

Review

Open Access

## Experimental obstructive cholestasis: the wound-like inflammatory liver response

María-Angeles Aller<sup>1</sup>, Jorge-Luis Arias<sup>2</sup>, Jose García-Domínguez<sup>3</sup>, Jose-Ignacio Arias<sup>4</sup>, Manuel Durán<sup>5</sup> and Jaime Arias\*<sup>1</sup>

Address: <sup>1</sup>Department of Surgery I, School of Medicine, Complutense University of Madrid, Spain, <sup>2</sup>Neurosciences Laboratory, School of Psychology, University of Oviedo, Asturias, Spain, <sup>3</sup>Plastic and Reconstructive Surgery Unit, Getafe Hospital, Madrid, Spain, <sup>4</sup>General Surgery Unit, Monte Naranco Hospital, Oviedo, Consejería de Salud y Servicios Sanitarios, Principado de Asturias, Spain and <sup>5</sup>Health Sciences School, King Juan Carlos University, Madrid, Spain

Email: María-Angeles Aller - maaller@med.ucm.es; Jorge-Luis Arias - jarias@uniovi.es; Jose García-Domínguez - eixil10@telefonica.net; Jose-Ignacio Arias - joseignacio.arias@sespa.princast.es; Manuel Durán - mduran.hflr@salud.madrid.org; Jaime Arias\* - jarias@med.ucm.es

\* Corresponding author

Published: 3 November 2008

Received: 26 May 2008

*Fibrogenesis & Tissue Repair* 2008, **1**:6 doi:10.1186/1755-1536-1-6

Accepted: 3 November 2008

This article is available from: <http://www.fibrogenesis.com/content/1/1/6>

© 2008 Aller et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

Obstructive cholestasis causes hepatic cirrhosis and portal hypertension. The pathophysiological mechanisms involved in the development of liver disease are multiple and linked. We propose grouping these mechanisms according to the three phenotypes mainly expressed in the interstitial space in order to integrate them.

Experimental extrahepatic cholestasis is the model most frequently used to study obstructive cholestasis. The early liver interstitial alterations described in these experimental models would produce an ischemia/reperfusion phenotype with oxidative and nitrosative stress. Then, the hyperexpression of a leukocytic phenotype, in which Kupffer cells and neutrophils participate, would induce enzymatic stress. And finally, an angiogenic phenotype, responsible for peribiliary plexus development with sinusoidal arterIALIZATION, occurs. In addition, an intense cholangiocyte proliferation, which acquires neuroendocrine abilities, stands out. This histopathological finding is also associated with fibrosis.

It is proposed that the sequence of these inflammatory phenotypes, perhaps with a trophic meaning, ultimately produces a benign tumoral biliary process – although it poses severe hepatocytic insufficiency. Moreover, the persistence of this benign tumor disease would induce a higher degree of dedifferentiation and autonomy and, therefore, its malign degeneration.

### Background

Obstructive cholestasis is characterized clinically by jaundice, discolored urine, pale stools and pruritus [1-3]. Obstruction of the biliary tree, either intrahepatic or extrahepatic, induces a characteristic pattern of early and late liver morphologic features that can be attributed to the evolution of an inflammatory response [3].

Hepatic inflammation is an important feature of cholestasis liver disease in both humans [1] and experimental animals [4,5]. Inflammatory features of obstructive cholestasis include portal tract edema [3], neutrophil infiltration in the portal tracts [3,5], proliferation of the biliary epithelial cells [1,3] and portal tract fibrosis [1,3,6].

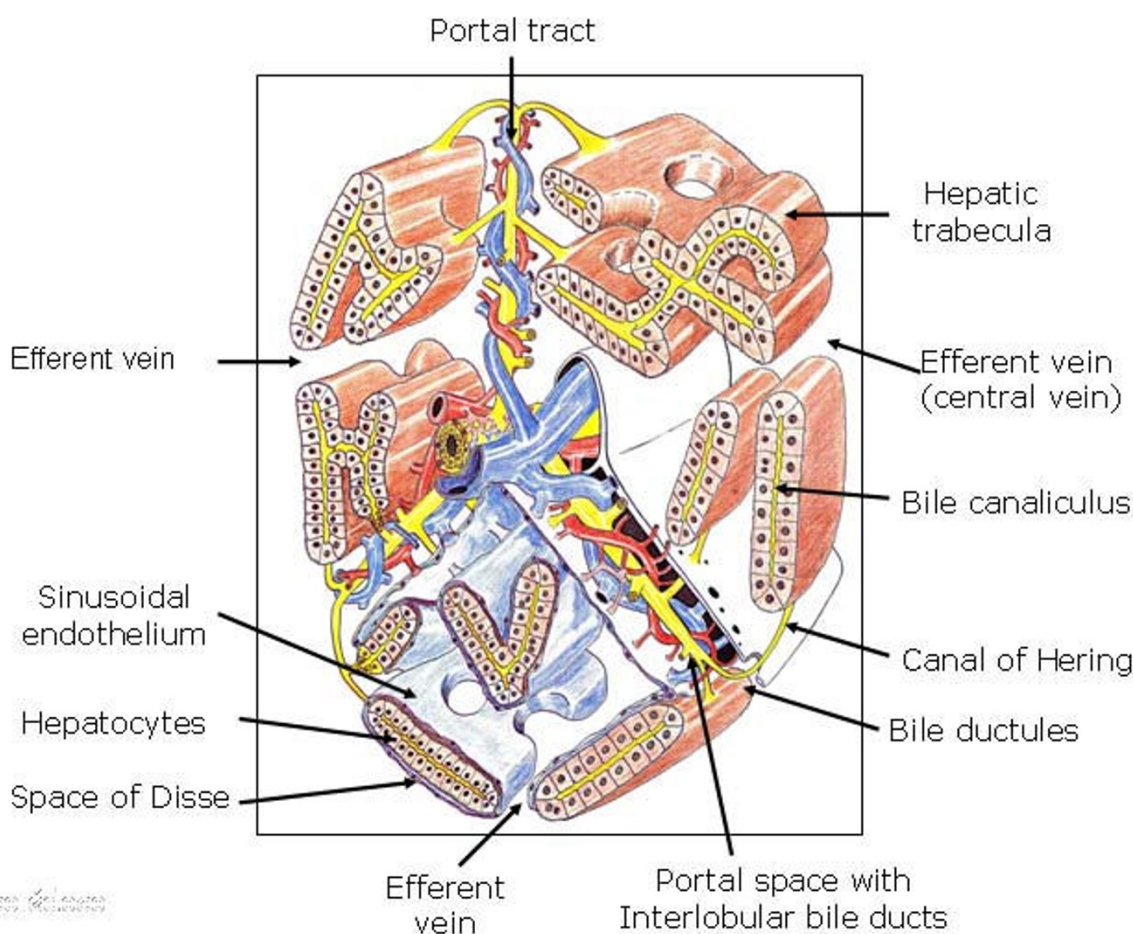
The preferable localization of the inflammatory response in the portal tract reflects the importance that this space occupies in the development of the hepatic cholestasis pathology. In this way, the main role of the portal tract in hepatic inflammation is similar to that of the interstitial space during inflammation of other organs or tissues [7,8]. This is why both spaces, the portal tract and the interstitium, can be considered similar to a certain extent.

**The functional biliary tree in the interstitial space**

The biliary tree begins with the 'source of the bile', which is the bile canaliculus, made up by the canalicular domain

of adjacent hepatocytes [3]. Bile canaliculi form a network of channels between hepatocytes and drain into the canals of Hering (intrahepatic bile ducts) that are lined by hepatocytes and cholangiocytes and also contain hepatic stem cells [3,9-11].

The canals of Hering continue into the bile ductules, which in turn drain into interlobular bile ducts, located in the portal space (Figure 1). Bile ductules and interlobular bile ducts are composed entirely of cholangiocytes. Interlobular bile ducts then continue into progressively larger ducts and finally drain into the extrahepatic biliary tract. The extrahepatic biliary tract is grossly divided into the



**Figure 1**  
**Schematic 3D representation of a complex acinus according to Rappaport.** Three portal tracts diverge from one axis, made up of the bile duct, the hepatic arterial and portal venous branches. At the same time, several trabeculae or laminae hepatis that are two cells thick arise from these three portal spaces and are oriented towards the efferent veins (central veins). In the lower part of the drawing, the liver plates are covered by the sinusoidal endothelium and the space of Disse located between both structures contains tissue fluid, which flows outwards into the lymphatics of the portal zones. The space of Disse continues with the portal space, and they both make up the interstitial space of the acinus.

common hepatic duct, the common bile duct, the cystic duct and the gall bladder [3,9,11]. The intrahepatic and extrahepatic biliary tract are supplied and nourished by a network of fine vessels called the peribiliary vascular (or capillary) plexus, derived from the hepatic artery [3,9]. The liver has a connective envelope, that inflexes at the level of the hilum and follows ducts and vessels within the organ until the periportal spaces. The periportal spaces represent the center of the functional units of the organ. From the portal tract, the blood supplies feed into vascular sinusoids that are bounded by laminae of hepatocytes that finally drain into the efferent central vein [12].

Between the sinusoidal endothelium and the vascular pole of the hepatocytes lies the space of Disse (perisinusoidal space) [12] (Figure 1). This space contains the extracellular matrix (ECM) and hepatic stellate cells (HSCs) (also referred to as Ito cells, lipocyte, perisinusoidal or fat-storing cells), and constitutes the framework of the acinus [6,12,13].

The composition of the ECM within the liver is not homogeneous. The connective tissue of the space of Disse is different from the connective tissue of the rest of the liver, such as the external capsule, septa, periductal and perivascular areas, and portal tracts [12]. Thus, two types of ECM are present in the normal liver acinus, namely the ECM sinusoidal tract in the space of Disse with HSCs, and the ECM in the portal and central vein tracts, where myofibroblasts are present [12,14,15].

Therefore, both the space of Disse and portal-central vein spaces, are considered to be interstitial spaces of the liver acinus, with the functional ability to synthesize and to degrade the ECM [12,15]. Thus, the ECM is not only a scaffold, having a mechanical role in supporting and maintaining tissue structures, but is also a complex and dynamic meshwork influencing many biological cell functions. The ECM has profound influences on the structure, viability and function of cells. However, it has also been recognized that the effect of the ECM on cells extends to immune and inflammatory cells [16]. Since the biliary microcirculation moves immersed in the ECM of the liver interstitial spaces, Disse and portal spaces, it is not bold to propose that in obstructive cholestasis the liver ECM plays a key etiopathogenic role.

