Cr(VI) in the soils of case study A, concentrations of Cr(VI) between 4 and 14 μ g g⁻¹ were 276 found in the most contaminated samples. These values were higher than the legal threshold of 2 µg 277 g^{-1} set by Italian Parliament for residential/urban soil (Italian-Parliament, 2006). 278

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280 3.1.2. Cr(VI) quantification by single-spike IDMS

According to the preliminary results obtained using the USEPA methods, an alternative method 281 based on IDMS was tested in the soil samples from case study A. Table 3 shows the concentration 282 of Cr(VI) (µg g⁻¹) obtained by species-specific single-spiking IDMS. The 27 analysed soil samples 283 were classified in two different groups. The first group includes 20 soil samples (A1-A20), in which 284 Cr(VI) quantification by IDMS was expected to be accurate due of the lower levels of spike 285 reduction and did not require any soil pre-treatment. The second group comprises 7 soil samples 286 (A21-A27), mostly from pedological profiles and trenches most-contaminated by pseudo-total Cr. 287 in which Cr(VI) quantification by IDMS was only possible after a soil pre-treatment with n-hexane 288 to remove reducing organic compounds. 289

In Italy soil quality standards for agricultural areas have not yet been established but the Italian 290 Parliament has established screening values to assess potential contamination of residential/urban 291 soils (Italian-Parliament, 2006). The amount of Cr(VI) extracted from the first group of samples 292 (A1-A20) ranged from 0.15 to 11.18 μ g g⁻¹ (median 2.98 μ g g⁻¹), with 70 % of samples exceeding 293 the Cr(VI) screening value (2 μ g g⁻¹) and 85 % the pseudo-total Cr screening value (150 μ g g⁻¹) set 294 by Italian Parliament for residential/urban soil (Italian-Parliament, 2006) with concentrations up to 295 6,831 μ g g⁻¹. Although the neutral to moderate-alkaline pH (7.4-8.0) and the sandy-loam texture of 296 297 the soil may favour the occurrence of Cr(VI) in the soil, the medium to high content in organic matter (13-56 mg g⁻¹; Table 3) strongly promotes the reduction of the mobile Cr(VI) to the more 298 thermodynamically stable and insoluble Cr(III). Part of this organic matter consists of heavy 299 hydrocarbons (C>12), likely derived from leather-based industrial wastes illegally buried into the 300 soil. In fact, the 80 % of the A1-A20 samples are potentially contaminated by heavy hydrocarbons 301 (C>12) with concentrations exceeding the C>12 screening value (50 μ g g⁻¹) set by Italian 302 Parliament for residential/urban soil (Italian-Parliament, 2006). The Cr(VI) extracted from the A1-303 A20 soil samples is present in concentrations higher than $2 \mu g g^{-1}$ and positively correlated with 304 their own pseudo-total Cr content (r=0.828; Table 4). However, Cr(VI) represents less than 1 % of 305 the pseudo-total Cr in each of A1-A20 samples (Table 3 and Figure 1). Therefore, assuming a 306 negligible occurrence of Cr(0), pseudo-total Cr in the soil is predominantly or almost exclusively 307 present as Cr(III), namely the less toxic and immobile Cr species. Similar results were also 308 observed by Moreira et al. (2018), who proved that Cr(VI) was always less than 0.2 % (0.7-2.0 µg 309 g^{-1}) in contaminated soils of Amazon forest containing up to 2,346 µg of pseudo-total Cr per g of 310 soil. During microwave-assisted extraction, both organic matter [acting as an electron donor (Tian 311 et al., 2010)] and, even more, heavy hydrocarbons (C>12) resulted to be powerful reducing agents 312 of ⁵⁰Cr-enriched Cr(VI) spike. The reduction of the ⁵⁰Cr-enriched Cr(VI) spike occurred during 313 extraction (Table 3, A1-A20), was basically moderate and significantly correlated to organic matter 314 (r=0.605; Table 4) and heavy hydrocarbons (r=0.780; Table 4). In any case, Cr(VI) reduction is 315

corrected by the isotope dilution methodology applied. The addition of EDTA to the extractant 316 solution enabled the formation of a stable complex - with the two most important cationic 317 contaminants of the case study A [Cr(III) and Zn(II)]. Indeed, the formation of the Cr(III)-EDTA 318 was evident in all chromatograms acquired during the LC-ICP-MS running. Figures SM1 and SM2 319 in the Supplementary Material also shows that EDTA forms a stable complex with Fe(III) detected 320 at m/z 54 but its interference with ⁵⁴Cr is chromatographically resolved. Zinc complexation by 321 EDTA was also desirable to correctly quantify Cr(VI) in these soils, since it forms chromates 322 relatively insoluble in water (Saha et al., 2011). Differences in relative abundance of Cr(VI) (% of 323 the pseudo-total content) in A1-A20 soil samples - collected at various depths (d1, d2 and d3) and 324 from pedological profiles and trenches (pt, depth ranging from 30 to 180 cm) - have been found 325 (Figure 1). The most oxidised soil layers [0-20 cm (d1), 30-60 cm (d2)] showed wider and greater 326 boxes and upper whiskers in comparison to the deeper and less oxidised soil layer [70-90 cm (d3)]. 327 328 This indicates a higher Cr(VI) content in the more aerated soil horizons, probably due to a greater probability of re-oxidation of manganese oxides involved in the chromium cycle in the soil 329 330 (Bartlett, 1991). As observed in Figure 1, the box of Cr(VI) (% of the pseudo-total content) of the 331 samples collected from pedological profiles and trenches (pt; A1, A5-A7, A13, A20), although shows the highest median, is narrower and lower than d1 and d2 boxes, probably because the 332 abundance of organic matter in these samples strongly promotes the reduction of Cr(VI) to Cr(III), 333 thus minimising the presence of the toxic and mobile anion in the soil (Antoniadis et al., 2017; 334 Kalcikova et al., 2016; Xiao et al., 2013). Indeed, Zhang et al. (2017) stated that humic substances 335 can limit the mobility of Cr(VI) in the soil by complexation-coupled reduction mechanisms. On the 336 other hand, Wu et al. (2017) demonstrated that humic substances from cattle manure reduced 337 Cr(VI) in the soil, then the produced Cr(III) formed outer sphere complexes with –OH and inner 338 sphere complex with carboxyl groups of humic acids. 339

