



ACADEMIC
PRESS

Molecular Phylogenetics and Evolution xxx (2002) xxx–xxx

MOLECULAR
PHYLOGENETICS
AND
EVOLUTION

www.academicpress.com

Phylogeography of chamois (*Rupicapra* spp.) inferred from microsatellites

Trinidad Pérez, Jesús Albornoz, and Ana Domínguez*

Departamento de Biología Funcional, Área de Genética, Universidad de Oviedo, Oviedo 33071, Spain

Received 2 January 2002; received in revised form 10 June 2002

Abstract

Evolutionary relationships among populations of chamois (*Rupicapra* spp.) across their current range from the Caucasus to the Cantabrian Mountains were investigated. The allelic variation in 23 microsatellite loci was assessed in eight geographical populations, recognised as subspecies of the two closely related species *R. pyrenaica* and *R. rupicapra*. Analysis of variance in allele frequencies (*Fst*, statistics) and in repeat numbers (*Rst*, statistics) showed these data to be highly structured. Two genetic distances between pairs of populations, *Ds* and $(\delta\mu)^2$, were computed and phylogenetic trees were constructed. Similar patterns were produced by the different statistics. All trees indicate a deep divergence between the two recognised species, which is compatible with archaeological data that place their split in the Riss–Würm interglacial period. Genetic distances between pairs of populations are highly correlated with geographical distance. This suggests that the history of the genus during Pleistocene glacial-interglacial periods was dominated by expansions and contractions within limited geographic regions, leading to alternate contact and isolation of contiguous populations. In addition, the alpine barrier has played a substantial role in West–East differentiation. © 2002 Elsevier Science (USA). All rights reserved.

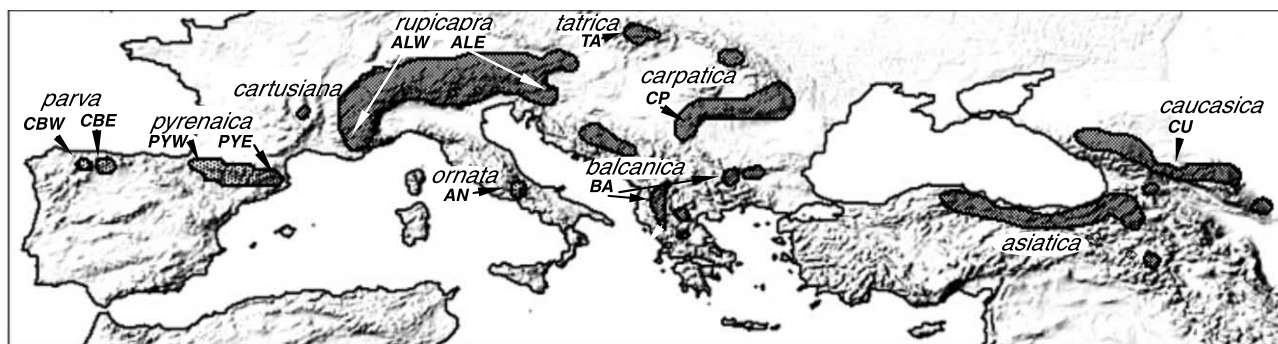
1. Introduction

Study of the genetic differentiation among geographical populations of related taxa allows past historical and evolutionary events, leading to current phylogeographic structure to be inferred (Avice et al., 1987). An increasing number of studies based on DNA polymorphism provide information about the influence of the Pleistocene glaciations on species expansions and contractions (Avice et al., 1998; Hewitt, 1996; Taberlet et al., 1998).

Chamois (genus *Rupicapra*) are mountain ungulates of the subfamily Caprinae, presently distributed over most of the medium to high altitude mountain ranges of Southern Europe, the Balkans, and the Near East. Paleontological evidence shows that the Rupicaprini originated during the Miocene in Asia and that *Rupicapra* spread to Europe during the middle Pleistocene (Masini and Lovari, 1988). There are 10 distinct geographical populations of chamois that have been recognised as subspecies (Couturier, 1938 and Dolan, 1963; cited in Masini and Lovari, 1988). The geographical

distribution of living chamois, as well as population sizes given by Masini and Lovari (1988), presented in Fig. 1. Today, at least three subspecies have been drastically reduced in size: there are about 2000 individuals of *R. r. caucasica* (Jason Badridge, pers. comm.); the populations of *R. r. balcanica* from Greece and Bulgaria have declined severely over the last few years (Haritakis Pappoioannou and Michael Brown, pers. comm.) and the population of *R. r. tatica* has been reduced to about 200 individuals (Wojciech Gasienica Byrcyn, pers. comm.). In recent years, geographical populations have been grouped into two species on the basis of morphological and behavioural characters, *Rupicapra pyrenaica* (with the subspecies *parva*, *pyrenaica*, and *ornata*) from southwestern Europe and *R. rupicapra* (with the subspecies *cartusiana*, *rupicapra*, *tatica*, *carpatica*, *balcanica*, *asiatica*, and *caucasica*) from north-eastern Europe. Analysis of genetic variation of a limited number of populations for allozyme loci (Nascetti et al., 1985) and RFLPs of mitochondrial DNA (Hammer et al., 1995) showed a considerably higher divergence between populations of the two proposed species than between populations within the same species. This was interpreted as support for the two species distinction.

* Corresponding author. Fax: +34-85-10-35-34.
E-mail address: sanjurjo@correo.uniovi.es (A. Domínguez).



<i>Rupicapra pyrenaica</i>		<i>Rupicapra rupicapra</i>	
<i>parva</i>	(6.000)	<i>cartusiana</i>	(100)
<i>pyrenaica</i>	(25.000)	<i>rupicapra</i>	(450.000)
<i>ornata</i>	(350)	<i>tatrica</i>	(900)
		<i>carpatica</i>	(2.500)
		<i>balcanica</i>	(25.000)
		<i>caucasica</i>	(13.000)
		<i>asiatica</i>	(unknown)

Fig. 1. Present distribution and population sizes of the genus *Rupicapra* (based on Masini and Lovari, 1988). Sampled locations are marked with an arrowhead and labelled with the abbreviations used throughout.

64 Microsatellites have been found to be very useful in
65 the study of phylogenetic relationships among popula-
66 tions within species or between closely related species
67 (Bowcock et al., 1994; Forbes et al., 1995; Goldstein et
68 al., 1995; MacHugh et al., 1997; Paetkau et al., 1997;
69 Polzheim et al., 2000; Richard and Thorpe, 2001). In this
70 paper, we use microsatellite polymorphisms to investi-
71 gate the genetic variation in chamois across its geo-
72 graphical range. We screened 25 bovine and caprine
73 microsatellite loci, amplifying a specific product in
74 chamois (Pérez et al., 2000) in 145 individuals com-
75 prising eight different subspecies. The data were used to
76 quantify levels of genetic variability within local popu-
77 lations as well as to investigate genetic relationships
78 among the proposed species and subspecies to gain in-
79 sights into *Rupicapra* phylogeny. The results were re-
80 lated to the fossil record and the influence of Pleistocene
81 glaciations on the population dynamics.

82 2. Materials and methods

83 2.1. Population samples

84 Our objective was to collect at least 20 samples from
85 each location across the geographical range of the genus
86 *Rupicapra*. The protected status of most populations,
87 together with other practical difficulties, meant that we
88 were unable to obtain any samples from the massif of
89 Chartreuse (*R. r. cartusiana*) or from Turkey (*R. r. asi-*
90 *atica*) and that we obtained only a small number of
91 samples from some other populations. It has been
92 shown that increasing the number of loci has a larger
93 and more important effect on the sampling variance

94 than increasing the sample size (Shriver et al., 1995;
95 Takezaki and Nei, 1996). Therefore, the study was based
96 on a large number of loci to counterbalance the small
97 number of samples of some populations. Samples were
98 collected from 1995 to 2001. For large populations,
99 where hunting is allowed, samples were of either muscle
100 or skin preserved in 96% ethanol by gamekeepers or
101 teeth from skulls sent to taxidermists. For protected
102 populations, samples were obtained from accidentally
103 dead or poisoned animals; tissues as well as their con-
104 servation method were diverse (bone, salted skin, muscle
105 in ethanol, and muscle in formalin) and were sent by
106 biologists. A total of 145 samples were collected from 11
107 locations (Fig. 1).