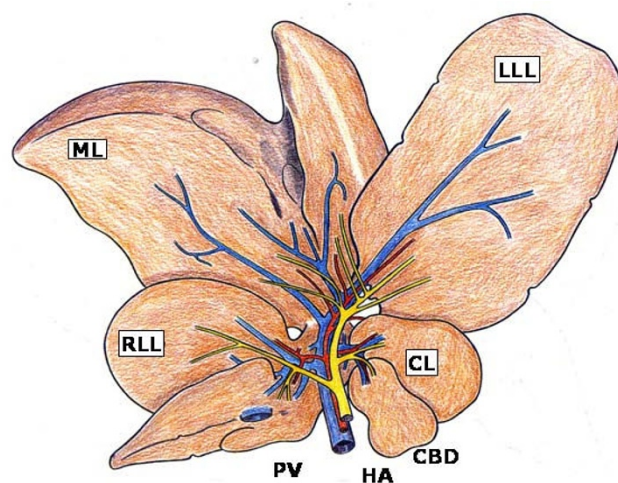
### Surgical experimental cholestasis

Obstructive jaundice causes a high rate of morbidity and mortality in the human clinical field [17]. The serious repercussions of cholestasis on the liver and on the systemic level [1-3,17] have led to the creation of many experimental models so as to better understand its pathogenesis, prophylaxis, and treatment.

Several surgical techniques for developing extrahepatic cholestasis have been described, especially in the rat, based on the section of the bile duct between ligatures [18,19]. These techniques are models of reversible obstructive jaundice, since they imply a high incidence of recanalization of the extrahepatic biliary route, which can be avoided by placing the duodenum and the distal part of the stomach between the two ligated and sectioned ends of the bile duct [19]. These macrosurgical techniques of extrahepatic cholestasis, called bile duct ligation (BDL), caused development of infected hilar biliary pseudocysts by dilation of the proximal end of the bile duct. As a result, the animals died during the first 2 weeks of the postoperative period due to sepsis caused by multiple abscesses in the intraperitoneal, hepatic and pulmonary areas [20-22].

The hepatic parenchyma in the rat has four lobes: the right lateral, middle, left lateral and caudate lobes, which in turn have independent portal and arterial vascularization and a separate biliar drainage [23] (Figure 2).

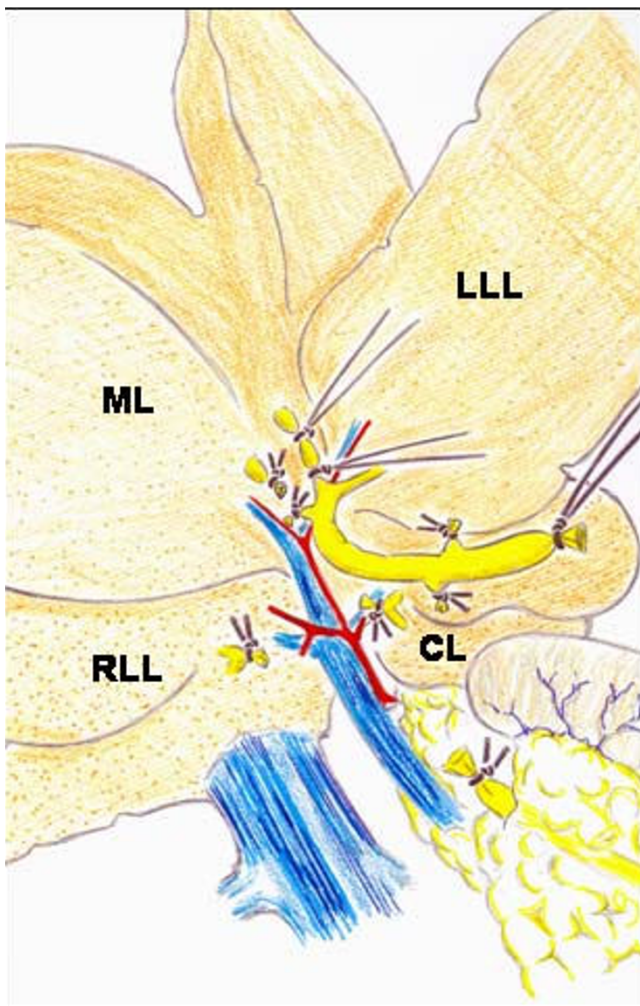
This anatomic feature makes it possible to resect the bile ducts that drain the four lobes of the liver in continuity with the common bile duct up to the beginning of its intrapancreatic segment by means of a microsurgical technique [22,24]. First, the common bile duct is ligated and sectioned close to its intrapancreatic portion. This maneuver, which produces dilation of the extrahepatic biliary tract, facilitates the posterior dissection of the common



**Figure 2**  
Representation of the rat's liver made up of four lobes: median (ML), left lateral (LLL), right lateral (RLL) and caudate (CL). In the hilum, the relation between the portal, arterial and biliary branches is appreciated, as well as the inexistence of the gall bladder. CBD, common bile duct; HA, hepatic artery; PV, portal vein.

bile duct and the lobular biliary branches of the four hepatic lobes.

Once the common bile duct is sectioned, it is shifted upwards. The dissection and section between ligatures of all the biliary branches that drain the hepatic lobes is possible by using a binocular operative microscope (Zeiss, OPMI 1-FR, Oberkochen, Germany). First, the biliary branch of the caudate lobe and then the biliary branch of the right lateral lobe are dissected, ligated and sectioned close to the hepatic parenchyma (Figure 3). The upward dissection of the extrahepatic biliary tract makes it possible to individualize, ligate and section the biliary

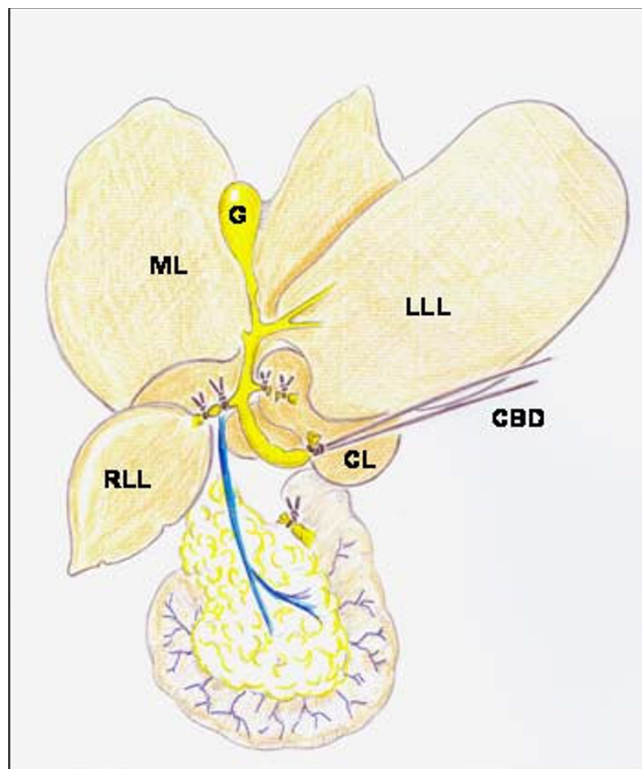


**Figure 3**  
**Microsurgical technique of extrahepatic cholestasis in the rat.** The common bile duct and the lobular bile ducts are sectioned between ligations. The dissection, ligation and sectioning of the lobular bile ducts must be performed without damaging either the portal or arterial vascularization of these lobes. CL, caudate lobe; LLL, left lateral lobe; ML, middle lobe; RLL, right lateral lobe.

branches draining the middle lobe and, finally, the same procedure is followed with the biliary branch of the left lateral lobe [22,24] (Figure 3).

An advantage of the microsurgical technique of extrahepatic cholestasis in the rat is the absence of large biliary pseudocyst formation, which would explain why early mortality is not present. It is possible that the absence of the hilus pseudocyst in this microsurgical model of cholestasis decreases the incidence of hepatopulmonary infection and thus prevents mortality related to sepsis [22,24].

The macrosurgical extrahepatic cholestasis in mice consists normally in double ligation of the common bile duct with 4-0 braided silk sutures and then, sectioning between the ligatures. Finally, the cystic duct is ligated [25]. However, the microsurgical technique can be also applied to mice. The mouse liver, just like the rat's, is composed of four lobes, with the same names (Figure 4). The main difference is that the mouse liver has a gall bladder. That is why, if BDL is performed, it is followed by a marked dila-



**Figure 4**  
**Microsurgical technique for producing extrahepatic cholestasis in mice.** CBD, common bile duct with a long ligature to facilitate handling; CL, caudate lobe; D, duodenum; G, gall bladder; LLL, left lateral lobe; LM, medial lobe; RLL, right lateral lobe.

tion of the gall bladder, which may lead to perforation and choleperitoneum [25].

Microsurgical extrahepatic cholestasis in the mouse also consists in the resection of the four lobular bile ducts in continuity with the common bile duct (Figure 4), but cholecystectomy must be added. The dissection, ligation and section of the bile ducts from the middle and left lateral lobes are simplified if the dissection of the gall bladder and the cystic duct is performed beforehand (Figure 5).

The use of broad-spectrum prophylactic antibiotics and weekly administration of vitamin K (8 mg.kg<sup>-1</sup> intramuscularly) has been proposed to reduce the early death of the animals [25-27]. In rats with microsurgical extrahepatic cholestasis, the weekly administration of antibiotics and vitamin K makes it possible for rodents to survive over 8 weeks.

In the long-term evolution, both macrosurgical (BDL) and microsurgical experimental cholestasis models develop hepatomegaly with a marked ductular proliferation and fibrosis, but the loss of normal liver architecture, typical of cirrhosis, is seldom found [18,24,25,28]. In relation to extrahepatic alterations, jaundice, choluria [27,28], portal hypertension with enlarged spleen and collateral portosystemic circulation [24,27-30], hepatic encephalopathy [31,32] and ascytes [27] stand out. Therefore, experimental extrahepatic cholestasis is not only a good model for studying the hepatic pathology related to biliary obstruction, but also for studying extrahepatic complications.

However, the aim of this review is limited to coverage of hepatic pathology related to obstructive cholestasis. The etiopathogenic mechanisms described in its production could be compared to those that play the main role in the evolution of inflammatory response related to other injuries. [7,8,33]. It is worth mentioning that in obstructive cholestasis and other inflammatory conditions, the tissue alterations mainly occur in the interstitial space. That is why their alterations and their respective production mechanisms are mainly referred to in this context.

### The interstitial space and the inflammatory response

The interstitial space always seems to be the battlefield for inflammation, whether it is due to trauma [7,8,34], infection [8,34] or tumors [35-38].

The successive pathophysiological mechanisms that develop in the interstitial space of tissues when they undergo acute post-traumatic inflammation are considered increasingly complex trophic functional systems for



**Figure 5**  
**Cholecystectomy during the production of extrahepatic cholestasis in mice.** The gall bladder is grasped with forceps near the fundus. The cystic artery is divided by caught and then the mesentery is cut with scissors (on top). The gall bladder is held in the left hand and the cystic duct is cleared of soft tissue by gentle blunt dissection. Then, the lobular bile ducts of the median and left lateral lobes (on bottom) are sectioned between ligations.

using oxygen [7,8,34]. Although their extent is apparently different, the hypothetical similarity between local and systemic responses to mechanical injury could be attributed to a general response mechanism to injury in the body. This mechanism is based on the successive and predominant expression of the nervous, immune and endocrine pathological functions [7,8].