In contrast to the A1-A20 soil samples, the quantification of Cr(VI) by IDMS in the second group of soil samples (A21-A27) was done after a soil pre-treatment with n-hexane, which removed part

of the abundant Cr(VI)-reducing organic compounds (see Table 3). These samples were mainly 342 collected from pedological profiles and trenches and were most-contaminated by pseudo-total Cr 343 $(1,314-10,046 \ \mu g \ g^{-1})$. The C>12 content in this samples ranged from 509 to 2,150 $\ \mu g \ g^{-1}$ and they 344 were characterised by a high content in organic matter (47-287 mg g^{-1}). The complexity and 345 abundance of organic compounds in these samples caused the complete reduction (~99 %) of the 346 ⁵⁰Cr-enriched Cr(VI) spike during the microwave-assisted extraction (at 80 °C for 5 min). Our first 347 attempts to stabilise the organic compounds, by adding 0.5 g of Alumina, Silica gel or C18 bulk 348 resin or lowering the extraction time (from 5 to 2.5 min) and temperature (30, 40, 50, 60 or 70 °C), 349 did not provide any improvement. For instance, increasing amounts of Cr(VI) were extracted from 350 soil sample A21 up to 70°C, but contextually at 70°C the overall reduction of the ⁵⁰Cr-enriched 351 Cr(VI) spike raised to ~92 % (Figure SM3, Supplementary Material). A soil pre-treatment before 352 the microwave-assisted extraction (at 80 °C for 5 min) with n-hexane aiming to partially remove 353 heavy hydrocarbons from soil, namely the most powerful Cr(VI) reducing agents, allowed us to 354 quantify the Cr(VI) in the Cr-most-contaminated soil samples A21-A27 (Table 3). Even though the 355 reduction of the ⁵⁰Cr-enriched Cr(VI) spike was on average 98.4 %, Cr(VI) could be quantified in 356 357 these samples with low uncertainties (0.1-17 % of the mean value, except A24). Although the relative abundance of Cr(VI) (% of the pseudo-total content) is always lower than 2 %, the Cr(VI) 358 extracted by these 7 soil samples is very high, ranging from 22.0 to 107.1 μ g g⁻¹ (median 40.7 μ g g⁻¹ 359 ¹: Table 3, A21-A27), with 100 % of samples exceeding the Cr(VI) screening value of 2 μ g g⁻¹ set 360 by Italian legislation for residential/urban soil (Italian-Parliament, 2006). 361

362

363 **3.2.** Cr(VI) quantification in soil samples from case study B

The concentration ($\mu g g^{-1}$) of Cr(VI) in the 12 soil samples from case study B, quantified by singlespike IDMS, is given in Table 5. The Cr(VI) amount extracted from these samples ranges from 0.70 to 5.79 $\mu g g^{-1}$ (median 1.81 $\mu g g^{-1}$), with 42 % of samples exceeding the Cr(VI) screening value of 2 $\mu g g^{-1}$ set by Italian legislation for residential/urban soil (Italian-Parliament, 2006). This is the first

evidence of the Cr(VI) occurrence in these widely investigated soils. Table 6 shows that the amount 368 of Cr(VI) significantly increases with pseudo-total Cr content (r=0.984). However, it represents less 369 than 1 % of the pseudo-total Cr (Table 5). Therefore, in case study B, the pseudo-total Cr is almost 370 exclusively present as Cr(III), namely the less toxic and immobile Cr species. The pseudo-total Cr 371 determined in 100 % of the samples of case study B exceeds the screening value (150 μ g g⁻¹) set by 372 Italian Parliament for residential/urban soil (Italian-Parliament, 2006) (Table 5), with concentrations 373 up to 594 µg g⁻¹. According to Adamo et al. (2003), soils of Solofrana valley are neutral (soil pH 374 around 7.0) and fine to medium textured sandy-silty or clay-sandy soils with 50-60 % of the soil in 375 the clay-silt fraction. These soils are also characterised by a high content in organic matter, 376 conceivably raised by the periodic depositions of the river sediments. Indeed the 12 analysed soils 377 show an organic matter content ranging from 70 to 83 mg g^{-1} (median 75 mg g^{-1} ; Table 5), 378 particularly high for Mediterranean soils. The presence of this considerable pool of organic matter, 379 acting as Cr(VI)-reducing agent (Barajas-Aceves et al., 2007; Jardine et al., 1999), justifies the low 380 relative abundance of Cr(VI) (<1 % of the pseudo-total content; Table 5). The low occurrence of 381 Cr(VI) (at least as % the pseudo-total content) is also in part due the low potential of these soils to 382 oxidise Cr(III) to Cr(VI), as assessed by Adamo et al. (2003) through the standard Cr net oxidation 383 test proposed by Bartlett and James (1996). Extracting these soils by BCR (Community Bureau of 384 Reference, European Commission) sequential extraction procedure (Ure et al., 1993), Adamo et al. 385 (2003; 2006) also demonstrated that the majority of the pseudo-total Cr is associated to the organic 386 substances, which easily complex and stabilise the Cr(III) in the soil (Gustafsson et al., 2014). 387

The high organic matter content in the analysed soils (Table 5) caused a significant reduction of the 50 Cr-enriched Cr(VI) spike during the microwave-assisted extraction. The reduction factor obtained in the analysis of these set of samples ranged from 19 to 46 %. Nevertheless the quantification of Cr(VI) by IDMS was characterised by a low uncertainty in all the samples (Table 5). In contrast to case study A, 50 Cr-enriched Cr(VI) spike reduction during extraction was not significantly correlated with organic matter content in soil samples (r=0.507; Table 6). This means that, instead

of the organic matter content, ⁵⁰Cr-enriched Cr(VI) reduction was affected by the complexity, the
 heterogeneity and the quality of the organic compounds in the soil samples.