108 2.2. Laboratory analysis

109 Two methods were used to isolate DNA for ampli-
110 fication. DNA from soft tissue was extracted with
111 Chelex, following Estoup et al. (1996). DNA from bone
112 or teeth was extracted from 1 g powdered material fol-
113 lowing Cattaneo et al. (1995) and purified further with
114 Chelex. After DNA precipitation, the pellet was resus-
115 pended in 250 µl sterile water and 50 µl was transferred
116 to a new tube and 450 µl Chelex 10% was added. The
117 mixture was incubated at 60 °C for 2 h. After testing the
118 quality and quantity of the DNA in a minigel, 2–5 µl
119 appropriate dilution (1/10–1/40) was used to perform
120 each PCR reaction.

121 The microsatellite markers used in this study included
122 14 caprine and 11 bovine loci, amplifying specific
123 products in chamois. PCR conditions were as described
124 in Pérez et al. (2000). PCR products were electropho-
125 resed in 6% denaturing polyacrylamide gels and visual-

126 ised by silver staining (Promega). Sequencing reactions
127 of pUC18 (silver sequence DNA sequencing system,
128 Promega) were used as standard markers to assign the
129 allele size. For loci with many alleles, additional gels, in
130 which individuals were ordered according to their pre-
131 viously determined allele size, were run to check further
132 genotypes.

133 2.3. Statistical analysis

134 Multilocus individual genotypes were arranged in a
135 matrix of 25 loci per 145 individuals. For three indi-
136 viduals (1 from Tatra, 1 from the Balkans, and 1 from
137 the Caucasus), multilocus genotypes were incomplete
138 because some loci could not be amplified.

139 Weinberg equilibrium for each locus and for each geo-
140 graphic area, as well as the test of disequilibrium for
141 pairs of loci, was performed using GENEPOP on the
142 web (<http://wbiomed.curtin.edu.au/genepop/>; Raymond
143 and Rousset, 1995). The sequential Bonferroni proce-
144 dure was applied to correct the significance level for
145 multiple comparisons (Sokal and Rohlf, 1995). In each
146 population, every locus was tested for departure from
147 Hardy–Weinberg by the “exact HW test” (Weir, 1996).
148 The algorithm used to estimate the exact *P* value was a
149 Markov-chain method, with the defaults recommended
150 by the authors. Global tests across loci for each popu-
151 lation or across populations for each locus were con-
152 structed using Fisher’s method. Linkage disequilibrium
153 was tested for all possible pairs of loci in each popula-
154 tion and globally for each pair of loci across popula-
155 tions. Observed and expected heterozygosities were also
156 calculated with GENEPOP.

157 Differences in the extent of genetic variation between
158 studied areas were tested by an ANOVA of the number
159 of heterozygous loci per individual, a variable indicating
160 heterozygosity (Weir, 1996). Non-HW loci were ex-
161 cluded; therefore, the values were based on 23 loci.
162 Comparisons among all possible pairs of samples were
163 carried out using Student’s *t* test and the significance
164 level was corrected by the sequential test of Bonferroni.

165 The allele-sharing distance between every pair of in-
166 dividuals (Bowcock et al., 1994) was calculated using the
167 calculator at <http://www.biology.ualberta.ca/jbrzusto/>,
168 and a neighbour-joining tree (Saitou and Nei, 1987) was
169 constructed from the resulting distance matrix.

170 The genetic structure of the populations was analysed
171 by both Wright’s *F*-statistics (Weir, 1996), based on
172 differences in allele frequencies, and by *R*_{st}-statistics
173 (Slatkin, 1995), based on differences in the allele size. *R*_{st}
174 is more appropriate for studying the levels of genetic
175 variation under the stepwise mutation models thought
176 to apply to microsatellites. The proportion of genetic
177 variation, both in allele frequencies or in allele sizes
178 (without standardisation), accounted for by the different
179 phylogenetic levels was analysed by a hierarchical

analysis of variance (Weir, 1996) with the aid of the
FSTAT 2.9.1 (Goudet, 2000). Bootstrapping over loci
was used to obtain confidence intervals for values of *F*_{st}
(15,000 bootstraps).

Recent studies have tested the performance of different
genetic distance measures in resolving the evolutionary
relations of closely related populations or species from
microsatellite data (Paetkau et al., 1997; Takezaki and
Nei, 1996). The results have shown that Nei’s standard
distance, *D*_s (Nei, 1972) and $(\delta\mu)^2$, specifically developed
for microsatellite loci, (Goldstein et al., 1995) performed
well. *D*_s is more appropriate for studying the fine-scale
population differentiation, while $(\delta\mu)^2$ is better for res-
olving the relationships among very distinct populations
and closely related species and for estimating evolution-
ary times. Consequently, we calculated both distances, *D*_s
and $(\delta\mu)^2$. The GENDIST program in PHYLIP 3.5c
(Felsenstein, 1993) was used to obtain *D*_s, while MICRO-
SAT (Minch, 1995) was used to calculate $(\delta\mu)^2$. UPGMA
and Neighbour-Joining trees were produced with the
NEIGHBOR program from PHYLIP 3.5c. Bootstrapping
gene frequencies over loci were achieved with SEQ-
BOOT from PHYLIP 3.5c for *D*_s and with MICROSAT
for $(\delta\mu)^2$. These multiple data sets were used to obtain
consensus trees with the CONSENSE program, in
PHYLIP 3.5c. Three diagrams were obtained with Tree-
ViewPPC 1.6 (Page, 2000). Distances were based on 20
loci; two loci in broad HW disequilibrium (see Section 3)
and three monomorphic loci were excluded.

The relation between genetic and geographical dis-
tances was analysed by regression of $(\delta\mu)^2$ on the dis-
tance, in hundreds of kilometres, between pairs of
studied areas. The significance of the association was
tested by a Mantel test performed with GENEPOP on
the web. This program computes significance by deter-
mining the distribution of the Spearman Rank correla-
tion coefficient under the null hypothesis of
independence and comparing the observed value with
this distribution. We used 100,000 permutations to de-
termine the rejection zone of the correlation.

3. Results

3.1. Within-population data

The number of heterozygotes for loci SR-CRSP07 and
INRA040 had already been shown to be much lower than
expected in the Cantabrian population (Pérez et al., 2000).
Further analysis of these two loci showed that SR-
CRSP07 may be X-linked; no male, out of 33, showed
more than one allele, while 17 out of the 49 known females
were heterozygotes. Locus INRA040 has non-amplifying
alleles, the Cantabrian population showed a deficit of
heterozygotes but amplification products were obtained
for all the individuals analysed. However, it was impos-

180
181
182
183
184
185
186
187
188
189
190
191
192
193
194
195
196
197
198
199
200
201
202
203
204
205
206
207
208
209
210
211
212
213
214
215
216
217
218
219

220

221

222
223
224
225
226
227
228
229
230
231

sible to amplify INRA040 from two individuals from the Pyrenees, from three individuals from the Alps or from any of the 17 individuals from the Carpathians. We conclude that INRA040 has one or more non-amplifying alleles that are highly frequent in the species *R. rupicapra*. These two loci were excluded from posterior analysis. Another three loci, SR-CRSP02, BM1824, and ILSTS008, were monomorphic.