The nervous or immediate functional system has ischemia/reperfusion and edema, which work by diffusion through the injured tissue. This trophic mechanism has a low energy requirement that does not require oxygen (ischemia), or in some circumstances the oxygen is not correctly used, with subsequent development of reactive oxygen and nitrogen species (ROS/RNS) (reperfusion) (Table 1) [7,8].

The immune or intermediate functional system produces infiltration of the injured tissue by inflammatory cells, especially by leukocytes. The immune cell residents in the interstitial space of the affected tissues and organs are also activated. Hence the capacity of these inflammatory cells for extracellular digestion by enzyme release (fermentation) and intracellular digestion (phagocytosis) could be associated with a hypothetical trophic capacity. Improper use of oxygen persists in this immune phase. Also during this phase the lymphatic circulation continues to play an important role [7,8,34,39] (Table 1).

During the evolution of the nervous and immune phase of the inflammatory response, the body loses its more specialized functions and structures. In this progressive deconstruction, depletion of the hydrocarbonate, lipid and protein stores occurs, as well as successive dysfunction and posterior failure, apoptosis, autophagy or necrosis of the specialized epithelium (that is, gastrointestinal, hepatic, pulmonary and renal). Although these alterations are considered a harmless consequence of the inflamma-

tory response, they are also mechanisms through which there is a redistribution of carbohydrates, lipids and amino acids in the body. Consequently, the redistribution of metabolic resources responds to the different trophic requirements of the body as the inflammation progresses. Nevertheless, consumption of the substrate deposits and the dysfunction or failure of the specialized epithelia could also represent an accelerated process of epithelial dedifferentiation [40].

The hypothetical ability to involute or dedifferentiate could constitute an effective defense mechanism against injury since it could make retracing a well-known route possible, (that is, the prenatal specialization phase during the last or endocrine phase of the inflammatory response). This specialization would require or return to prominence of oxidative metabolism, and thus angiogenesis, in the affected epithelial organs, to create a capillary bed that would make regeneration of the specialized epithelial cells possible or for carrying out the repair through fibrosis or scarring [7,8,34,39,41] (Table 1).

**The liver interstitium in obstructive cholestasis**

If we consider that the interstitial alterations produced during the inflammatory response are common to different conditions, the successive pathophysiological mechanisms that develop in the interstitial space of the tissues when they undergo acute traumatic inflammation [7,8] would also be expressed in the liver interstitium when suffering inflammation related to obstructive cholestasis. Thus, in the experimental cholestatic obstructive inflammatory liver disease, three inflammatory phenotypes would be expressed during its evolution: the ischemia/reperfusion phenotype (nervous), the leukocyte phenotype (immune), and the angiogenic phenotype (endocrine).

**Table 1: Phases of the inflammatory interstitial response**

| Phase                      | Response   |
|----------------------------|--|
| I. Immediate or nervous    | Ischemia/reperfusion<br>Edema<br>Oxidative and nitrosative stress  |
| II. Intermediate or immune | Activation of resident inflammatory cells<br>Infiltration by inflammatory cells<br>Toxin and bacterial translocation<br>Enzymatic stress |
| III. Late or endocrine     | Angiogenesis<br>Cell proliferation<br>Cell specialization<br>Energetic stress  |

The hepatic interstitium shares both biochemical and structural characteristics with other interstitial spaces of the body. The molecules secreted by the cells occupying the body tissues interact to create a complex network, which constitutes the ECM. As there are many different tissues in the body, there are also different organizations of cells and matrices [42].

Phylogenetic data generated from recently completed genome sequencing projects have shown that the molecules of the ECM, especially those related to cell-matrix adhesion are 'ancient and exquisitely conserved in multicellular animals' [42,43]. In general, two classes of molecules are produced by the ECM: fibrous proteins (collagen, laminin and elastin) and glycosaminoglycans (GAGs) that can be non-sulfated (hyaluronic acid) and sulfated [42]. Because of their high net negative charge, GAGs and proteoglycans play pivotal roles in biological processes, such as permeoselectivity of basement membranes, activation of chemokines and cytokines, cell-cell interactions and sequestration of growth factors [44,45]. This is the reason why their change has important implications in proinflammatory and anti-inflammatory activities [46-49].

Immediately after complete bile duct obstruction in the rat, an intense increase (60%) in biliary ductal pressure is produced [50] and this is quickly followed by pathological ECM changes [51]. By contrast, biliary decompression, by relieving mechanical stress, reverses liver lesions induced by BDL [52,53]. These experiments reflect the major importance that mechanical energy has in the etiopathology of liver injury in relation to biliary obstruction.

The response of the murine liver to the biliary obstructive injury implies its transcriptional reprogramming favoring the activation of genes regulating metabolism, cell proliferation and matrix remodeling in a time-restricted and sequential fashion [54]. Although there are a dominant activation of metabolic genes in all phases following BDL, from the immediate (1 day) to the later (21 days) phase, involvement of specific pathways varied according to the duration of obstruction [54]. Moreover, where some genes are upregulated, (that is, genes related to disruption of lipid metabolism and fibrosis) in the early stage of cholestasis, other genes are downregulated, (that is, genes involved in mechanisms of cell protection against the accumulation of toxic bile acids) [55].

The three inflammatory phenotypes hypothetically expressed in the murine liver interstitium during long-term cholestasis induced by BDL could help to integrate the etiopathogenic mechanisms that have been described. These inflammatory phenotypes would associate the

genetic factors (up- and downregulated) with metabolic and histological alterations.

### **The ischemia/reperfusion phenotype**

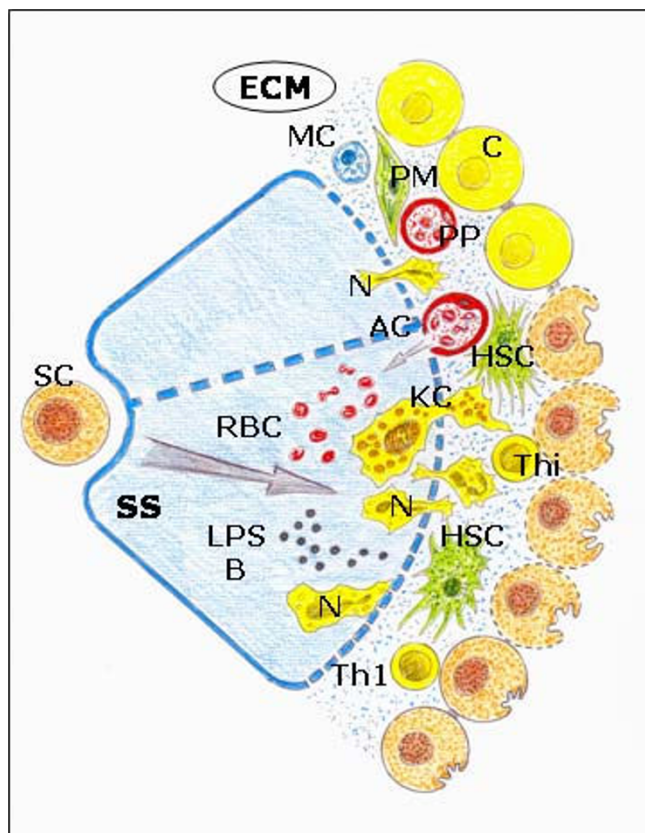
After BDL, the liver rat suffers severe hemodynamic alterations, both portal and arterial, to which the effects of ischemia/reperfusion and oxidative stress can be attributed. The increase of vascular resistance in the liver portal system related to extrahepatic cholestasis results in portal hypertension [56,57] and liver ischemia, associated with a deficient production of inducible nitric oxide synthase (iNOS) and NO [58].

The biliary tree is nourished by the peribiliary plexus [59,60] and around the smaller ducts the plexus gets progressively simpler and thinner [59]. That is why it could be assumed that the increase in intraductal pressure with bile duct dilatation in extrahepatic cholestasis could induce compression of the peribiliary plexus and, consequently bile tract ischemia. However, it has been described that after the decrease in portal vein flow, an increase in hepatic arterial blood flow or a 'hepatic arterial buffer response' is produced [61]. Furthermore, after 2 weeks of BDL in the rat, a significant peribiliary plexus proliferation is produced that is drained by small venules in both the portal vein branches and hepatic sinusoids [59]. Ischemia/reperfusion injury has been in turn involved in the pathogenesis of intrahepatic cholestasis [62].

Rats subjected to BDL could have excessive accumulation of hydrophobic bile acids, which are considered the main cause of hepatotoxicity [12]. They exhibited partial impairment of mitochondrial electron transport chain functions in the liver and oxidative stress [12,27]. Retention and accumulation of hydrophobic bile salts (that is, tauro- and glycochenodeoxycholate) may decrease antioxidant activities of hepatic catalase, glutathione peroxidase, reduced glutathione (GSH) and superoxide dismutase levels and induce hepatocyte necrosis by activating mitochondrial membrane permeability transition [63-67] (Figure 6).

It is accepted that there is a strong correlation between experimental obstructive jaundice and oxidative stress [64,65,68]. However, BDL mainly impairs the liver ability of antioxidant regeneration, especially at the mitochondrial level [66]. Thus, it has been demonstrated that treatment with antioxidants improves the hepatic cellular redox status [68,69]. Indeed, antioxidants have a protective effect on hepatocellular integrity and liver functions by inhibiting reactive oxygen species formation [66,67,69].

In summary, in this early stage of BDL the insufficient supply of oxygen suffered by the liver, related to hemody-



**Figure 6**  
**Ischemia-revascularization and leukocytic phenotypes during the evolution of obstructive cholestasis.**  
 Predominance of oxidative stress with edema and enzymatic stress with infiltration by leukocytes and Kupffer cell activation. AC: arterial capillar; B: bacteria; C: cholangiocyte; ECM: extracellular matrix; HSC: hepatic stellate cell; KC: Kupffer cell; LPS: lipopolysaccharide; MC: mast cell; N: Neutrophyl; PM: portal myofibroblast; PP: peribiliary arterial plexus; RBC: red blood cells; SC: stem cells; SS: sinusoidal space; Th<sub>1</sub>: T cell h<sub>1</sub>; Thi: intraepithelial lymphocyte.

dynamic alterations, as well the incorrect use of oxygen derived from bile salt hepatotoxicity, would constitute the essential factors that would induce the reduction of hepatic energy metabolism. Consequently, the liver reduces its functional capacity to meet tissue metabolic needs.