396

397 **4.** Conclusions

Chromium speciation in soil is of paramount importance to assess the environmental and human 398 health risks of potentially contaminated sites and to select and apply appropriate management and 399 remediation strategies. This study investigated the extent of Cr(VI) occurrence in two Italian 400 agricultural soils potentially contaminated by tannery-deriving Cr. The application of IDMS as 401 calibration technique ensured the accurate Cr(VI) quantification in soil samples. The use of 402 EDTA at pH 10 as extractant avoids Cr(III) oxidation and enables the separation of Cr species by 403 anion exchange chromatography. In both case studies and in all of the analysed samples, it was 404 found that the presence of medium-high level of organic matter in soil keeps the relative abundance 405 406 (% of the pseudo-total content) of Cr(VI), below 2 %. However, in most samples the amount of Cr(VI) was found to be above the Italian screening value of 2 µg g⁻¹, suggesting that the toxic and 407 408 carcinogenic anion may represent a potential risk for the surrounding environment and for human 409 health. This is particularly important for case study B, as it is the first evidence of Cr(VI) occurrence in soils of Solofrana valley irrigated or flooded by river waters. 410

The chemical complexity of the most contaminated soil samples from case study A, a farmland 411 potentially contaminated by pseudo-total Cr and Zn and heavy hydrocarbons, made the 412 quantification of Cr(VI) very challenging for all the available analytical techniques; the similar 413 critical issues in the extraction and quantification of Cr(VI) faced by classical USEPA 7196A and 414 7199 methods and innovative IDMS method probably indicate a redox potential of these soil 415 samples that does not allow a high occurrence of Cr(VI) in the soil. Further investigations are 416 417 probably necessary considering a direct quantification of the Cr(VI) in the solid soil matrix of the Cr-most-contaminated samples by X-ray absorption near edge structure spectroscopy (XANES). 418

Lately, the findings of this study suggest that the maintenance of a medium-high content of organic matter in soil, promoting Cr(VI) reduction and lowering its bioavailability, along with no-tillage (i.e., no cyclic soil aeration) and possible supply of elemental sulphur [a Cr(VI)-reducing agent in soils impacted by leather tanneries (Shi et al., 2016)], might be appropriate actions to minimise the Cr(VI) occurrence in potentially contaminated soils as those investigated in this paper. The monitoring over time of the fate and stability of Cr(VI) in soil is also essential to manage and minimise the risks for environment and human health.

426

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435 **References**

Adamo, P., Denaix, L., Terribile, F., Zampella, M., 2003. Characterization of heavy metals in
contaminated volcanic soils of the Solofrana river valley (southern Italy). Geoderma 117, 347366.

Adamo, P., Zampella, M., Gianfreda, L., Renella, G., Rutigliano, F.A., Terribile, F., 2006. Impact
of river overflowing on trace element contamination of volcanic soils in South Italy: Part I. Trace
element speciation in relation to soil properties. Environ Pollut 144, 308-316.

- Antoniadis, V., Polyzois, T., Golia, E.E., Petropoulos, S.A., 2017. Hexavalent chromium
 availability and phytoremediation potential of Cichorium spinosum as affect by manure, zeolite
 and soil ageing. Chemosphere 171, 729-734.
- 445 Avudainayagam, S., Megharaj, A., Owens, G., Kookana, R.S., Chittleborough, D., Naidu, R., 2003.
- 446 Chemistry of chromium in soils with emphasis on tannery waste sites. Rev Environ Contam T
- 447 178, 53-91.
- Banks, M.K., Schwab, A.P., Henderson, C., 2006. Leaching and reduction of chromium in soil as
 affected by soil organic content and plants. Chemosphere 62, 255-264.
- 450 Barajas-Aceves, M., Corona-Hernandez, J., Rodriguez-Vazquez, R., 2007. Chromium fractionation
- 451 in semi-arid soils amended with chromium and tannery sludge. J Hazard Mater 146, 91-97.
- Bartlett, R.J., 1991. Chromium Cycling in Soils and Water Links, Gaps, and Methods. Environ
 Health Persp 92, 17-24.
- 454 Bartlett, R.J., James, B.R., 1996. Chromium. In: Sparks, D.L. (Ed.), Methods of Soil Analysis. Part
- 455 3-Chemical Methods. SSSA Book Series, vol. 5. SSSA–ASA, Madison, WI, USA, pp. 683–701.
- Becquer, T., Quantin, C., Sicot, M., Boudot, J.P., 2003. Chromium availability in ultramafic soils
 from New Caledonia. Sci Total Environ 301, 251-261.
- Bolan, N.S., Adriano, D.C., Natesa, R., Koo, B.J., 2003. Effects of organic amendments on the
 reduction and phytoavailability of chromate in mineral soil. J Environ Qual 32, 120-128.
- 460 Caporale, A.G., Adamo, P., Capozzi, F., Langella, G., Terribile, F., Vingiani, S., 2018. Monitoring
- 461 metal pollution in soils using portable-XRF and conventional laboratory-based techniques:
- 462 Evaluation of the performance and limitations according to metal properties and sources. Sci Total463 Environ 643, 516-526.
- 464 Chon, C.M., Kim, J.G., Lee, G.H., Kim, T.H., 2008. Influence of extractable soil manganese on
- 465 oxidation capacity of different soils in Korea. Environ Geol 55, 763-773.