Deviations from Hardy–Weinberg equilibrium were tested for every combination of locus per study area showing polymorphism and with a sample size higher than 5. Significant deviations were observed for one, CP-SR-CRSP13, out of the 146 populations per loci combinations tested and for two loci, SR-CRSP14 and SR-CRSP15, when global tests across populations were performed ($\alpha = 0.05$). In the three cases, there was heterozygote deficiency that could arise due to non-amplifying alleles or to population subdivision. In the case of SR-CRSP14, non-amplifying alleles may be the most plausible explanation because there is a deficit of heterozygotes in populations that are in clear equilibrium for other loci. The other two are probably due to population subdivision. Global tests across loci show a deficit of heterozygotes for populations CP and BA (Table 1). Disequilibrium between pairs of loci was non-significant in every comparison.

In total, 179 alleles were detected across the 23 loci and the 11 study areas. This gives a mean number of alleles per locus of 7.78. The mean number of alleles per locus in each population is 3.22. This mean is biased downwards by the populations with very small sample sizes. The total number of alleles in the two species was 132 for *R. pyrenaica* and 148 for *R. rupicapra*.

Observed heterozygosities were, in general, slightly lower than expected, indicating a general excess of homozygotes, which in the Carpathians and the Balkans was significant ($P < 0.001$). An analysis of variance of individual observed heterozygosities revealed differences

between populations ($F_{10,131} = 9.88$; $P = 3.4 \times 10^{-12}$). Populations from the Alps, the West-Pyrenees, and the East-Cantabrian Mountains are significantly more variable ($\alpha = 0.05$) than populations from the Apennines, the Carpathians, the Caucasus, and the West-Cantabrian Mountains. Differences between the pairs AN–TA, AN–PYE, AN–CBW, PYE–ALW, and ALW–BA were also significant.

3.2. Among-population data

A neighbour-joining tree of 142 individuals (3 individuals with incomplete multilocus genotypes were excluded) based on allele sharing (Fig. 2) shows striking differences between populations pertaining to different species. The individual from the Apennines and one of the individuals from the Balkans do not group so closely with their specific group.

Species-private alleles, 30 for *R. pyrenaica* and 47 for *R. rupicapra*, were found in 17 loci (Fig. 3). Of these, only one locus, SR-CRSP06, can be considered diagnostic, six alleles were found in *R. pyrenaica*, ranging in size between 143 and 153 base pairs, while *R. rupicapra* has only two alleles of sizes 139 and 141 bp. Among the species-private alleles, 33 were also population specific (Table 1). It may be noted that the population of the Apennines, with only one sampled individual, showed two private alleles.

Analysis of variance indicates a significant structure of the data. The percentages of the genetic variance accounted for by differences between species, between study areas, and within study areas were 29.85, 11.78, and 58.37 for frequencies and 44.90, 9.08, and 46.02 for allele sizes, respectively. Every pairwise comparison of genetic differentiation between studied areas was significant ($\alpha = 0.05$, data not shown).

Pairwise genetic distances, D_s and $(\delta\mu)^2$ (Table 2), were highly correlated ($r = 0.92$). For distantly related pairs of populations, $(\delta\mu)^2$ increases more than D_s .

Table 1
Descriptive statistics for each population over all loci

Study area	Abbreviation	<i>n</i>	LP	<i>A</i>	PA	<i>P</i> -HW	He	Ho
Cantabrian-West	CBW	19.00	0.68	3.00	2	0.1856	37.91	36.16
Cantabrian-East	CBE	21.00	0.76	3.72		0.3800	44.92	45.13
Pyrenees-West	PYW	24.00	0.76	4.44	12	0.1069	44.40	44.02
Pyrenees-East	PYE	17.00	0.76	4.16		0.2278	43.01	39.13
Apennines	AN	1.00	0.08	1.08	2	–	–	8.70
Alps-West	ALW	18.00	0.83	4.08	2	0.3373	50.77	47.58
Alps-East	ALE	11.00	0.83	3.71		0.1143	48.15	46.25
Tatra	TA	2.91	0.79	2.29	1	0.7848	46.96	42.75
Carpathians	CP	17.00	0.79	3.04	3	0.0007	37.78	30.69
Balkans	BA	6.78	0.74	3.09	5	0.0005	48.76	35.61
Caucasus	CU	6.43	0.58	2.83	6	0.1733	37.74	32.09

n, mean number of individuals typed per locus (it equals the number of individuals sampled in each population, except for the samples TA, BA, and CU for which some amplifications failed due to bad quality of the DNA sample); LP, proportion of polymorphic loci; *A*, mean number of alleles per locus; PA, number of private alleles; *P*-HW, exact *P* value associated with the Hardy–Weinberg equilibrium; He, expected heterozygosity in %; Ho, observed heterozygosity in %.

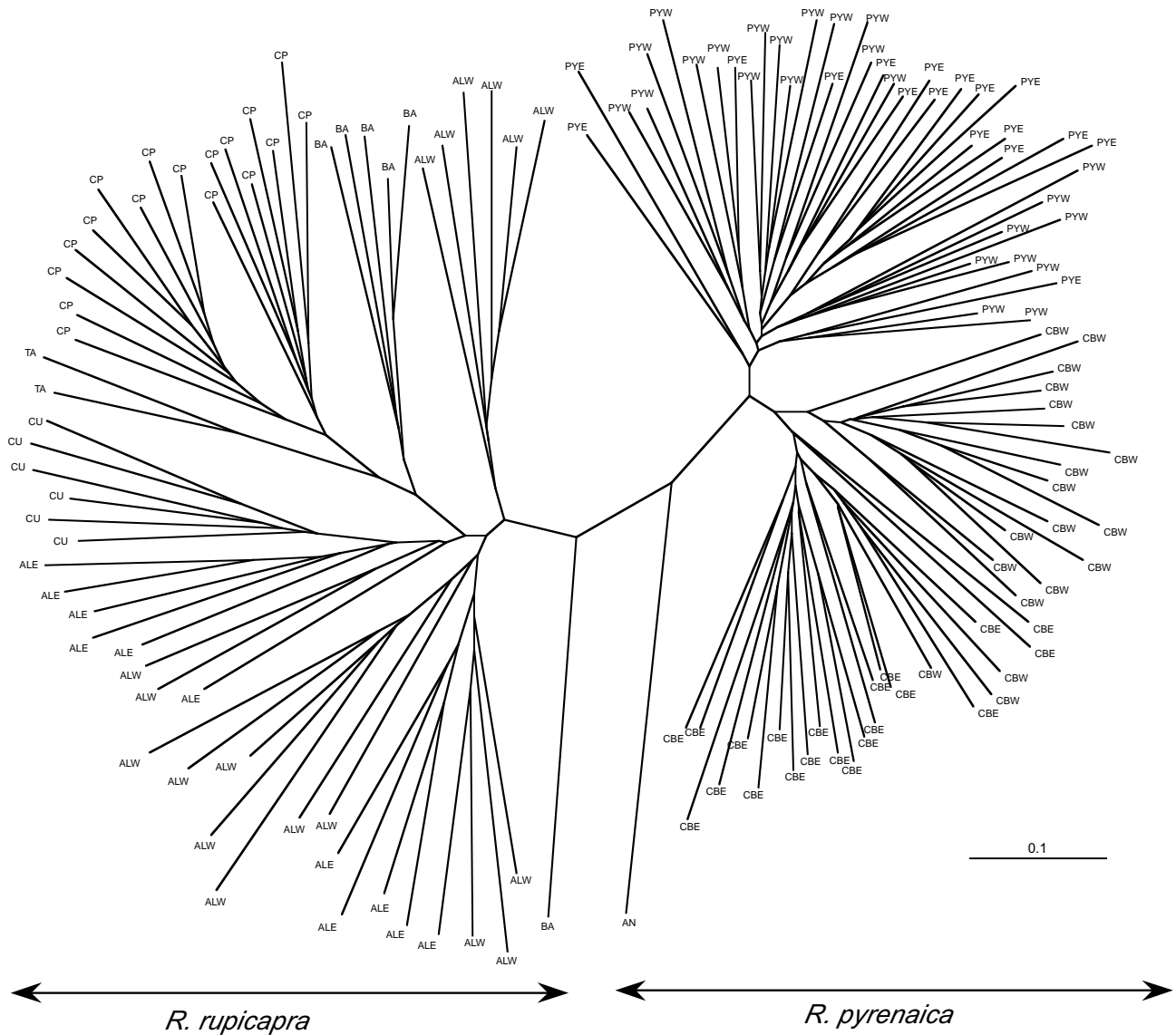


Fig. 2. Neighbour-joining tree of individuals based on allele-sharing distance for multilocus genotypes.