Oxidative liver damage could decrease the intracellular content of proteins participating in energy production and membrane function, (that is, proteins regulating water and ion transport) [66] inducing cellular and interstitial edema. Also, increased hepatic lipid peroxidation, a high oxidative stress marker [66], can occur with increased membrane permeability, increased degradation of components of the ECM and edema [70]. The accumulation of fragments of GAG has been proposed as an important

mechanism for edema formation because of the hydrophilic properties of GAGs, and particularly of hyaluronan [49,70]. GAGs attract and entrap water and ions, thereby forming hydrated gels, while permitting the flow of cellular nutrients [42,49,71]. Under inflammatory conditions hyaluronan is more polydisperse with a preponderance of lower-molecular forms, and favors edematous infiltration of the tissues [49] as well as the interstitial fluid flow and the tissue lymph pressure gradient [72]. Additionally, mechanical strain by bile tree dilatation related to BDL can lead from mechanotransduction to modifications in proteoglycans and GAGs remodeling the interstitium [73,74]. Matrix stiffness and the mechanical tension that results from cellular adhesion to stiff substrates are instrumental in determining the phenotype of many cells in culture [75].

Early mechanical stiffness has been described in the rat CCl<sub>4</sub> model of fibrosis. This increase in liver stiffness appears to result from matrix crosslinking, and possibly other unknown variables, in addition to matrix quantity [76]. Also, early changes in mechanical stiffness of the liver could induce myofibroblast differentiation in early liver diseases [76,77].

The myofibroblast function endows activated hepatic stellate cells with the ability to behave like smooth muscle cells. Accordingly, activated stellate cells respond by contraction to vasoactive substances. Also, the subsequent constriction potentially regulates the diameter of liver vasculature and affects hepatic blood flow and pressure [78].

### The leukocyte phenotype

Acquiring an active immune phenotype through the cholestatic liver involves both parenchymal (hepatocytes and cholangiocytes) and non-parenchymal cells (sinusoidal endothelial cells, Kupffer cells and hepatic myofibroblasts); blood cells that migrate to the liver interstitium [77-81]. However, the interstitial space seems to orchestrate the inflammatory immune cell activity post BDL in the rat.

In particular, ECM fragments and their receptors exhibit important effects on inflammatory cells and therefore are considered to be clearly implicated in the evolution of the immune interstitial response [16,45,80]. Matrix metalloproteinase (MMPs) are a family of enzymes that degrade components of the ECM and are expressed in the diseased tissues that are inflamed [82], and are particularly present in cholestatic liver injury [83,84]. The enzymatic destruction of the ECM causes the immediate release of the mediators sequestered in its network [44,45]. Furthermore, some fragments of the ECM are molecules that have proinflammatory functions, which can enhance the immune response by activating innate and acquired



immune responses [80]. Fragments of ECM proteins and hyaluronan have the ability to promote inflammation by binding Toll-like receptors (TLR)-4 and TLR-2 [81], by activating the transcriptional regulatory complex of nuclear factor (NF) $\kappa$ B/I $\kappa$ B $\alpha$  and by production of proinflammatory cytokines (that is, TNF $\alpha$ , interleukin (IL)1 $\beta$ ) and chemokines that induce the activation and interstitial recruitment of leukocytes [80].

Upon activation, T cells undergo polarization with different cytokine profiles. Type 1 (Th1) produces interferon (IFN) $\gamma$  and IL2 and type 2 (Th2) produces IL4, IL5, IL9, IL10 and IL13. In particular, Th2 cytokines are mostly involved in mediating allergic inflammation and chronic fibroproliferative disorders [79].

The liver tissue macrophages, or Kupffer cells, are mainly found in the periportal area of the lobule and, due to their location, could play a key role in ischemia/reperfusion injury [85]. But Kupffer cells are also involved in liver inflammation mediated by cholestasis through the release of biologically active substances that promote the immunopathogenic process [86] (Figure 6). Kupffer cells are clearly altered in biliary obstruction [86], with an increased phagocytic ability and a marked proinflammatory response to endotoxin and the lipopolysaccharide binding protein (LBP), which are both increased in extrahepatic cholestasis [85-87]. The hypersensitivity to endotoxin in cholestasis is the cause of increased proinflammatory cytokine synthesis and increased lipid peroxidation [88], with a worsening of apoptosis and trigger progressing to necrosis [89]. However, depletion of Kupffer cells aggravates hepatocellular necrosis and inflammation in cholestatic mice [90]. The LPS-induced proinflammatory response is downregulated by high-density lipoproteins (HDL) that decrease the hepatic proinflammatory signals, restores eNOS activity and lowers portal pressure [91].

Neutrophils are key components of the initial inflammatory response to liver cholestatic injury [92]. In experimental extrahepatic cholestasis, neutrophil interstitial infiltration occurs in early phases, 3 days after BDL [5,93,94]. Biliary cells contribute to hepatic inflammation by producing neutrophil chemoattractants [4]. In long-term BDL, rats continue to show an important cell migration around the portal triad and the central vein, associated with a proinflammatory cytokine liver increase [95]. However, proinflammatory cytokines mediate a heterogeneous hepatocyte response to cholestatic stimuli, with selective hepatocyte down regulation in the periportal zone [96] (Figure 6).

Both cells, HSCs and myofibroblasts, present in the liver interstitium have the ability to express an immune pheno-

type. In particular, HSCs secrete a broad spectrum of inflammatory mediators, (that is, chemokines, MCP-1 and RANTES). PAF (platelet activation factor), IL8 and leukocyte adhesion molecules (ICAM-1, VCAM) are required for the recruitment and activation of leukocytes in the interstitium [6,13,14,97,98]. The homing of these cells to the liver interstitial space is favored by HSCs since these cells express MMPs, which enhance the degradation of the extracellular matrix [14,97,98]. Therefore, HSCs change the initial contractile phenotype to the immune phenotype. It is considered that these phenotypes are intricately related and even interdependent [97] (Figure 6).

Bacterial translocation is a complication of portal hypertension that is capable of inducing proinflammatory cytokines [99], and therefore is produced in BDL rats [99,100]. Furthermore, bacterial translocation provides a mechanism for the pathogenesis of bacterial infections in experimental cholestasis [100]. Increased production of TNF $\alpha$  may play an important role in the process of bacterial translocation in rats with cirrhosis and ascitis because TNF $\alpha$  blockade is able to downregulate it without increasing the incidence of systemic infections [101].

It has been proposed that the immune response, with expression of pro and anti-inflammatory mediators and recruitment of immune cells, may differ over the course of time of obstructive jaundice [102]. Thus, after the initial proinflammatory immune response, a regulating anti-inflammatory activity is established [80] in which T cells and mast cells could participate [103-106]. Dendritic cell differentiation in a cholestatic hepatic environment may lead to Th2 polarization and secretion of IL4 and IL10, rather than IFN $\gamma$  [107]. In the presence of extrahepatic biliary obstruction, the activation of p38, c-Jun N-terminal kinase (JNK) and extracellular signal-regulated kinase (ERK) considered 'stress kinases' [108] would be produced. p38 MAP kinase in particular has been suggested to regulate IL10 synthesis through activation of Sp1 transcription factor rather than through the NF $\kappa$ B pathway [109]. Since IL10 expression is significantly upregulated 14 days after BDL mice [102], anti-inflammatory mediators may modulate the production of proinflammatory cytokines in long-term cholestasis, thus resulting in susceptibility to bacterial translocation and infection [102]. In this way, ECM molecules, like hyaluronan networks, might serve as scaffolds to prevent the loss of ECM components during inflammation and to sequester proinflammatory mediators. That is why a protective or 'counter-inflammatory' role has been suggested for the highly crosslinked hyaluronan [46,47].

Jaundice is also an important mediator of the liver inflammatory response in this experimental model of cholesta-

sis. Bilirubin is produced via reduction of heme-derived biliverdin by biliverdin reductase [110]. Bilirubin has a number of new and interesting biochemical and biological properties [111]. In addition to having a protective role against oxidative stress [111], bilirubin has antiapoptotic and antimutagenic properties [112] as well as a strong role as an immune modulator [113,114]. Thus, in a mouse model of endotoxemia, a single dose of bilirubin in addition to its antioxidant effects also exerts a strong anti-inflammatory activity [114].

Cholestatic jaundice also occurs in the setting of sepsis [114]. Liver abnormalities in sepsis include cholestasis and hyperbilirubinemia. Hyperbilirubinemia particularly develops in sepsis in the setting of bacteremia and precedes positive blood cultures in a third of all cases [115].

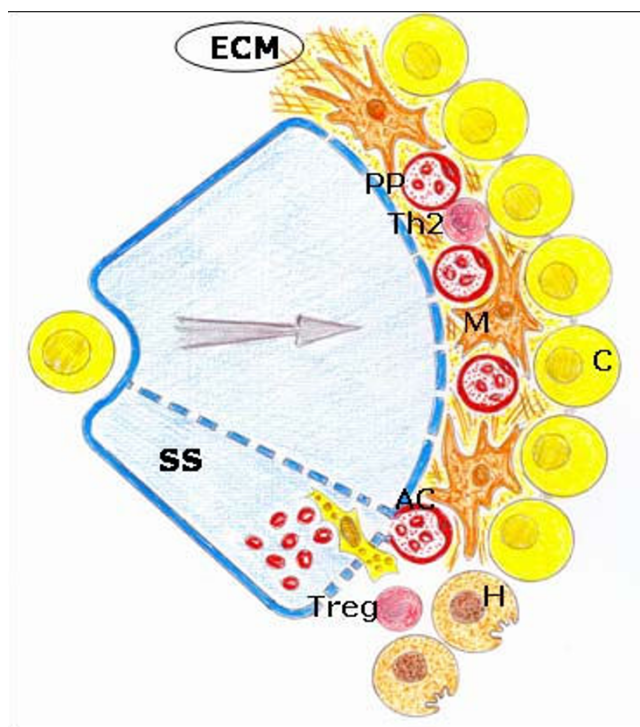
### The angiogenic phenotype

The late evolutive phase in the development of surgical experimental liver cholestasis or endocrine phase is characterized by the predominance of angiogenesis. Angiogenesis is defined as the growth of new vessels from preexisting ones [116].

Although the final objective of endothelial growth is to form new vessels for oxygen, substrates and blood cells (vascular phase), other functions could also be carried out, like antioxidative and anti-immune properties, before the new vessels are formed (prevascular phase) [117,118].

Angiogenesis requires migration of endothelial cells into the interstitial space with the subsequent proliferation and differentiation into capillaries [118]. In BDL rats the proliferation of bile ductules, like in liver organogenesis, precedes the proliferation of the escorting microvessels. After 1 week of BDL, despite the noticeable proliferation of bile ducts, the peribiliary arterial plexus maintains its normal architecture. By contrast, after 2 and 4 weeks of BDL significant microvasculature proliferation is developed, extending from the peribiliary plexus of bile tracts [59] (Figure 7).