- 466 Choppala, G., Bolan, N., Lamb, D., Kunhikrishnan, A., 2013. Comparative sorption and mobility of
- 467 Cr(III) and Cr(VI) species in a range of soils: implications to bioavailability. Water Air Soil Poll
 468 224.
- 469 Choppala, G., Kunhikrishnan, A., Seshadri, B., Park, J.H., Bush, R., Bolan, N., 2018. Comparative
- sorption of chromium species as influenced by pH, surface charge and organic matter content in
- 471 contaminated soils. J Geochem Explor 184, 255-260.
- 472 D'Ascoli, R., Rao, M.A., Adamo, P., Renella, G., Landi, L., Rutigliano, F.A., Terribile, F.,
- 473 Gianfreda, L., 2006. Impact of river overflowing on trace element contamination of volcanic soils
- 474 in South Italy: Part II. Soil biological and biochemical properties in relation to trace element
 475 speciation. Environ Pollut 144, 317-326.
- 476 Di Gennaro, A., Aronne, G., De Mascellis, R., Vingiani, S., Sarnataro, M., Abalsamo, P., Cona, F.,
- Vitelli, L., Arpaia, G., 2002. I sistemi di terre della Campania. Monografia e carta 1:250.000, con
 legenda. S.E.L.C.A., Firenze, pp. 63.
- Di Vito, M.A., Castaldo, N., de Vita, S., Bishop, J., Vecchio, G., 2013. Human colonization and
 volcanic activity in the eastern Campania Plain (Italy) between the Eneolithic and Late Roman
 periods. Quatern Int 303, 132-141.
- 482 Fabregat-Cabello, N., Rodriguez-Gonzalez, P., Castillo, A., Malherbe, J., Roig-Navarro, A.F.,
- Long, S.E., Alonso, J.I.G., 2012. Fast and accurate procedure for the determination of Cr(VI) in
- solid samples by isotope dilution mass spectrometry. Environ Sci Technol 46, 12542-12549.
- 485 FAO, 1998. World reference base for soil resources, Food And Agriculture Organization of the
- 486 United Nations, Rome, 84 World Soil Resources Reports International Society of Soil Science.
- 487 Garcia Alonso, J.I., Rodriguez-Gonzalez, P., 2013. Isotope dilution mass spectrometry. ISBN: 978-
- 488 1-84973-333-5, RSC Publishing, Cambridge, UK.
- 489 Guidotti, L., Abad, S.Q., Rodriguez-Gonzalez, P., Alonso, J.I.G., Beone, G.M., 2015.
- 490 Quantification of Cr(VI) in soil samples from a contaminated area in northern Italy by isotope
- dilution mass spectrometry. Environ Sci Pollut R 22, 17569-17576.

- Gustafsson, J.P., Persson, I., Oromieh, A.G., van Schaik, J.W.J., Sjostedt, C., Kleja, D.B., 2014.
 Chromium(III) complexation to natural organic matter: mechanisms and modeling. Environ Sci
 Technol 48, 1753-1761.
- IARC, 2012. International Agency for Research on Cancer Chromium, Nickel and Welding, IARC
 Monographs on the Education of Carcinogenic Risks to Huamans, 49 World Health
 Organizationm Lyon, France.
- ISO11466, 1995. Soil quality Extraction of trace elements soluble in aqua regia. First edition,
 ISO/TC 190/SC 3, 03-1995.
- 500 Italian-Parliament, 2006. Decreto legislativo 3 aprile 2006, n. 152. Norme in materia ambientale
- 501 (Gazzetta Ufficiale della Repubblica Italiana n. 88, Supplemento Ordinario no. 96, 14 aprile
 502 2006).
- Jacobs, J.A., Testa, S.M., 2005. Overview of chromium(VI) in the environment: background and
 history. In: Chromium(VI) handbook. New York, CRC Press, pp. 1–21.
- James, B.R., Petura, J.C., Vitale, R.J., Mussoline, G.R., 1995. Hexavalent Chromium Extraction
 from Soils a Comparison of 5 Methods. Environ Sci Technol 29, 2377-2381.
- Jardine, P.M., Fendorf, S.E., Mayes, M.A., Larsen, I.L., Brooks, S.C., Bailey, W.B., 1999. Fate and
- transport of hexavalent chromium in undisturbed heterogeneous soil. Environ Sci Technol 33,
 2939-2944.
- Kalcikova, G., Zupancic, M., Jemec, A., Gotvajn, A.Z., 2016. The impact of humic acid on
 chromium phytoextraction by aquatic macrophyte Lemna minor. Chemosphere 147, 311-317.
- 512 Korolczuk, M., Grabarczyk, M., 2005. Evaluation of ammonia buffer containing EDTA as an
- extractant for Cr(VI) from solid samples. Talanta 66, 1320-1325.
- Kotas, J., Stasicka, Z., 2000. Chromium occurrence in the environment and methods of its
 speciation. Environ Pollut 107, 263-283.
- Leita, L., Margon, A., Pastrello, A., Arcon, I., Contin, M., Mosetti, D., 2009. Soil humic acids may
- favour the persistence of hexavalent chromium in soil. Environ Pollut 157, 1862-1866.