306 Neighbour-joining trees (Fig. 4) were constructed from
 307 matrixes of both genetic distances, D_s and $(\delta\mu)^2$. Pop-
 308 ulations group into two main clusters representing the
 309 two proposed species *R. pyrenaica* and *R. rupicapra*.
 310 There is close agreement between trees based on D_s or
 311 $(\delta\mu)^2$ that only differ in the branches of the Caucasia
 312 population. In the tree based on $(\delta\mu)^2$, the populations
 313 from the Alps, the Carpathians, and the Tatra, group
 314 close together and the Caucasia population is more
 315 distantly related than in the tree based on D_s .

316 The neighbour-joining tree of $(\delta\mu)^2$ distance recapit-
 317 ulates the geographical distribution areas of chamois.
 318 Pairwise $(\delta\mu)^2$ distances were represented against esti-
 319 mated geographical distances between pairs of study
 320 areas (Fig. 5). There is a clear correlation ($r = 0.66$) be-
 321 tween both measures ($P = 0.00034$, one-tailed Mantel
 322 test). It can be observed that the relationships between

genetic and geographical distances (the regression coef- 323
 324 ficients) are equal whether at the level of pairs of pop-
 325 ulations within species ($b = 0.15 \pm 0.06$) or in pairs of
 326 populations between species ($b = 0.17 \pm 0.03$). The
 327 intercepts of the two regression lines are 1.59 ± 0.80 for
 328 pairs of study areas within species and 6.93 ± 0.69 for
 329 pairs between species (t Student = 5.07, $df = 51$,
 330 $P = 2.8 \times 10^{-6}$). Therefore, mean genetic distances ad-
 331 justed by geographical distance are larger for inter-
 332 specific comparisons.

4. Discussion 333

334 For the 23 autosomal standard microsatellite loci as-
 335 sayed (Pérez et al., 2000), 20 were polymorphic in the
 336 genus *Rupicapra*, with a mean of 8.80 alleles per locus.

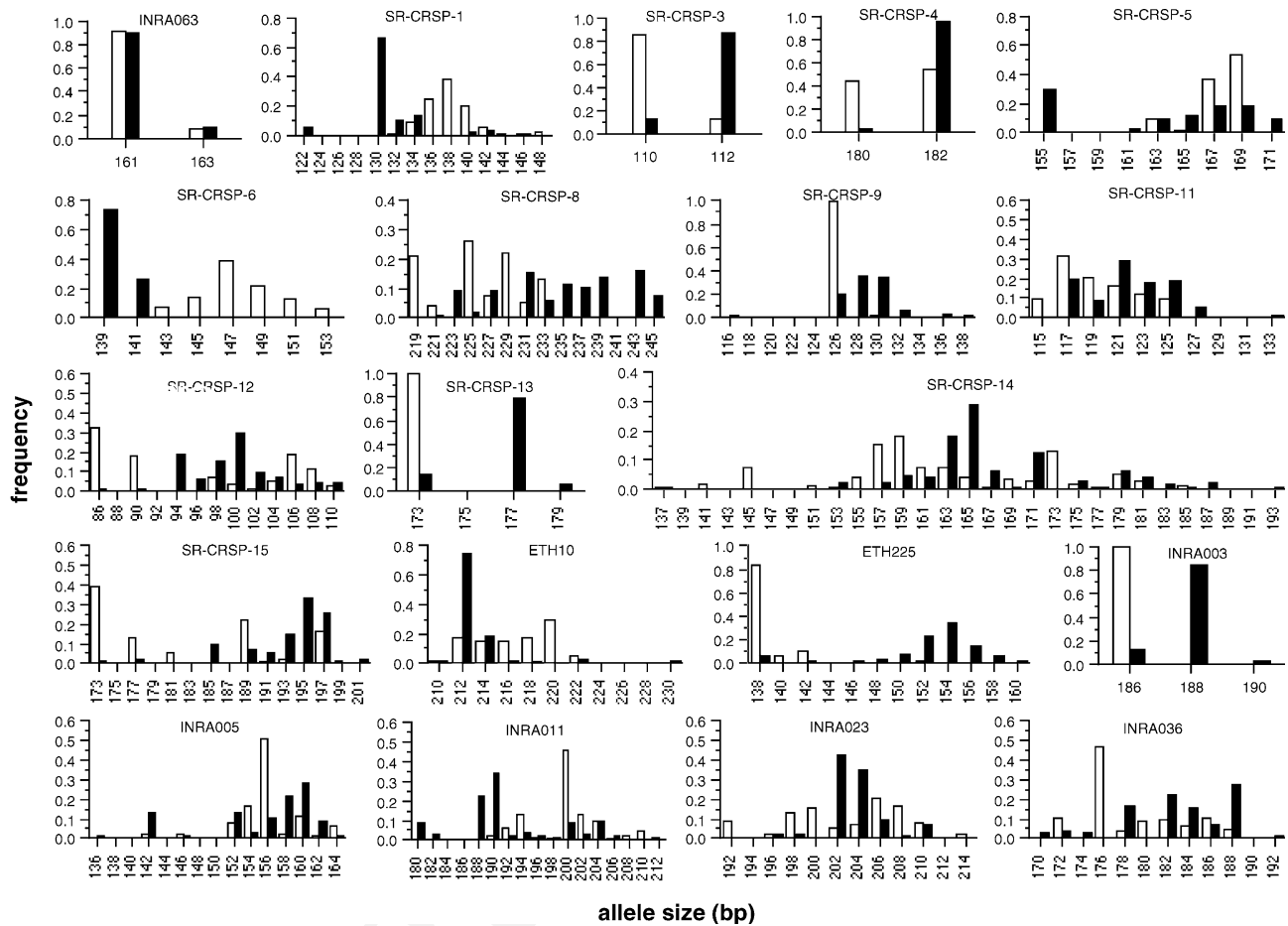


Fig. 3. Allele frequency distributions for polymorphic loci in *R. pyrenaica* (white bars) and *R. rupicapra* (black bars).

Table 2
Pairwise genetic distances between study areas: D_s above diagonal, $(\delta\mu)^2$ below

	CBW	CBE	PYW	PYE	AN	ALW	ALE	TA	CP	BA	CU
CBW		0.16	0.25	0.28	0.82	0.99	1.06	1.18	1.46	1.01	1.25
CBE	0.66		0.26	0.29	0.79	0.89	1.02	1.24	1.58	1.03	1.25
PYW	1.64	2.10		0.08	0.77	0.97	1.03	1.20	1.65	1.13	1.34
PYE	2.78	1.88	1.82		0.90	0.91	1.04	1.24	1.64	1.08	1.34
AN	6.89	5.93	7.32	10.10		0.85	0.89	1.37	1.37	1.24	1.01
ALW	8.30	6.72	8.72	8.50	7.32		0.15	0.39	0.33	0.27	0.22
ALE	9.37	8.19	9.02	9.60	8.48	0.55		0.38	0.37	0.28	0.25
TA	10.10	8.14	11.99	10.72	9.40	2.01	2.14		0.34	0.36	0.48
CP	12.64	10.51	13.76	12.83	9.99	1.56	2.05	1.53		0.37	0.38
BA	8.42	6.36	10.00	9.35	7.49	2.47	2.70	2.04	2.53		0.39
CU	15.64	13.36	14.30	14.01	12.66	3.71	4.43	6.27	2.63	5.63	

Rectangles highlight interspecific comparisons.