The main role of the cholangiocyte function in angiogenesis post BDL has been corroborated, associating hepatic artery ligation to cholestasis induced by BDL in the rat. The liver suffers increased cholangiocyte apoptosis, impaired cholangiocyte proliferation, decreased cholangiocyte vascular endothelial growth factor (VEGF) secretion and the disappearance of the peribiliary plexus. Interestingly enough, cholangiocyte functions and, thus, the integrity of the peribiliary plexus are prevented by treatment with recombinant VEGF-A [119]. Likewise, in human liver transplantation biliary regeneration occurs as an initial proliferation of the epithelial compartment, fol-



**Figure 7**  
**Angiogenic phenotype during the evolution of obstructive cholestasis.** Increase in the proliferation of cholangiocytes with an important development of the peribiliary plexus and sinusoidal arterializations with hepatocytary aplasia. AC: arterial capillar; C: cholangiocyte; ECM: extracellular matrix; H: hepatocyte; M: myofibroblast; PP: peribiliary arterial plexus; SS: sinusoidal space; ; Th<sub>2</sub>: T cell h<sub>2</sub>; Treg: regulatory T cell.

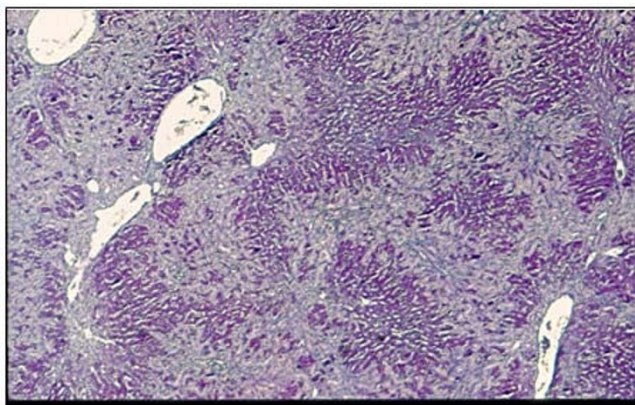
lowed by the vascular compartment, which seems to be supported by induced VEGF-A expression by the epithelial compartment [120].

The ECM plays critical roles in most blood vessel formation processes. In the angiogenic process, ECM components and their fragments provide direction for regulating vessel cell migration, proliferation, differentiation and survival [42]. Integrins are the major type of ECM receptor in endothelial cells [121].

Mast cell hyperplasia is associated with the proliferation of bile ductules during extrahepatic cholestasis [105,106]. These findings suggest that mast cells accumulating in the portal triads may be involved in bile duct proliferation. At the same time, the recanalization of the ligated common bile duct led to an abrupt and transient increase in the number of mast cells associated with a rapid increase in the number of apoptotic biliary epithelial cells. These findings suggested that liver interstitial mast cells may

relate to the hepatic remodeling through the induction of apoptosis [105].

In long-term extrahepatic cholestasis, the predominating hepatic alteration is marked ductular proliferation with a mild portal inflammatory infiltration and apoptosis [122]. However, extrahepatic cholestasis also makes it possible to create a model of biliary fibrosis in the long term [122,123] (Figure 8). Hepatic fibrosis post BDL in rodents is the consequence of an inflammatory process of biliary origin [124,125]. Liver fibrosis is the excessive accumulation of ECM proteins including collagen [6,97,98]. A fundamental concept regarding the pathogenesis of hepatic fibrosis is that the process represents the body's wound-healing response to injury and is similar to the response of other organs to recurrent injury [97]. HSC activation is a key pathogenic feature that underlies liver fibrosis because the resulting myofibroblasts are mainly responsible for connective tissue reassembly [97,98,126]. Multiple and varied stimuli contribute to the induction and maintenance of activation, including oxidative stress, neurotransmitters (norepinephrine), the renin-angiotensin cascade, cytokines (IL4, IL13), chemokines and growth factors, that is, transforming growth factor (TGF)- $\beta$ 1 and connective tissue growth factor (CTGF) [13,77,97,98,122,126,127]. HSCs are further stimulated in a paracrine mode by invaded thrombocytes, polymorphonuclear leucocytes, mast cells and lymphocytes but also by activated Kupffer cells, sinusoidal endothelial cells and hepatocytes to transdifferentiate to myofibroblasts [81,98,122]. HSCs in the liver could also originate from the bone marrow and acquire the myofibroblast phenotype if the adequate, injurious microenvironment of the liver is present [13,98] (Figure 7).



**Figure 8**  
**Periportal biliary proliferation that invades Rappaport spaces I and II.** Peribiliary fibrosis is observed (hematoxylin and eosin (H&E) stain, 50  $\times$  magnification).

Hepatic myofibroblasts are the principal cell type responsible for promoting the deposition of crosslinked fibrillar collagen in the cholestatic liver [6,98]. The coexistence of epithelial-mesenchymal transition from biliary epithelial or hepatocyte cells has also been proposed [13,15,128,129].

During the establishment of liver fibrosis, the persistence of injurious agents and the inflammatory response are followed by 'sinusoidal capillarization', which mainly consists of the transformation of fenestrated hepatic sinusoids into continuous capillaries, accompanied by the deposition of a continuous basement membrane near the endothelial cells and hepatocytes [106]. Mast cells in fibrotic livers can also be involved in hepatic arterialization [106,130]. Capillarization hinders the normal exchanges between plasma and hepatocytes and is the main cause of worsening liver function [130]. In turn, hepatic macrophages can regulate the influx of neutrophils, which may play a direct role in matrix degradation [131]. Also, infiltrating neutrophils seem to accumulate preferentially near the proliferating bile ductules and therefore they could influence the remodeling of biliary epithelial cells [131].

Oxidative and enzymatic stress would, respectively, be produced during ischemia/reperfusion. Furthermore, leukocyte phenotypes in experimental obstructive cholestasis could be involved in the pathogenesis of bile duct epithelial cell proliferation and in the looping and reduplication of the duct and ductules [132]. Thus, it has been suggested that the hepatoprotective effect of honey in BDL rats could be attributed to both to their antioxidant and anti-inflammatory activities [132].

Intense biliary proliferation in the portal spaces characterizes long-term extrahepatic microsurgical cholestasis in the rat. The proliferating bile ducts invade zones 1 and 2 or Rappaport acinus, but not zone 3 or the pericentral zone [28]. This is why it could be considered an 'atypical' proliferation [11]. In essence, the pathophysiologic response of the liver, when bile flow (extrahepatic cholestasis) and/or portal venous (sinusoidal capillarization with portal hypertension) are impaired, is atrophy of the involved hepatic area and hypertrophy of the uninvolved area [133]. Histopathologically the atrophy-hypertrophy complex is characterized by septal fibrosis in the atrophic liver with biliary piecemeal necrosis, apoptosis and ductular proliferation [132,133] (Figure 7).

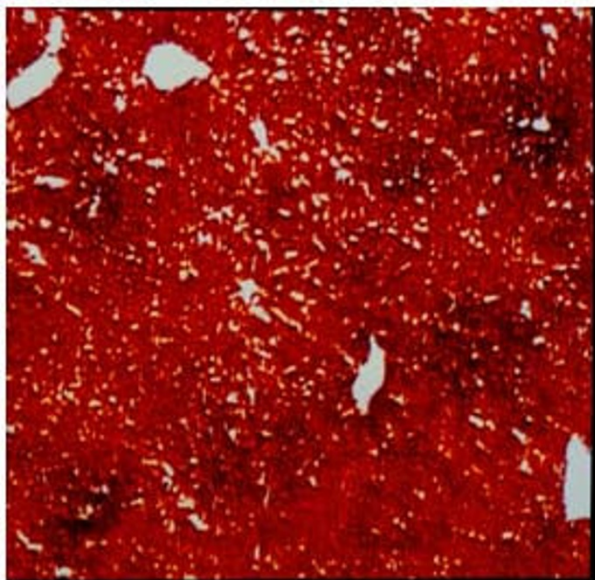
Cholangiocytes are considered biologically important epithelia because of the diverse array of cellular processes in which they participate, including transport of water, ions and solutes [134]. The cholangiocytes have been proposed to be the principle target cell for bile acids in the

liver. Bile acids significantly alter cholangiocyte secretion, proliferation and survival [135]. Thus, bile acids can counteract the loss of bile ducts induced by cholinergic denervation in the BDL rat [136]. However, during their intense proliferation in obstructive cholestasis, proliferating cholangiocytes acquire the phenotype of neuroendocrine cells and secrete different substances including neurotransmitters (serotonin) [137], neuropeptides (opioid peptides such as met-enkephalin) [138-141], hormones (prolactin) [142] and their receptors (estrogens) [11] and growth factors, (that is, insulin-like growth factor (IGF), platelet-derived growth factor (PDGF), hepatocyte growth factor (HGF), TGF $\beta$  and VEGF) [11,136]. A great deal of evidence indicates that hepatic progenitor cell activation in the cholestatic liver is regulated by neural and neuroendocrine factors in modulating non-malignant and malignant cholangiocyte biology [136].

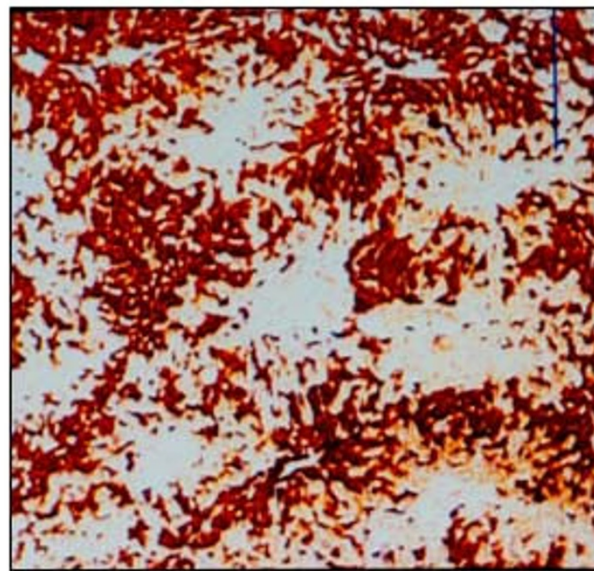
Microsurgical extrahepatic cholestasis decreases the liver cytochrome *c* oxidase activity [143]. Cytochrome oxidation accounts for more than 90% of oxygen consumption by living organisms on earth, and is essential for vital organs such as the liver [144]. Therefore, changes in the phenotype of bile ductule cells in association with a decrease in cytochrome *c* oxidase activity in the cholestatic liver may be attributable to the lower energy requirements

of the neuroendocrine phenotype expressed by these cells [143] (Figure 9).