- Lilli, M.A., Moraetis, D., Nikolaidis, N.P., Karatzas, G.P., Kalogerakis, N., 2015. Characterization
- and mobility of geogenic chromium in soils and river bed sediments of Asopos basin. J HazardMater 281, 12-19.
- 521 Ministerial-Decree, 1999. Metodi ufficiali di analisi chimica del suolo. Metodo V.2 522 Determinazione del calcio carbonato attivo, 13/09/1999.
- 523 Moreira, L.J.D., da Silva, E.B., Fontes, M.P.F., Liu, X., Ma, L.Q., 2018. Speciation, bioaccessibility
- and potential risk of chromium in Amazon forest soils. Environ Pollut 239, 384-391.
- Mpouras, T., Chrysochoou, M., Dermatas, D., 2017. Investigation of hexavalent chromium sorption
 in serpentine sediments. J Contam Hydrol 197, 29-38.
- 527 Orsi, G., Di Vito, M.A., Isaia, R., 2004. Volcanic hazard assessment at the restless Campi Flegrei
 528 caldera. B Volcanol 66, 514-530.
- Palmer, C.D., Puls, R.W., 1994. Natural attenuation of hexavalent chromium in groundwater and
 soils. Office of Research and Development, USEPA/540/5-94/505.
- Pettine, M., Capri, S., 2005. Digestion treatments and risks of Cr(III)-Cr(VI) interconversions
 during Cr(VI) determination in soils and sediments a review. Anal Chim Acta 540, 231-238.
- 533 Pinel-Raffaitin, P., Rodriguez-Gonzalez, P., Ponthieu, M., Amouroux, D., Le Hecho, I., Mazeas, L.,
- Donard, O.F.X., Potin-Gautier, M., 2007. Determination of alkylated tin compounds in landfill
 leachates using isotopically enriched tin species with GC-ICP-MS detection. J Anal Atom
 Spectrom 22, 258-266.
- Rahman, G.M.M., Kingston, H.M.S., Towns, T.G., Vitale, R.J., Clay, K.R., 2005. Determination of
 hexavalent chromium by using speciated isotope-dilution mass spectrometry after microwave
 speciated extraction of environmental and other solid materials. Anal Bioanal Chem 382, 11111120.
- 541 Rauret, G., Lopez-Sanchez, J.F., Sahuquillo, A., Rubio, R., Davidson, C., Ure, A., Quevauviller, P.,
- 542 1999. Improvement of the BCR three step sequential extraction procedure prior to the certification
- 543 of new sediment and soil reference materials. J Environ Monitor 1, 57-61.

- Saha, R., Nandi, R., Saha, B., 2011. Sources and toxicity of hexavalent chromium. J Coord Chem
 64, 1782-1806.
- Ščančar, J., Milačič, R., 2014. A critical overview of Cr speciation analysis based on high
 performance liquid chromatography and spectrometric techniques. J Anal Atom Spectrom 29,
 427-443.
- Séby, F., Vacchina, V., 2018. Critical assessment of hexavalent chromium species from different
 solid environmental, industrial and food matrices. Trends in Analytical Chemistry 104, 54-68.
- 551 Shahid, M., Shamshad, S., Rafiq, M., Khalid, S., Bibi, I., Niazi, N.K., Dumat, C., Rashid, M.I.,
- 552 2017. Chromium speciation, bioavailability, uptake, toxicity and detoxification in soil-plant
- system: A review. Chemosphere 178, 513-533.
- 554 Shi, J.J., Chen, H.L., Arocena, J.M., Whitcombe, T., Thring, R.W., Memiaghe, J.N., 2016.
- Elemental sulfur amendment decreases bio-available Cr-VI in soils impacted by leather tanneries.
 Environ Pollut 212, 57-64.
- Tchounwou, P.B., Yedjou, C.G., Patlolla, A.K., Sutton, D.J., 2012. Heavy metal toxicity and the
 environment. EXS 101, 133-164.
- 559 Tian, X.F., Gao, X.C., Yang, F., Lan, Y.Q., Mao, J.D., Zhou, L.X., 2010. Catalytic role of soils in
- the transformation of Cr(VI) to Cr(III) in the presence of organic acids containing alpha-OH groups. Geoderma 159, 270-275.
- UNI-EN-ISO-16703, 2011. Soil quality Determination of content of hydrocarbon in the range C10
 to C40 by gas chromatography. ICS 13.080.10, July 21, 2011.
- Ure, A.M., Quevauviller, P., Muntau, H., Griepink, B., 1993. Speciation of heavy-metals in soils
 and sediments an account of the improvement and harmonization of extraction techniques
 undertaken under the auspices of the Bcr of the Commission-of-the-European-Communities. Int J
 Environ an Ch 51, 135-151.
- 568 USEPA, 1992. Method 7196A. Chromium, Hexavalent (Colorimetric), SW-846 Third Edition,
- 569 Update V. U.S. Government Printing Office, Washington DC.