337 Chamois populations have three to four alleles per locus
 338 and expected heterozygosities between 38% and 51%.
 339 These estimates of population genetic diversity are low
 340 when compared with values reported for other wild or
 341 domestic artiodactyls (e.g., Bancroft et al., 1995; Fickel
 342 and Reinsch, 2000; Forbes et al., 1995; MacHugh et al.,
 343 1997) and comparable to reported diversities in North
 344 American red deer, wapiti (Polziehn et al., 2000) and in

345 vicuña (Kadwell et al., 2001). The relatively low levels of
 346 diversity in our study may be explained by the bias to high
 347 variability in the choice of microsatellite loci in the origi-
 348 nal species (Pepin et al., 1995). This bias would not affect
 349 homologous species, where microsatellite loci could have
 350 smaller or altered repeats (Forbes et al., 1995). Differences
 351 among heterozygosities seem to reflect population sizes.
 352 The large Alpine populations showed the largest values,

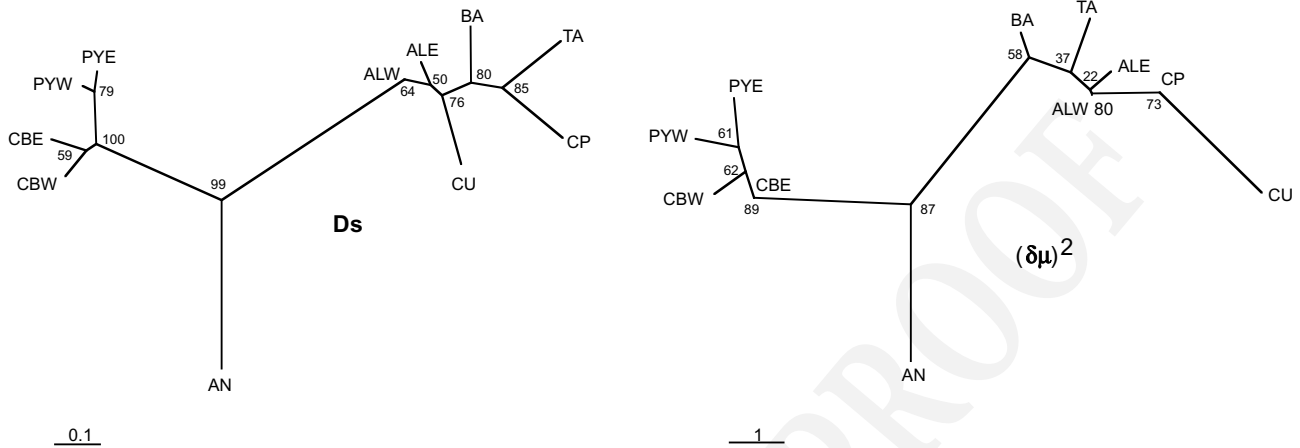


Fig. 4. Neighbour-joining trees summarising phylogeographic relationships between populations based on genetic distances D_s and $(\delta\mu)^2$. Bootstrap values indicating the degree of support of each branch point are shown besides the node as the number of replicates, out of 100, in which the cluster was formed.

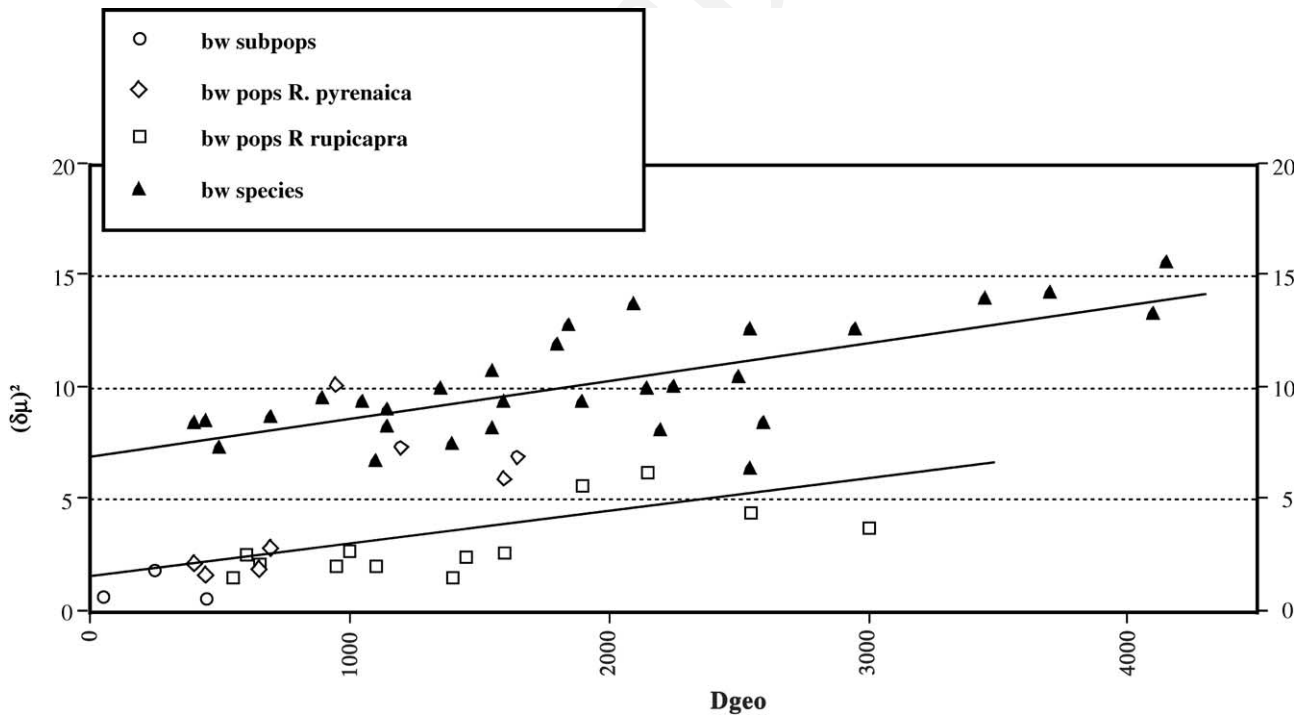


Fig. 5. Regression of $(\delta\mu)^2$ values on geographical distance.

353 while populations of reduced size such as those of the
354 Apennines and the Carpathian Mountains were more
355 homozygous. Worth noting is the low diversity of the
356 Caucasic population, which may be related to the re-
357 cently suffered drastic reduction in population size.

358 Observed heterozygosities were generally lower than
359 expected, reflecting the spatial structuring of popula-
360 tions. Population structuring into subpopulations is re-
361 vealed further by the differentiation between areas
362 within mountain ranges. Moritz (1994) has suggested
363 that regions with significantly different allele frequen-

cies, and hence, with low migration rates are important
from a local management perspective and defined them
as management units (MUs). From this viewpoint, it
may be noted that local subpopulations of chamois
within a mountain range are significantly different from
one another and it is therefore important to prevent
local declines to avoid the loss of genetic variability.