Bile duct ligation in mice has been widely used to define specific phases of acute and chronic injury and repair in the different cellular compartments of the liver [145]. Bile duct ligation elicits dynamic changes in mouse liver. Acute liver injury, with necrotic and apoptotic cell death and biliary infarcts, is followed by continuous tissue repair, lymphocyte and Kupffer cell infiltration and accumulation of collagen during the second week of postoperative evolution. In this way, Kupffer cells abrogate liver injury in mice by cytokine-dependent mechanisms that include the production of IL6 [146]. It has been demonstrated that endogenous hepatocyte growth factor (HGF) is a reasonable strategy to attenuate hepatic inflammation, necrosis and apoptosis and it has regenerative potential against cholestatic hepatitis [147]. Growth hormone (GH) administration also upregulates hepatocyte proliferation and attenuates fibrogenic response at day 28 of bile duct ligation in mice. Therefore, this endocrine pathway is a potential mechanism to modulate the liver repair response to bile duct ligation [148]. In mice biliary obstruction, as a model of liver repair response to biliary injury, many targeted genes with metabolic-, profibrotic- and proliferation-specific functions are likely involved in



Control



Cholestasis

**Figure 9**  
**Significant inhibition of liver cytochrome oxidase activity after microsurgical extrahepatic cholestasis in the rat.**

the acute phase of injury (first week) and in the chronic phase (4 weeks) [149]. However, although cholangiocellular proliferation occurs early in large bile ducts on days 2–3 and in small bile ducts on day 5, it only produces the rupture of normal liver architecture in the chronic evolutive stages [145]. In summary, these time-related changes in extrahepatic cholestatic mouse are similar to those previously described in rat. Since mice are frequently used in knockout studies, this experimental model could be very useful to study novel mechanistic/molecular biology insights into the pathobiology of obstructive cholestasis in rodent models.

Depression of cholangiocyte mitochondrial respiration in obstructive cholestasis could induce hypoxia-inducible factor (HIF)1 $\alpha$  activation and overexpression, by a similar mechanism to one that has been described in tumor cells [150,151]. This supposed overactivation of HIF1 $\alpha$ , secondary to the decrease in oxygen metabolism with reduced ATP generation, induces the obtaining of energy via other mechanisms. For example, cholangiocytes could generate sufficient reduced nicotinamide dinucleotide phosphate (NADPH) for their biological functions through the continuous replenishment of Krebs cycle intermediates [152]. By these anaplerotic mechanisms, cholangiocytes could obtain sufficient energy not only for the new functions acquired but also for proliferation [152,153].

### Conclusion

Given the plasticity of HSCs and hepatic parenchymal cells (hepatocyte-cholangiocyte axis) it should be kept in mind that while the cholestatic liver develops, they can express the same phenotypes as the post-traumatic inflammatory response [7,8] such as: an ischemic/reperfusion phenotype (hypoxic) a leukocytic phenotype (with pro- and anti-immune response) and finally an angiogenic phenotype with cholangiocyte proliferation and fibrosis.

During these evolutive phases, it could be considered that the cholangiocyte adopts a progressive metabolic complexity (neuroendocrine), which is associated with a growing structural complexity. Cholangiocytes proliferation is a key mechanism capable of conditioning the evolution of liver damage. In fact, proliferating cholangiocytes acquire the phenotype of neuroendocrine cells and secrete different substances that represent the tools of crosstalk with other hepatic cells [11]. In this way, the activation of this neuroendocrine compartment can result in the persistence of the inflammatory response, which would increase the chance for malignant cell transformation. Thus, this 'atypical' biliary proliferation seems to be able to induce an inflammatory response in the remaining liver and includes a concept of premalignancy [11].

In essence, the cholestatic liver changes include fibroblastic cells and extracellular matrix production, inflammation with an immune response, represented by lymphocytes, macrophages and dendritic cells and finally, angiogenesis, shown by newly formed blood vessels [154]. Essentially, all of the elements that constitute the inflammatory response participate in this 'host liver reaction', which may have a trophic purpose for the development of the stiffened biliary cholestatic liver. The persistence of this inflammatory response through a longer evolution would induce an 'atypical' ductular proliferation with the development of a neuroendocrine compartment [11,36-38] and, finally, a malignant cell transformation as it occurs in humans by producing biliary tract cancer (cholangiocarcinoma) [136].

### List of abbreviations

BDL: bile duct ligation; ECM: extracellular matrix; ERK: extracellular signal-regulated kinase; GAG: glycosaminoglycans; HGF: hepatocyte growth factor; HSC: hepatic stellate cell; HDL: high-density lipoprotein; HIF: hypoxia-inducible factor; IGF: insulin-like growth factor; IL: interleukin; iNOS: inducible nitric oxide synthase; IFN $\gamma$ : interferon gamma; JNK: c-Jun N-terminal kinase; LBP: lipopolysaccharide binding protein; LPS: lipopolysaccharide; MMP: matrix metalloproteinase; NADPH: reduced nicotinamide dinucleotide phosphate; NO: nitric oxide; PAF: platelet activation factor; PDGF: platelet-derived growth factor; ROS: reactive oxygen species; RNS: reactive nitrogen species; TLR: Toll-like receptors; TGF: transforming growth factor; TNF $\alpha$ : tumor necrosis factor  $\alpha$ ; VEGF: vascular endothelial growth factor.

### Competing interests

The authors declare that they have no competing interests.

### Authors' contributions

MAA, JA and JLA conceived the integration of inflammation in experimental cholestasis. JLA, MD and JGD supported important clinical aspects and discussed the manuscript. MAA, JLA and JA wrote the final version of the manuscript.

### Acknowledgements

We would like to acknowledge the librarians of the School of Medicine Library (UCM), especially the Director, Juan Carlos Domínguez, María-José Valdemoro, María Elena Vicente for her assistance in preparing the manuscript and Elizabeth Mascola for translating the text into English. This study was supported in part by a Grant from MEC.SEJ 2004/07445 and the Department of Health. Castilla-La Mancha Regional Council (Ref. 04047-00).

### References

- Schaffner F, Popper H: **Classification and mechanism of cholestasis**. In *Liver and biliary disease Pathophysiology Diagnosis Management* Edited by: Wright R, Alberti KGMM, Karran S, Millward-