- USEPA, 1995. Alkaline Digestion of Hexavalent Chromium. Test Methods for Evaluating Solid 570
- Waste, Physical/Chemical Methods, SW-846, 3rd edition. U.S. Environmental Protection Agency: 571 Washington, DC, USA.
- 572
- USEPA, 1996. Method 7199: Determination of hexavalent chromium in drinking water, 573
- groundwater and industrial wastewater effluents by ion chromatography. By USEPA and the 574
- American Society for Testing and Materials (ASTM). 575
- USEPA, 2007. Method 6800: Elemental and Speciated Isotope Dilution Mass Spectrometry. U.S. 576
- Environmental Protection Agency: Washington, DC, USA. 577
- Walkley, A., Black, I.A., 1937. An examination of Degtjareff method for determining soil organic 578
- matter and a proposed modification of the chromic acid titration method. Soil Sci 37, 29-37. 579
- Weibel, G., Waber, H.N., Eggenberger, U., Mader, U.K., 2016. Influence of sample matrix on the 580
- alkaline extraction of Cr(VI) in soils and industrial materials. Environ Earth Sci 75. 581
- 582 Wu, M., Li, G.X., Jiang, X.L., Xiao, Q.Q., Niu, M.X., Wang, Z.Y., Wang, Y.Y., 2017. Non-
- biological reduction of Cr(VI) by reacting with humic acids composted from cattle manure. Rsc 583 584 Adv 7, 26903-26911.
- Xiao, W.D., Yang, X.E., He, Z.L., Rafiq, M.T., Hou, D.D., Li, T.Q., 2013. Model for evaluation of 585
- the phytoavailability of chromium (Cr) to Rice (Oryza sativa L.) in representative Chinese soils. J 586 Agr Food Chem 61, 2925-2932. 587
- Zampella, M., Adamo, P., Caner, L., Petit, S., Righi, D., Terribile, F., 2010. Chromium and copper 588 in micromorphological features and clay fractions of volcanic soils with andic properties. 589 Geoderma 157, 185-195. 590
- Zhang, J., Chen, L.P., Yin, H.L., Jin, S., Liu, F., Chen, H.H., 2017. Mechanism study of humic acid 591
- functional groups for Cr(VI) retention: Two-dimensional FTIR and C-13 CP/MAS NMR 592 correlation spectroscopic analysis. Environ Pollut 225, 86-92. 593
- Zuliani, T., Scancar, J., Milacic, R., 2013. The use of stable isotopes for Cr(VI) determination in 594
- silty-clay soil solution. Anal Bioanal Chem 405, 7231-7240. 595

Figure 1.



Isotope	Abundance (%)
50	96.10 ± 0.23
52	3.82 ± 0.19
53	0.08 ± 0.04
54	Not detected
Cr(VI) conce	ntration (µg g ⁻¹)
134.8	23 ± 3.44

Table 1. Isotopic composition (% abundance) and concentration of the ⁵⁰Cr-enriched Cr(VI) spike solution employed in this work.

	Extraction/determination methods								
	<mark>US</mark> 3060A	EPA /7196A	USEPA 3060A [Al ₂ (SO ₄) ₃ purification]/7196A		USEPA 30 extractio	USEPA 3060A (cold extraction)/7196A		USEPA 3060A/7199	
Sample ID	Cr(VI)	Spike reduction	Cr(VI)	Spike reduction	Cr(VI)	Spike reduction	Cr(VI)	Spike reduction	
	$(\mu g g^{-1})$	(%)	(µg g ⁻¹)	(%)	(µg g ⁻¹)	(%)	(µg g ⁻¹)	(%)	
A2 (d2)	1.7 ± 0.1	46 <u>± 1</u>	1.0 ± 0.1	24 ± 1	0.5 ± 0.1	10 ± 1			
A13 (pt)							10.5 ± 0.4	83 ± 3	
A16 (d3)	4.0 ± 0.2	52 <u>±</u> 2	4.9 ± 0.2	44 ± 2	2.2 ± 0.1	21 ± 1	7.1 ± 0.3	28 ± 1	
A18 (d3)							8.5 ± 0.1	42 ± 1	
A20 (pt)							13.9 ± 0.6	88 ± 3	
A27 (pt)					(8.9 ± 0.1	96 ± 2	
A28 (d2)*					3.2 ± 0.2	62 <u>±</u> 2	5.2 ± 0.2	89 ± 4	

Table 2. Preliminary Cr(VI) quantification in selected soil samples from case study A by USEPA 3060A extraction and colorimetric-based USEPA 7196A and USEPA 7199 methods.

* Pseudo-total Cr content = $1458 \pm 30 \ \mu g \ g^{-1}$, organic matter content = $49 \pm 3 \ m g \ g^{-1}$. Pseudo-total Cr and organic matter contents of the A2, A13, A16, A18, A20 and A27 soil samples are shown in Table 3.

--- Analysis not performed

d2 and d3 indicate soil samples collected at 30-60 cm and 70-90 cm of depth, respectively; pt indicates soil samples collected from pedological profiles and trenches opened in the field.

Table 3. Pseudo-total ($\mu g g^{-1}$) and hexavalent ($\mu g g^{-1}$; % of the pseudo-total) Cr, organic matter (mg g⁻¹) and heavy hydrocarbons (HC C>12, $\mu g g^{-1}$) contents in soil samples from case study A. Samples A21-A27 were pre-treated with n-hexane, respectively, before microwave-assisted extraction. The reduction (%) of the ⁵⁰Cr(VI)-enriched spike after microwave-assisted extraction is shown as well.