Geographically isolated populations of the different
mountain ranges were ascribed to 10 subspecies. These
10 subspecies were considered either within a single
species by Couturier (1938) and Dolan (1963) (cited in

364
365
366
367
368
369
370
371
372
373
374

Masini and Lovari, 1988) or they have more recently been grouped into two species *R. pyrenaica* and *R. rupicapra* (Lovari and Scala, 1980; Nascetti et al., 1985). Microsatellite analysis of 8 of the 10 proposed subspecies showed a clear differentiation between every pair of populations. Trees based on genetic distances, either individual band-sharing distances or between population distances, clearly separate two groups corresponding to the two proposed species of chamois. The single individual from the Apennines is closer to the Pyrenaica group than to Rupicapra, but assignment is not clear. It can also be noted that this individual is homozygous for two private alleles. Camerano (1914) distinguished the species *R. ornata* besides the two currently accepted ones. Our data offer some support for this classification, but obviously additional samples have to be analysed to clarify the phylogenetic relationships of Apennine chamois. Furthermore, it may be noted that the current genetic constitution of the Apennine chamois may have been largely determined by extreme genetic drift. The subspecies was nearly extinct early in the 20th century and in the late 1940s (Lovari, 1985) and today is composed of a reduced number of individuals.

There was substantial genetic variation between species, 30% for allele frequencies and 45% for allele sizes. Variation between studied areas within species accounted for approximately 10% of the genetic variance, irrespective of whether *Fst* or *Rst* estimates were used. In most cases but not always, genetic distances reflected a larger differentiation among pairs of populations in different mountain ranges, or proposed subspecies than between subpopulations. The lowest Nei's standard genetic distance between proposed subspecies ($D_s = 0.22$ for the pair ALW–CU) was greater than the largest D_s value within mountain ranges. When $(\delta\mu)^2$ is considered, three distances (CBW–PYW, ALW–CP, and TA–CP) are lower than the value of 1.82 observed between the two Pyrenean populations. Genetic distances can be compared with microsatellite-based values reported for other pairs of proposed subspecies. For example, Nei's genetic distance between isolated bear populations ranged between 0.4 and 1.5 (Paetkau et al., 1999), while distances among populations of bighorn sheep ranged between 0.17 and 1.38 (Forbes and Hogg, 1999). D_s distances between subspecies of chamois (0.22–0.90) are at the lower end of these values and are comparable to D_s between proposed subspecies of the North American deer, wapiti ($D_s = 0.18$ –0.69; Polziehn et al., 2000) and between North American populations of grey wolves ($D_s = 0.13$ –0.67; Roy et al., 1994). In contrast with the observation of Forbes and Hogg (1999) in bighorn sheep, $(\delta\mu)^2$ reveals more differentiation. Though not dramatic, those between pairs of proposed subspecies are greater than differences between subpopulations of the same subspecies. In addition, there is the certainty of actual discontinuities between populations in different mountain ranges. Tak-

ing these factors altogether, it might be proposed that, though not very differentiated, populations in different mountain ranges have undergone significant independent evolution sensu Moritz (1994), and hence, they could be considered evolutionarily significant units (ESUs) or subspecies. However, this matter cannot be solved by microsatellite analysis alone and must be the object of mtDNA analysis.

The level of divergence between populations can be compared with archaeological data. The Rupicaprinae originated in Asia during the late Miocene. Masini and Lovari (1988) proposed that the chamois, or its direct ancestor, may have reached Europe as a late immigrant during the early or middle Pleistocene and moved westward, along the mountain chains of the Alpine System. They related its arrival to a cold climatic phase, marked by the arrival in Europe of waves of taxa from cold or open environment. The *Rupicapra* genus is thought to have evolved during the middle and late Pleistocene in West-Eurasia. In the middle Pleistocene, chamois occurred in the same geographic area that living species currently occupy.

Genetic distances are correlated with distances between mountain chains, which is fully compatible with the Asiatic origin of *Rupicapra* and the European colonisation westward along the mountain ranges. The close association of genetic and geographical distance implies that populations differentiated "in situ" and no major migrations occurred after the initial colonisation. Within this general scenario, it was shown that, for equally distant areas, mean genetic distances between the pairs *R. pyrenaica*–*R. rupicapra* are greater than within species, implying that there was an additional barrier to gene flow between the two taxonomic groups. This observation is in agreement with the proposed split of the *Rupicapra* genus into *R. pyrenaica* and *R. rupicapra*. Divergence times (Fig. 6) were calculated from $E(\delta\mu)^2 = 2\beta\tau$, where β is the mutation rate and τ is the time of generation (Goldstein et al., 1995). We lack an estimate for the mutation rate of the microsatellite loci studied and have used the average mutation rate of 5.6×10^{-4} , calculated for 15 microsatellite loci in humans (Weber and Wong, 1993). The generation time in chamois was estimated as 6.24 years/generation (Gaillard, 1992). Separation times were calculated from the UPGMA tree, based on the $(\delta\mu)^2$ distance. It should be noted that these estimates have a considerable error due to the error of the distance itself, on the one hand, and to the added error in the mutation rate estimate, on the other. This error may be large because microsatellite mutation rates differ between loci and between species (i.e., Di Rienzo et al., 1998; Vázquez et al., 2000). The estimated divergence time between the two putative species, *R. rupicapra* and *R. pyrenaica*, is 57,000 years. Our estimate is lower than the 280,000 years of separation estimated from mean nucleotide divergence among mitochondrial RFLP haplotypes of three sub-

431
432
433
434
435
436
437
438
439
440
441
442
443
444
445
446
447
448
449
450
451
452
453
454
455
456
457
458
459
460
461
462
463
464
465
466
467
468
469
470
471
472
473
474
475
476
477
478
479
480
481
482
483
484
485
486