- Sadler GH. London. Oxford: WB Saunders Company Ltd; 1979:296-323.
2. Sherlock S: **Cholestasis**. In *Diseases of the liver and biliary System* 8th edition. Edited by: Sherlock S. London: Blackwell Scientific Publications; 1989:248-272.
  3. Li MK, Crawford JM: **The pathology of cholestasis**. *Semin Liver Dis* 2004, **24**:21-24.
  4. Saito JM, Maher JJ: **Bile duct ligation in rats induces biliary expression of cytokine-induced neutrophil chemoattractant**. *Gastroenterology* 2000, **118**(6):1157-1168.
  5. Gujral JS, Farhood A, Bajt ML, Jaeschke H: **Neutrophils aggravate acute liver injury during obstructive cholestasis in bile duct-ligated mice**. *Hepatology* 2003, **38**:355-363.
  6. Batailler R, Brenner DA: **Liver fibrosis**. *J Clin Invest* 2005, **115**:209-218.
  7. Aller MA, Arias JL, Nava MP, Arias J: **Posttraumatic inflammation is a complex response based on the pathological expression of the nervous, immune, and endocrine functional systems**. *Exp Biol Med (Maywood)* 2004, **229**(2):170-181.
  8. Aller MA, Arias JL, Sanchez-Patan E, Arias J: **The inflammatory response: an efficient way of life**. *Med Sci Monit* 2006, **12**:RA225-234.
  9. Nakanuma Y, Hosono M, Sanzen T, Sasaki M: **Microstructure and development of the normal and pathological biliary tract in humans, including blood supply**. *Microsc Res Tech* 1997, **38**(6):552-570.
  10. Theise ND, Saxena R, Portman BC, Thung SN, Yee H, Chiriboga L, Kumar A, Crawford JM: **The canals of Hering and hepatic stem cells in humans**. *Hepatology* 1999, **30**:1425-1433.
  11. Alvaro D, Mancino MG, Glaser S, Gaudio E, Marzioni M, Francis H, Alpini G: **Proliferating cholangiocytes: a neuroendocrine compartment in the diseased liver**. *Gastroenterology* 2007, **132**:415-431.
  12. Poli G: **Pathogenesis of liver fibrosis: role of oxidative stress**. *Mol Aspects Med* 2000, **21**:49-98.
  13. Friedman SL: **Protean, multifunctional and enigmatic cells of the liver**. *Physiol Rev* 2008, **88**:125-172.
  14. Iredale JP: **Models of liver fibrosis: exploring the dynamic nature of inflammation and repair in a solid organ**. *J Clin Invest* 2007, **117**:539-548.
  15. Henderson NC, Iredale JP: **Liver fibrosis: cellular mechanisms of progression and resolution**. *Clin Sci* 2007, **112**:265-280.
  16. Adair-Kirk TL, Senior RM: **Fragments of extracellular matrix as mediators of inflammation**. *Int J Biochem Cell Biol* 2008, **40**:1101-1110.
  17. O'Connor M: **Mechanical biliary obstruction. A review of the multisystemic consequences of obstructive jaundice and their impact on perioperative morbidity and mortality**. *Am Surgeon* 1985, **51**:245-251.
  18. Kountouras J, Billing BH, Scheuer PJ: **Prolonged bile duct obstruction: a new experimental model for cirrhosis in the rat**. *Br J Exp Pathol* 1984, **65**:305-311.
  19. Holmberg JT, Hederstrom E, Ihse I: **A method to prevent recanalization of the transected bile duct in the rat**. *Scand J Gastroenterol* 1985, **20**:428-432.
  20. Aller MA, Lorente L, Arias J: **Hepatic artery ischemia in rats with cholestasis. Different response of the lobes**. *J Hepatol* 1991, **13**(Suppl):S87.
  21. Aller MA, Lorente L, Alonso S, Rodriguez G, Arias J: **Different types of response to hepatic arterial ischemia in rats with cholestasis**. *Surg Res Commun* 1993, **14**:269-280.
  22. Aller MA, Duran M, Ortega L, Arias JL, Nava MP, Prieto I, Arias J: **Comparative study of macro- and microsurgical extrahepatic cholestasis in the rat**. *Microsurgery* 2004, **24**:442-447.
  23. Castaing D, Houssin D, Bismuth H: **Anatomy of the liver and portal system of the rat**. In *Hepatic and portal surgery in the rat* Paris: Editorial Masson; 1980:27-46.
  24. Aller MA, Lorente L, Alonso S, Arias J: **A model of cholestasis in the rat, using a microsurgical technique**. *Scand J Gastroenterol* 1993, **28**:10-14.
  25. Abalde JG, Pasarin M, Garcia-Pagan JC: **Animal models of portal hypertension**. *World J Gastroenterol* 2006, **12**:6577-6584.
  26. Beck PL, Lee SS: **Vitamin K1 improves survival in bile-duct-ligated rats with cirrhosis**. *J Hepatol* 1995, **23**:235.
  27. Huang Y-T, Hsu Y-C, Chen C-Y, Liu C-T, Wei Y-H: **Oxidative-stress-related changes in the livers of bile-duct-ligated rats**. *J Biomed Sci* 2003, **10**:170-178.
  28. Aller MA, Nava MP, Arias JL, Duran M, Prieto I, Llamas MA, Arias J: **Microsurgical extrahepatic cholestasis in the rat: a long-term study**. *J Invest Surg* 2004, **17**:99-104.
  29. Chan C-C, Wang S-S, Lee FY, Chang FY, Lin HC, Hou MC, Huang MC, Lee SD: **Effects of endothelin-1 on portal-systemic collaterals of common bile duct-ligated cirrhotic rats**. *Eur J Clin Invest* 2004, **34**:290-296.
  30. Katsuta Y, Zhang X-J, Ohsuga M, Akimoto T, Komeichi H, Shimizu S, Inami T, Miyamoto A, Satomura K, Takano T: **Hemodynamic features of advanced cirrhosis due to chronic bile duct ligation**. *J Nippon Med Sch* 2005, **72**:217-225.
  31. Garcia-Moreno LM, Aller MA, Conejo NM, Gomez MA, Martin FR, Arias J, Arias JL: **Brain Ag-NOR activity in cholestatic rats with hepatic encephalopathy**. *Hepatology* 2002, **24**:275-281.
  32. Garcia-Moreno LM, Conejo NM, Gonzalez-Pardo H, Aller MA, Nava MP, Arias J, Arias JL: **Evaluation of two experimental models of hepatic encephalopathy in rats**. *Braz J Med Biol Res* 2005, **38**:127-132.
  33. Lorente L, Aller MA, Arias JL, Arias J: **Nitric oxide (NO) is a highly reactive free radical with a multitude of organ specific regulatory functions**. *Ann Surg* 1996, **224**:688-689.
  34. Aller MA, Arias JL, Arias J: **Post-traumatic inflammatory response: perhaps a succession of phases with a nutritional purpose**. *Med Hypotheses* 2004, **63**:42-46.
  35. Freitas I, Baronzio GF, Bono B, Griffini P, Bertone V, Sonzini N, Magrassi GR, Bonandrini L, Gerzeli G: **Tumor interstitial fluid: misconsidered component of the internal milieu of a solid tumor**. *Anticancer Res* 1997, **17**:165-172.
  36. Arias JL, Aller MA, Arias J: **The use of inflammation by tumor cells**. *Cancer* 2005, **104**:223-228.
  37. Arias JL, Aller MA, Sanchez-Patan F, Arias J: **Inflammation and cancer: is trophism the link?** *Surg Oncol* 2006, **15**:235-242.
  38. Arias JL, Aller MA, Arias J: **Cancer cell: using inflammation to invade the host**. *Mol Cancer* 2007, **6**:29.
  39. Aller MA, Arias JL, Nava MP, Arias J: **Evolutionary trophic phases of the systemic acute inflammatory response, oxygen use mechanisms and metamorphosis**. *Psicothema* 2004, **16**:369-372.
  40. Selye H: **The part of inflammation in the local adaptation syndrome**. *Rev Can Biol* 1953, **12**:155-175.
  41. Aller MA, Arias JL, Arias JL, Sanchez-Patan F, Arias J: **The inflammatory response recapitulates phylogeny through trophic mechanisms to the injured tissue**. *Med Hypotheses* 2007, **68**:202-209.
  42. Rhodes JM, Simons M: **The extracellular matrix and blood vessels formation: not just a scaffold**. *J Cell Mol Med* 2007, **11**:176-205.
  43. Hynes RO, Zhao Q: **The evolution of cell adhesion**. *J Cell Biol* 2000, **150**:F89-96.
  44. Stern R, Asari AA, Sugahara KN: **Hyaluronan fragments: an information-rich system**. *Eur J Cell Biol* 2006, **85**:699-715.
  45. Yung S, Chan TM: **Glycosaminoglycans and proteoglycans: overlooked entities?** *Perit Dial Int* 2007, **27**:S104-109.
  46. Day AJ, De la Motte CA: **Hyaluronan cross-linking: a protective mechanism in inflammation?** *TRENDS Immunol* 2005, **26**:637-643.
  47. Cantor JO, Nadkarni PP: **Hyaluronan: the Jekyll and Hyde molecule**. *Inflamm Allergy Drug Targets* 2006, **5**:257-260.
  48. Bartlett AH, Hayashida K, Park PW: **Molecular and cellular mechanisms of syndecans in tissue injury and inflammation**. *Mol Cells* 2007, **24**:153-166.
  49. Jiang D, Liang J, Noble PW: **Hyaluronan in tissue injury and repair**. *Ann Rev Cell Dev Biol* 2007, **23**:435-461.
  50. Slott PA, Liu MH, Tavoloni N: **Origin, pattern, and mechanism of bile duct proliferation following biliary obstruction in the rat**. *Gastroenterology* 1990, **99**:466-477.
  51. Desmouliere A, Darby Z, Costa AMA, Raccurt M, Tuchweber B, Sommer P, Gabbiani G: **Extracellular matrix deposition, lysyl-oxidase expression and myofibroblastic differentiation during the initial stages of cholestatic fibrosis in the rat**. *Lab Invest* 1997, **76**:765-778.
  52. Ramm GA, Carr SC, Bridle KR, Li L, Britton RS, Crawford DH, Vogler CA, Bacon BR, Tracy TF: **Morphology of liver repair following cholestatic liver injury: resolution of ductal hyperplasia,**