Sample ID	Pseudo-total Cr	Cr(VI)		Spike reduction	Organic matter	HC C>12
	(µg g ⁻¹)	$(\mu g g^{-1})$	(% of tot)	(%)	(mg g ⁻¹)	$(\mu g \ g^{-1})$
A1 (pt)	48 ± 1	0.15 ± 0.10	0.31 ± 0.22	1.6 ± 0.5	13 ± 1	20 ± 5
A2 (d2)	103 ± 4	1.04 ± 0.26	1.01 ± 0.26	2.1 ± 0.2	14 ± 1	19 ± 7
A3 (d1)	136 ± 3	0.99 ± 0.11	0.73 ± 0.08	50 ± 17	21 ± 1	27 ± 8
A4 (d1)	298 ± 8	2.30 ± 0.27	0.77 ± 0.09	10 ± 1	19 ± 2	45 ± 9
A5 (pt)	331 ± 8	1.97 ± 0.10	0.59 ± 0.03	20 ± 2	54 ± 3	<u>68</u> ± 11
A6 (pt)	<u>426</u> ± 9	2.36 ± 0.27	0.55 ± 0.06	17 ± 3	36 ± 2	<u>73</u> ± 10
A7 (pt)	<u>454</u> ± 11	2.73 ± 0.10	0.60 ± 0.02	16 ± 3	34 ± 2	143 ± 23
A8 (d1)	<u>546</u> ± 13	1.65 ± 0.34	0.30 ± 0.06	8.6 ± 0.8	26 ± 1	<u>69</u> ± 12
A9 (d2)	955 ± 24	3.48 ± 0.11	0.36 ± 0.01	23 ± 14	23 ± 1	$\underline{200} \pm 29$
A10 (d3)	<u>960 ± 22</u>	5.25 ± 0.34	0.55 ± 0.04	44 ± 8	43 ± 3	<u>307</u> ± 44
A11 (d1)	$\underline{1096} \pm 27$	2.74 ± 0.10	0.25 ± 0.01	11 ± 1	42 ± 2	<u>59</u> ± 11
A12 (d2)	$\underline{1161} \pm 29$	5.45 ± 0.36	0.47 ± 0.03	2.8 ± 0.2	26 ± 1	$\underline{106} \pm 16$
A13 (pt)	1230 ± 25	4.87 ± 0.12	0.40 ± 0.01	82 ± 2	55 ± 3	680 ± 81
A14 (d1)	$\underline{1413} \pm 34$	3.21 ± 0.11	0.23 ± 0.01	11 ± 2	34 ± 2	<u>79</u> ± 13
A15 (d3)	1713 ± 45	1.31 ± 0.25	0.08 ± 0.01	17.9 ± 0.5	22 ± 1	<u>60</u> ± 11
A16 (d3)	<u>2372</u> ± 57	7.08 ± 0.11	0.30 ± 0.01	13 ± 2	20 ± 1	<u>93</u> ± 15
A17 (d3)	2802 ± 73	6.81 ± 0.62	0.24 ± 0.02	7 ± 1	34 ± 2	$\underline{218} \pm 32$
A18 (d3)	3456 ± 85	6.28 ± 0.12	0.18 ± 0.01	11 ± 6	27 ± 1	<u>162</u> ± 24
A19 (d2)	3572 ± 92	$\underline{11.18}\pm0.26$	0.31 ± 0.01	6 ± 6	33 ± 2	158 ± 24
A20 (pt)	$\underline{6831} \pm 158$	9.45 ± 0.11	0.14 ± 0.01	57 ± 4	56 ± 3	331 ± 42
Mean	1495	4.02	0.42	21	32	146
Min	48	0.15	0.08	2	13	19
P25	402	1.89	0.25	8	22	60
Median	1028	2.98	0.34	12	30	86
P75	1878	5.66	0.56	20	38	172
Max	6831	11.18	1.01	82	56	680
IT sc. val. [*]	<u>150</u>	<u>2</u>				<u>50</u>
Sam>ITs.v.	85%	70%				80%
A21 (d2)	1314 ± 31	<u>22.5</u> ± 2.1	1.71 ± 0.16	97.7 ± 2.6	47 ± 3	710 ± 98
A22 (d2)	<u>1946</u> ± 49	<u>22.0</u> ± 3.1	1.13 ± 0.16	98.3 ± 3.2	80 ± 5	<u>509</u> ± 72
A23 (d3)	3857 ± 97	26.2 ± 2.0	0.68 ± 0.05	97.5 ± 2.8	177 ± 9	$\underline{1240}\pm180$
A24 (pt)	<u>4162</u> ± 86	<u>61.4</u> ± 33.3	1.47 ± 0.80	98.9 ± 2.1	272 ± 8	<u>960</u> ± 145
A25 (pt)	<u>4756</u> ± 112	40.7 ± 0.1	0.86 ± 0.01	98.0 ± 2.5	260 ± 10	$\underline{1070} \pm 129$
A26 (pt)	<u>5563</u> ± 141	$\underline{107.1}\pm0.1$	1.93 ± 0.01	99.2 ± 3.4	287 ± 12	<u>930</u> ± 151
A27 (pt)	10046 ± 275	46.7 ± 8.1	0.47 ± 0.08	99.5 ± 1.1	198 ± 8	$\underline{2150}\pm259$
Mean	4521	46.6	1.18	98.4	189	1081
Min	1314	22.0	0.47	97.5	47	509
P25	2902	24.3	0.77	97.8	129	820
Median	4162	40.7	1.13	98.3	198	960
P75	5160	54.1	1.59	99.1	266	1155
Max	10046	107.1	1.93	99.5	287	2150
IT sc. val.*	<u>150</u>	<u>2</u>				<u>50</u>
Sam>ITs.v.	100%	100%				100%

* Italian Legislative Decree No. 152 (2006). Attachments to the Part IV, Title V, Table 1, Column A.

Underlined values of pseudo-total and hexavalent Cr and HC C>12 exceed Italian screening values set by L.D. 152/2006.

d1, d2 and d3 indicate soil samples collected at 0-20 cm, 30-60 cm and 70-90 cm of depth, respectively; pt indicates soil samples collected from pedological profiles and trenches opened in the field.

Table 4. Values of Pearson's correlation coefficient (r) for pseudo-total ($\mu g g^{-1}$) and hexavalent ($\mu g g^{-1}$; % of the pseudo-total) Cr, organic matter (mg g⁻¹) and heavy hydrocarbons (HC C>12, $\mu g g^{-1}$) contents in 20 soil samples (A1-A20, not pre-treated with n-hexane) from case study A.