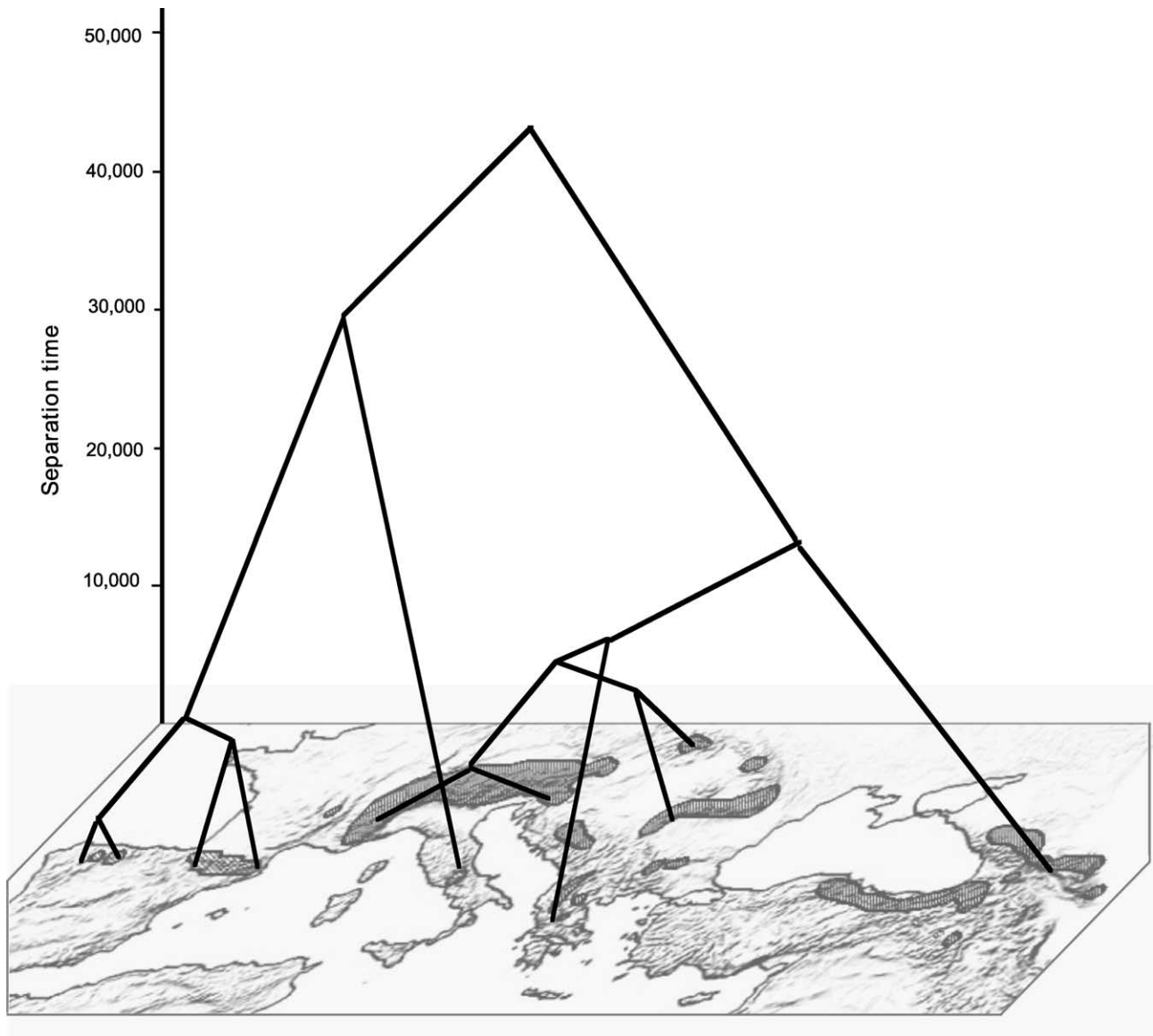


Fig. 6. Phylogeography of chamois.

487 species (Hammer et al., 1995). On the basis of the fossil
488 record, Masini and Lovari (1988) placed the split during
489 the interglacial period Riss–Würm, somewhere in the
490 middle of the two genetic distance-based estimates.

491 Genetic distances between pairs of populations within
492 species mostly depend on the geographical distance of the
493 pairs being considered. Masini and Lovari (1988) pro-
494 posed that *R. rupicapra* evolved in Eastern Europe or Asia
495 Minor during a time of geographic isolation and then
496 spread again to Western Europe during Würm II. In the
497 said case, no association between genetic and geographi-
498 cal distance needs to occur in the interspecific compari-
499 sons. Our results suggest instead that the history of the
500 genus during Pleistocene glacial-interglacial periods was
501 dominated by expansions and contractions within limited
502 geographic regions, leading to alternate contact and iso-
503 lation of contiguous populations. Finally, the warm cli-

mate of the Holocene had rendered the populations 504
definitively isolated occupying the top of the different 505
mountain ranges. 506

The phylogeography of chamois may be compared 507
with other taxa in Eurasia for which a small degree of 508
congruence was found (Hewitt, 1996; Taberlet et al., 509
1998). In general, the northern regions were colonised 510
from Iberian and Balkan refugees and the alpine barrier 511
often isolated Italian lineages. Contrary to the general 512
trend, *Rupicapra* differentiated without major migra- 513
tions, presumably because it is a cold-tolerant species. As 514
in other taxa, the alpine barrier has played a substantial 515
role in West–East differentiation that led to the two spe- 516
cies of the genus. Some studies have reported the evolu- 517
tion in isolation during Pleistocene periods of climatic 518
fluctuations (Hundertmark et al., 2002; Leonard et al., 519
2000; Paulo et al., 2001). Our results also point to isolation 520

521 by distance as the primary agent for differentiation in
522 chamois.

523 Acknowledgments

524 This work was partially funded by Grant PB-REC98-
525 01 from the FICYT. We are indebted to people who, in
526 one way or another, contributed to the collection of
527 chamois samples: the “Consejería de Agricultura” and
528 the “Guardería de Caza” of Asturias, Dr. Carlos Nores,
529 Dr. L. Rossi, Juan Carlos del Campo, J. Bejar, Paloma
530 Barracina, H. Papaioannou, M. Brown, Dr. W. Gasie-
531 nica-Byrcyn, Dr. T. Skalski, J. Meana, S. Erceg, Dr. P.
532 Veinberg, and Dr. J. Badridge. We thank M.W. Bruford
533 for his critical reading of the manuscript.

534 References

535 Avise, J.C., Arnold, R.M., Ball Jr., R.M., Bermingham, E., Lamb, T.,
536 Neigel, J.E., Reeb, C.A., Saunders, N.C., 1987. Intraspecific
537 phylogeography: the mitochondrial DNA bridge between popula-
538 tion genetics and systematics. *Annu. Rev. Ecol. Syst.* 18, 489–522.
539 Avise, J.C., Walker, D., Johns, G.C., 1998. Speciation durations and
540 Pleistocene effects on vertebrate phylogeography. *Proc. R. Soc.*
541 *Lond. B* 265, 1707–1712.
542 Bancroft, D.R., Pemberton, J.M., King, P., 1995. Extensive protein
543 and microsatellite variability in an isolated, cyclic ungulate
544 population. *Heredity* 74, 326–336.
545 Bowcock, A.M., Ruiz-Linares, A., Tomfohrde, J., Minch, E., Kidd,
546 J.R., Cavalli-Sforza, L.L., 1994. High resolution of human
547 evolutionary trees with polymorphic microsatellites. *Nature* 368,
548 455–457.
549 Camerano, L., 1914. Ricerche intorno ai camosci (Parte I a). *Memorie*
550 *Della Regia Accademia Di Scienze Di Torino (Classe Scienze*
551 *Fisiche Matematiche Naturali)* 64, 1–82.
552 Cattaneo, C., Smillie, D.M., Gelsthorpe, K., et al., 1995. A simple
553 method for extracting DNA from old skeletal material. *Forensic*
554 *Sci. Int.* 74, 167–174.
555 Di Rienzo, A., Donnelly, P., Toomajian, C., Sisk, B., Hill, A., Petzl-
556 Erler, M.L., Haines, G.K., Barch, D.H., 1998. Heterogeneity of
557 microsatellite mutations within and between loci, implications for
558 human demographic histories. *Genetics* 148, 1269–1284.
559 Estoup, A., Largiadèr, C.L., Perrot, E., Chourrout, D., 1996. Rapid
560 one-tube DNA extraction for reliable PCR detection of fish
561 polymorphic markers and transgenes. *Mol. Mar. Biol. Biotechnol.*
562 5, 295–298.
563 Felsenstein, J., 1993. PHYLIP-phylogenetic inference package. Version
564 3.5c University of Washington, Seattle. Available from [http://](http://evolution.genetics.washington.edu/phylip/getme.html)
565 evolution.genetics.washington.edu/phylip/getme.html.
566 Fickel, J., Reinsch, A., 2000. Microsatellite markers for the European
567 Roe deer (*Capreolus capreolus*). *Mol. Ecol.* 9, 994–995.
568 Forbes, S.F., Hogg, J.T., 1999. Assessing population structure at high
569 levels of differentiation: microsatellite comparisons of bighorn
570 sheep and large carnivores. *Anim. Conserv.* 2, 223–233.
571 Forbes, S.H., Hogg, J.T., Buchanan, F.C., Crawford, A.M., Allendorf,
572 F.W., 1995. Microsatellite evolution in congeneric mammals:
573 domestic and bighorn sheep. *Mol. Biol. Evol.* 12, 1106–1113.
574 Gaillard, J.M., 1992. Some demographic characteristics in ungulate
575 populations and their implications for management and conserva-
576 tion. *Ongulés/Ungulates* 91, 493–495.