Case study A	Pseudo- total Cr (μg g ⁻¹)	Cr(VI) (µg g ⁻¹)	Cr(VI) (% of pseudo- total)	Spike reduction (%)	Organic matter (mg g ⁻¹)
$Cr(VI) \ (\mu g \ g^{-1})$	0.828***				
Cr(VI) (% of pseudo-total)	-0.602**	-0.415			
Spike reduction (%)	0.221	0.160	-0.018		
Organic matter (mg g ⁻¹)	0.385	0.380	-0.224	0.605**	
HC C>12 (µg g ⁻¹)	0.363	0.460*	-0.218	0.780***	0.643**

***, ** and * indicate statistically significant correlations (2-tails) at p < 0.001, p < 0.01 and p < 0.05, respectively

Table 5. Pseudo-total (μ g g ⁻¹) and hexavalent (μ g g ⁻¹ ; % of the pseudo-total) Cr and organic matte (mg g ⁻¹) contents in soil samples from case study B. The reduction (%) of the ⁵⁰ Cr(VI)-enriched spike after microwave-assisted extraction is shown as well.		Pseudo-		Spike	Organic	
	(mg g) contents spike after microw	in soil samples fro vave-assisted extrac	om case study B. The reduction is shown as well.	ction (%) of the	³⁰ Cr(VI)-enrich	ed

Sample ID	Pseudo- total Cr	Cr(VI)	Spike reduction	Organic matter
	$(\mu g g^{-1})$	(µg g ⁻¹)	(% of tot)	(%)	(mg g ⁻¹)
B 1	209 ± 4	0.72 ± 0.02	0.34 ± 0.01	40.1 ± 0.6	73 ± 3
B2	<u>212</u> ± 5	0.70 ± 0.02	0.33 ± 0.01	46.0 ± 0.6	83 ± 5
B3	$\underline{213}\pm 4$	0.75 ± 0.01	0.35 ± 0.01	41.1 ± 0.1	76 ± 4
B4	$\underline{229}\pm 6$	1.61 ± 0.02	0.70 ± 0.02	38.9 ± 0.5	75 ± 4
B5	$\underline{243}\pm 6$	1.59 ± 0.01	0.66 ± 0.01	41.8 ± 0.2	73 ± 2
B6	<u>244</u> ± 3	1.62 ± 0.02	0.67 ± 0.01	38.2 ± 0.2	76 ± 3
B7	$\underline{282}\pm9$	$\underline{2.25} \pm 0.02$	0.80 ± 0.02	19 ± 1	75 ± 2
B8	$\underline{285}\pm7$	2.00 ± 0.02	0.70 ± 0.01	19.1 ± 0.8	70 ± 3
B9	$\underline{285} \pm 6$	$\underline{2.17}\pm0.02$	0.76 ± 0.02	36.7 ± 0.1	75 ± 2
B10	<u>543</u> ± 12	$\underline{4.99}\pm0.01$	0.92 ± 0.03	27.6 ± 0.7	75 ± 3
B11	<u>573</u> ± 14	$\underline{5.79} \pm 0.02$	1.01 ± 0.02	27 ± 1	78 ± 4
B12	<u>594</u> ± 11	5.11 ± 0.02	0.86 ± 0.01	24 ± 3	70 ± 3
Mean	326	2.44	0.68	33	75
Min	209	0.70	0.33	19	70
P25	225	1.38	0.58	26	73
Median	263	1.81	0.70	37	75
P75	350	2.94	0.81	40	76
Max	594	5.79	1.01	46	83
IT sc. val. $*$	<u>150</u>	2			
Sam>ITs.v.	100%	42%			

* Italian Legislative Decree No. 152 (2006). Attachments to the Part IV, Title V, Table 1, Column A.

Underlined values of pseudo-total and hexavalent Cr exceed the Italian screening values set by L.D. 152/2006.

Table 6. Values of Pearson's correlation coefficient (r) for pseudo-total ($\mu g g^{-1}$) and hexavalent ($\mu g g^{-1}$; % of the pseudo-total) Cr and organic matter (mg g⁻¹) contents in soil samples from case study B.

Case study B	Pseudo- total Cr (μg g ⁻¹)	Cr(VI) (μg g ⁻¹)	Cr(VI) (% of pseudo- total)	Spike reduction (%)
Cr(VI) (µg g ⁻¹)	0.984***			
Cr(VI) (% of pseudo-total)	0.776**	0.865***		
Spike reduction (%)	-0.579*	-0.616*	-0.696*	
Organic matter (mg g ⁻¹)	-0.192	-0.179	-0.283	0.507

***, ** and * indicate statistically significant correlations (2-tails) at p < 0.001, p < 0.01 and p < 0.05, respectively

Figure 1. Comparison on hexavalent Cr (% of the pseudo-total content) extracted from 20 soil samples (A1-A20, not pre-treated with n-hexane) collected at case study A from: i) sampling grid 20x20 m at 0-20 cm (d1; 5 samples), 30-60 cm (d2; 4 samples) or 70-90 cm (d3; 5 samples) of depth; ii) pedological profiles and trenches (pt, 6 samples: depth ranging from 30 to 180 cm) opened in the field.

Highlights

Most of samples from two case studies exceed the legal Italian Cr(VI) screening value

The Cr(VI) is significantly correlated to the pseudo-total Cr content in both soils

The Cr(VI) extracted from both soils is less than 2 % of the pseudo-total Cr content

Both soils A and B show a low potential to oxidise Cr(III) to Cr(VI)

Organic compounds caused a partial reduction of Cr(VI)-spike during extraction phase