Goldstein, D.B., Ruiz-Linares, A., Cavalli-Sforza, L.L., Feldman, 577
M.W., 1995. Genetic absolute dating based on microsatellites and 578
the origin of modern humans. *Proc. Natl. Acad. Sci. USA* 92, 579
6723–6727. 580
Goudet, J., 2000. FSTAT, a program to estimate and test gene 581
diversities and fixation indices (version 2.9.1). Available from 582
<http://www.unil.ch/izea/software/fstat.html>. Updated from Gou- 583
det (1995). 584
Hammer, S., Nadlinger, K., Hartl, G.B., 1995. Mitochondrial DNA 585
differentiation in chamois (genus *Rupicapra*): implications for 586
taxonomy, conservation, and management. *Acta Theriol. (Suppl.* 587
3), 145–155. 588
Hewitt, G.M., 1996. Some genetic consequences of ice ages and their 589
role in divergence and speciation. *Biol. J. Linn. Soc.* 58, 247–276. 590
Hundertmark, K.J., Shields, G.F., Udina, I.G., Bowyer, R.T., Danil- 591
kin, A.A., Schwartz, C.C., 2002. Mitochondrial phylogeography of 592
moose (*Alces alces*): late pleistocene divergence and population 593
expansion. *Mol. Phylogenet. Evol.* 22, 375–387. 594
Kadwell, M., Fernandez, M., Stanley, H.F., Baldi, R., Wheeler, J.C., 595
Rosadio, R., Bruford, M.W., 2001. Genetic analysis reveals the 596
wild ancestors of the llama and the alpaca. *Proc. R. Soc. Lond. B.* 597
Biol. Sci. 268, 2575–2584. 598
Leonard, J.A., Wayne, R.K., Cooper, A., 2000. Population genetics of 599
Ice Age brown bears. *Proc. Natl. Acad. Sci. USA* 97, 1651–1654. 600
Lovari, S., 1985. Behavioural repertoire of the Abruzzo chamois, 601
Rupicapra pyrenaica ornata Neumann, 1899 (Artiodactyla: Bovi- 602
dae). *Säugetierkundliche Mitteilungen* 32, 113–136. 603
Lovari, S., Scala, C., 1980. Revision of *Rupicapra* genus. I. A statistical 604
re-evaluation of Couturier’s data on the morphometry of six 605
chamois subspecies. *Boll. Zool.* 47, 113–124. 606
MacHugh, D.E., Shriver, M.D., Loftus, R.T., Cunningham, P., 607
Bradley, D.G., 1997. Microsatellite DNA variation and the 608
evolution, domestication and phylogeography of taurine and zebu 609
cattle (*Bos taurus* and *Bos indicus*). *Genetics* 146, 1071–1086. 610
Masini, F., Lovari, S., 1988. Systematics, phylogenetic relationships, 611
and dispersal of the chamois (*Rupicapra* spp.). *Quaternary Res.* 30,
612 339–349. 613
Minch, E., 1995. MICROSAT 1.4d, a program for calculating 614
distances from microsatellite data. Available from [http://hpgl.stan-](http://hpgl.stanford.edu/projects/microsat/)
615 [ford.edu/projects/microsat/](http://hpgl.stanford.edu/projects/microsat/). 616
Moritz, C., 1994. Defining “evolutionary significant units” for 617
conservation. *Trends Ecol. Evol.* 9, 373–375. 618
Nascetti, G., Lovari, S., Lanfranchi, P., Berducou, C., Mattiucci, S., 619
Rossi, L., et al., 1985. Revision of *Rupicapra* genus. III. Elec- 620
trophoretic studies demonstrating species distinction of chamois 621
populations of the Alps from those of the Apennines and Pyrenees. 622
In: Lovari, S. (Ed.), *The Biology and Management of Mountain* 623
Ungulates. Croom Helm, London, pp. 56–62. 624
Nei, M., 1972. Genetic distance between populations. *Am. Nat.* 106,
625 283–292. 626
Paetkau, D., Waits, L.P., Clarkson, P.L., Craighead, L., Strobeck, C., 627
1997. An empirical evaluation of genetic distance statistics using 628
microsatellite data from bear (Ursidae) populations. *Genetics* 147,
629 1943–1957. 630
Paetkau, D., Amstrup, S.C., Born, E.W., Calvert, W., Derocher, A.E., 631
Garner, G.W., et al., 1999. Genetic structure of the world’s polar 632
bear populations. *Mol. Ecol.* 8, 1571–1584. 633
Page, R.D.M., 2000. TreeViewPPC. Version 1.6.2 University of 634
Glasgow, Scotland. Available from [http://taxonomy.zoolo-](http://taxonomy.zoology.gla.ac.uk/rod/treeview.html)
635 [gy.gla.ac.uk/rod/treeview.html](http://taxonomy.zoology.gla.ac.uk/rod/treeview.html). Updated from Page (1996). 636
Paulo, O.S., Dias, C., Bruford, M.W., Jordam, W.C., Nichols, R.A., 637
2001. The persistence of liocene populations through the Pleisto- 638
cene climatic cycles: evidence from the phylogeography of an 639
Iberian lizard. *Proc. R. Soc. Lond. B. Biol. Sci.* 268, 1625–
640 1630. 641
Pepin, L., Amigues, Y.L.A., Berthier, J.-L., Bensaid, A., Vaiman, D., 642
1995. Sequence conservation of microsatellites between *Bos taurus* 643

- 644 (cattle), *Capra hircus* (goat) and related species. Examples of use in
645 parentage testing and phylogeny analysis. *Heredity* 74, 53–61.
- 646 Pérez, T., Albornoz, J., Domínguez, A., 2000. A panel of bovine and
647 caprine microsatellites suitable as markers in chamois. *Anim.*
648 *Genet.* 31, 344–345.
- 649 Polziehn, R.O., Hamr, J., Mallory, F.F., Strobeck, C., 2000. Micro-
650 satellite analysis of North American wapiti (*Cervus elaphus*)
651 populations. *Mol. Ecol.* 9, 1561–1576.
- 652 Raymond, M., Rousset, F., 1995. GENEPOP: population genetics
653 software for exact tests and ecumenicism. *J. Hered.* 86, 248–249.
- 654 Richard, M., Thorpe, R., 2001. Can microsatellites be used to infer
655 phylogenies? Evidence from population affinities of the Western
656 Canary Island lizard (*Gallotia galloti*). *Mol. Phylogenet. Evol.* 20,
657 351–360.
- 658 Roy, M.S., Geffen, E., Smith, D., Ostrander, E.A., Wayne, R.K., 1994.
659 Patterns of differentiation and hybridation in North American
660 wolflike canids, revealed by analysis of microsatellite loci. *Mol.*
661 *Biol. Evol.* 11, 553–570.
- 662 Saitou, N., Nei, M., 1987. The neighbour-joining method: a new
663 method for constructing phylogenetic trees. *Mol. Biol. Evol.* 4,
664 406–425.
- Shriver, M.D., Jin, L., Boerwinkle, E., Deka, R., Ferrel, R.E., 665
Chakraborty, R., 1995. A novel measure of genetic distance for 666
highly polymorphic tandem repeat loci. *Mol. Biol. Evol.* 12, 914– 667
920. 668
- Slatkin, M., 1995. A measure of population subdivision based on 669
microsatellite allele frequencies. *Genetics* 139, 457–462. 670
- Sokal, R.R., Rohlf, F.J., 1995. *Biometry: the principles and practice of* 671
statistics in biological research, third ed. Freeman and Company, 672
New York. 673
- Taberlet, P., Fumagalli, L., Wust-Saucy, A.G., Cosson, J.F., 1998. 674
Comparative phylogeography and postglacial colonization routes 675
in Europe. *Mol. Ecol.* 7, 453–464. 676
- Takezaki, N., Nei, M., 1996. Genetic distances and reconstruction of 677
phylogenetic trees from microsatellite DNA. *Genetics* 144, 389–399. 678
- Vázquez, J.F., Pérez, T., Albornoz, J., Domínguez, A., 2000. Estima- 679
tion of microsatellite mutation rates in *Drosophila melanogaster*. 680
Genet. Res. 76, 323–326. 681
- Weber, J.L., Wong, C., 1993. Mutation of human short tandem 682
repeats. *Hum. Mol. Genet.* 2, 1123–1128. 683
- Weir, B.S., 1996. *Genetic Data Analysis II*, second ed. Sinauer 684
Associates Inc., Sunderland, USA. 685