- matrix deposition and regression of myofibroblasts.** *Liver* 2000, **20**:387-396.
53. Costa AM, Tuchweber B, Lamireau T, Yousef IM, Balabaud C, Rosenbaum J, Desmouliere A: **Role of apoptosis in the remodeling of cholestatic liver injury following release of the mechanical stress.** *Virchows Arch* 2003, **442(4)**:372-380.
  54. Campbell KM, Sabla GE, Bezerra JA: **Transcriptional reprogramming in murine liver defines the physiologic consequences of biliary obstruction.** *J Hepatol* 2004, **40**:14-23.
  55. Kojima K, Hosokawa M, Kobayashi K, Tainaka H, Chiba K: **Microarray analysis of hepatic gene expression during long-term cholestasis induced by common bile duct ligation in rats.** *Res Commun Mol Pathol Pharmacol* 2004, **115-116**:63-75.
  56. Ohara N, Schaffner T, Reichen J: **Structure-function relationship in secondary biliary cirrhosis in the rat. Stereologic and hemodynamic characterization of a model.** *J Hepatol* 1993, **17**:155-162.
  57. Li T, Yang Z: **Research progress of vasculopathy in portal hypertension.** *World J Gastroenterol* 2005, **11**:6079-6084.
  58. Baron V, Hernandez J, Noyola M, Escalante B, Muriel P: **Nitric oxide and inducible nitric oxide synthase expression are downregulated in acute cholestasis in the rat accompanied by liver ischemia.** *Comp Biochem Physiol C Toxicol Pharmacol* 2000, **127(3)**:243-249.
  59. Gaudio E, Onori P, Pannarale L, Alvaro D: **Hepatic microcirculation and peribiliary plexus in experimental biliary cirrhosis: a morphology study.** *Gastroenterology* 1996, **111**:1118-1124.
  60. Li WG, Hu SH, Xue BD, Jiang ZG, Huang ZQ: **Observation of hepatohilar peribiliary vascular plexus with complete absence of hepatic artery blood supply in rats.** *Transplant Proc* 2007, **39**:3424-2428.
  61. Lauth WW: **Relationship between hepatic blood flow and overall metabolism: the hepatic arterial buffer response.** *Fed Proc* 1983, **42**:1662-1666.
  62. Tanaka Y, Chen C, Maher JM, Klaassen CD: **Kupffer cell-mediated downregulation of hepatic transporter expression in rat hepatic ischemia-reperfusion.** *Transplantation* 2006, **82**:258-266.
  63. Tahan G, Tarcin O, Tahan V, Eren F, Gedik N, Sahani S, Biberoglu N, Guzel S, Bozbas A, Tuzun N, Yucel O: **The effects of N-acetylcysteine on bile-duct ligation-induced liver fibrosis in rats.** *Dig Dis Sci* 2007, **52**:3348-3354.
  64. Wang G, Shen H, Rajaraman G, Roberts MS, Gong Y, Jiang P, Burczynski F: **Expression and antioxidant function of liver fatty acid binding protein in normal and bile-duct ligated rats.** *Eur J Pharmacol* 2007, **560**:61-68.
  65. Orellana M, Avalos N, Abarca M, Valdes E: **Microsomal and peroxisomal fatty acid oxidation in bile duct ligated rats: a comparative study between liver and kidney.** *Gen Pharmacol* 1997, **28(4)**:525-529.
  66. Portincasa P, Grattagliano I, Testini M, Caruso ML, Wang DQH, Moschetta A, Calamita G, Vacca M, Valentini AM, Renna G, Lissidini G, Palasciano G: **Parallel intestinal and liver injury during early cholestasis in the rat: modulation by bile salts and antioxidants.** *Free Rad Biol Med* 2007, **42**:1381-1391.
  67. Assimakopoulos SF, Maroulis I, Patsoukis N, Vagenas K, Scopa CD, Georgiou CD, Vagianos CE: **Effect of antioxidant treatments on the gut-liver axis oxidative status and function in bile duct-ligated rats.** *World J Surg* 2007, **31**:2023-32.
  68. Assimakopoulos SF, Vagianos CE, Patsoukis N, Georgiou C, Nikolopoulou V, Scopa CD: **Evidence for intestinal oxidative stress in obstructive jaundice-induced gut barrier dysfunction in rats.** *Acta Physiol Scand* 2004, **180**:177-185.
  69. Soylu AR, Aydogdu N, Basaran UN, Altaner S, Tarcin O, Gedik N, Umüt H, Tezel A, Dokmeci G, Galoglu H, Ture M, Kutlu K, Kaymak K: **Antioxidants vitamin E and C attenuate hepatic fibrosis in biliary obstructed rats.** *World J Gastroenterol* 2006, **12**:6835-6841.
  70. Kennett EC, Davies MJ: **Degradation of matrix glycosaminoglycans by peroxynitrite/peroxynitrous acid: evidence for a hydroxyl-radical-like mechanism.** *Free Rad Biol Med* 2007, **42**:1278-1289.
  71. Alberts B, Bray D, Lewis J, Raff M, Roberts K, Watson JD: **Molecular biology of the cell.** In *Cell-cell adhesion and the extracellular matrix* New York: Garland Publishing Inc; 1983:673-715.
  72. Chen B, Fu B: **A model for charged molecule transport in the interstitial space.** *Conf Proc IEEE Eng Med Biol Soc* 2005, **1**:55-58.
  73. Evanko SP, Tammi MI, Tammi RH, Wight TN: **Hyaluronan-dependent pericellular matrix.** *Adv Drug Deliv Rev* 2007, **59**:1351-1365.
  74. Ingber DE: **Cellular mechanotransduction: putting all the pieces together again.** *FASEB J* 2006, **20**:811-827.
  75. Bischofs IB, Schwarz US: **Cell organization in soft media due to active mechanosensing.** *Proc Natl Acad Sci USA* 2003, **100**:9274-9279.
  76. Georges PC, Hui JJ, Gombos Z, McCormick ME, Wang AY, Uemura M, Mick R, Janney PA, Furth EE, Wells RG: **Increased stiffness of the rat liver precedes matrix deposition: implications for fibrosis.** *Am J Physiol Gastrointest Liver Physiol* 2007, **293**:G1147-G1154.
  77. Muddu AK, Guha IN, Elsharkawy AM, Mann DA: **Resolving fibrosis in the diseased liver: translating the scientific promise to the clinic.** *Int J Biochem Cell Biol* 2007, **39**:695-714.
  78. Winau F, Quack C, Darmoise A, Kaufmann SHE: **Starring stellate cells in liver immunology.** *Curr Opin Immunol* 2008, **20**:68-74.
  79. Azouz A, Razzaque MS, El-Hallak M, Taguchi T: **Immunoinflammatory responses and fibrogenesis.** *Med Electron Microsc* 2004, **37**:141-148.
  80. Morwood SR, Nicholson LB: **Modulation of the immune response by extracellular matrix proteins.** *Arch Immunol Ther Exp (Warsz)* 2006, **54(6)**:367-374.
  81. Szabo G, Mandrakar P, Dolganiuc A: **Innate immune response and hepatic inflammation.** *Semin Liver Dis* 2007, **27(4)**:339-350.
  82. Cuenca J, Martin-Sanz P, Alvarez-Barrientos AM, Bosca L, Goren N: **Infiltration of inflammatory cells plays an important role in matrix metalloproteinase expression and activation in the heart during sepsis.** *Am J Pathol* 2006, **169**:1567-1576.
  83. Roeb E, Purucker E, Breuer B, Nguyen H, Heinrich PC, Rose-John S, Matern S: **TIMP expression in toxic and cholestatic liver injury in rat.** *J Hepatol* 1997, **27**:535-544.
  84. Bergheim I, Guo L, Davis MA, Duveau I, Arteel GE: **Critical role of palmsinogen activator inhibitor-1 in cholestatic liver injury and fibrosis.** *J Pharmacol Exp Ther* 2006, **316(2)**:592-600.
  85. Kolios G, Valatas V, Kouroumalis E: **Role of Kupffer cells in the pathogenesis of liver disease.** *World J Gastroenterol* 2006, **12**:7413-7420.
  86. Minter RM, Fan MH, Niederbichler A, Ipaktchi K, Arabi S, Hemmila MR, Remick DG, Wang SC, Su GL: **Altered Kupffer cell function in biliary obstruction.** *Surgery* 2005, **138**:236-245.
  87. O'Neil S, Hunt J, Filkins J, Gamelli R: **Obstructive jaundice in rats results in exaggerated hepatic production of tumor necrosis factor-alpha and tissue tumor necrosis factor-alpha levels after endotoxin.** *Surgery* 1997, **122**:281-286.
  88. Harry D, Anand R, Holt S, Davies S, Marley R, Fernando B, Goodier D, Moore K: **Increased sensitivity to endotoxemia in the bile duct-ligated cirrhotic rat.** *Hepatology* 1999, **30**:1198-1205.
  89. Moazzam FN, Brems JJ, Yong SL, Filkins JP, Fisher SG, Holt DR, Gamali RL, Ding JW: **Endotoxin potentiates hepatocyte apoptosis in cholestasis.** *J Am Coll Surg* 2002, **194**:731-739.
  90. Gehring S, Dickson EM, San Martin ME, Van Rooijen N, Papa EF, Harty MW, Tracy TF, Gregory SH: **Kupffer cells abrogate cholestatic liver injury in mice.** *Gastroenterology* 2006, **130**:810-822.
  91. Thabut D, Tzai KA, Bonnefont-Rousselot D, Aller M, Farges O, Guimont MC, Tellier Z, Guicharo C, Ogier-Denis E, Poynard T, Moureau R, Le Brech D: **High-density lipoprotein administration attenuates liver proinflammatory response, restores liver endothelial nitric oxide synthase activity, and lowers portal pressure in cirrhotic rats.** *Hepatology* 2007, **46**:1893-1906.
  92. Shivakumar P, Campbell KH, Sabla GE, Miethke A, Tiao G, McNeal MM, Ward RL, Bezerra JA: **Obstructive of extrahepatic bile ducts by lymphocytes is regulated by IFN-gamma in experimental biliary atresia.** *J Clin Invest* 2004, **114**:322-329.
  93. Georgiev P, Jochum W, Heinrich S, Jang JH, Nocito A, Dahm F, Clavien PA: **Characterization of time-related changes after experimental bile duct ligation.** *Br J Surg* 2008, **95**:646-656.
  94. Levy R, Schlaefke F, Keynan A, Nagauker O, Yaari A, Sikuler E: **Increased neutrophil function induced by bile duct ligation in a rat model.** *Hepatology* 1993, **17**:908-914.
  95. Fernandez-Martinez E, Perez-Alvarez V, Tsutsumi V, Shibayama M, Muriel P: **Chronic bile duct obstruction induces changes in plasma and hepatic levels of cytokines and nitric oxide in the rat.** *Exp Toxicol Pathol* 2006, **58**:49-58.
  96. Donner MG, Schumacher S, Warskulat U, Heinehann J, Häussinger D: **Obstructive cholestasis induces TNF- $\alpha$  and IL-1 $\beta$ -mediated**

- periportal downregulation of Bsep and zonal regulation of Ntcp, Oatp1a4, and Oatp1b2. *Am J Physiol Gastroenterol Liver Physiol* 2007, **293**:G1134-G1146.
97. Rockey D: **Hepatic fibrosis, stellate cells, and portal hypertension.** *Clin Liver Dis* 2006, **10**:459-479.
  98. Gressner OA, Weiskirchen R, Gressner AM: **Evolving concepts of liver fibrogenesis provide new diagnostic and therapeutic options.** *Comp Hepatol* 2007, **6**:7.
  99. Garcia-Tsao G, Wiest R: **Gut microflora in the pathogenesis of the complications of cirrhosis.** *Best Pract Res Clin Gastroenterol* 2004, **18**(2):353-372.
  100. Szymmf B, Libert JM, Mougeot C, Lebrec D, Mazmanian M, Humbert M, Herve P: **Cirrhotic rats with bacterial translocation have higher incidence and severity of hepatopulmonary syndrome.** *J Gastroenterol Hepatol* 2005, **20**:1538-1544.
  101. Frances R, Chiva M, Sánchez E, Gonzalez-Navajas JM, Llovet T, Zapater P, Soriano G, Muñoz C, Balazó J, Perez-Mateo M, Song X-Y, Guarner C, Such J: **Bacterial translocation is downregulated by anti-TNF- $\alpha$  monoclonal antibody administration in rats with cirrhosis and ascites.** *J Hepatol* 2007, **46**:797-803.
  102. Morita Y, Yoshidome H, Kimura F, Shimizu H, Ohtsuka M, Taxeuchi D, Mitsuhashi N, Iida A, Miyazaki M: **Excessive inflammation but decreased immunological response renders liver susceptible to infection in bile duct ligated mice.** *J Surg Res* 2008, **146**:262-270.
  103. Rioux KP, Sharkey KA, Wallace JL, Swain MG: **Hepatic mucosal mast cell hyperplasia in rats with secondary biliary cirrhosis.** *Hepatology* 1996, **23**:888-895.
  104. Schramm R, Thorlaciuc H: **Neutrophil recruitment in mast cell-dependent inflammation: inhibitory mechanisms of glucocorticoids.** *Inflamm Res* 2004, **53**:644-652.
  105. Takeshita A, Shibayama Y: **Role of mast cells in hepatic remodeling during cholestasis and its resolution: relevance to regulation of apoptosis.** *Exp Toxicol Pathol* 2005, **56**:273-280.
  106. Franceschini B, Ceva-Grimaldi G, Russo C, Dioguardi N, Grizzi F: **The complex functions of mast cells in chronic human liver diseases.** *Dig Dis Sci* 2006, **51**:2248-2256.
  107. Cabillie F, Rougier N, Basset C, Lecovillard I, Quelvenec E, Tojas L, Guguen-Guilouzon C, Corlu A: **Hepatic environment elicits monocyte differentiation into a dendritic cell subset directing Th2 response.** *J Hepatol* 2006, **44**:552-559.
  108. Samuel I: **Bile and pancreatic juice exclusion activates acinar stress kinases and exacerbates gallstone pancreatitis.** *Surgery* 2008, **143**:434-440.
  109. Waseem T, Duxbury M, Ito H, Ashley SW, Robinson MK: **Exogenous ghrelin modulates release of pro-inflammatory and anti-inflammatory cytokines in LPS-stimulated macrophages through distinct signaling pathways.** *Surgery* 2008, **143**:334-342.
  110. Schmidt R, McDonagh AF: **The enzymatic formation of bilirubin.** *Ann NY Acad Sci* 1975, **244**:533-552.
  111. Vitek L, Schwertner HA: **The heme catabolic pathway and its protective effects on oxidative stress-mediated diseases.** *Adv Clin Chem* 2007, **43**:1-57.
  112. Bulmer AC, Ried K, Blanchfield JT, Wagner KH: **The anti-mutagenic properties of bile pigments.** *Mutat Res* 2007, **658**:28-41.
  113. Selye H: **On the mechanism through which obstructive jaundice influences inflammatory processes.** *Ann Rheum Dis* 1954, **13**:102-108.
  114. Moseley RH: **Sepsis and cholestasis.** *Clin Liver Dis* 1999, **3**:465-475.
  115. Marrero J, Martinez FJ, Hyzy R: **Advances in critical care hepatology.** *Am J Respir Crit Care Med* 2003, **168**:1421-1426.
  116. Kurz H, Burri PH, Djonov VG: **Angiogenesis and vascular remodeling by intussusception: from form to function.** *News Physiol Sci* 2003, **18**:65-70.
  117. Aller MA, Arias JL, Arias J: **The mast cell integrates the splanchnic and systemic inflammatory response in portal hypertension.** *J Transl Med* 2007, **5**:44.
  118. Aller MA, Arias JL, Cruz A, Arias J: **Inflammation: a way to understanding the evolution of portal hypertension.** *Theor Biol Med Model* 2007, **4**:44.
  119. Gaudio E, Barbaro B, Alvaro D, Glaser S, Francis H, Franchitto A, Onori P, Ueno Y, Marzioni M, Fava G, Venter J, Reichenbach R, Summers R, Alpini G: **Administration of r-VEGF-A prevents hepatic artery ligation-induced bile duct damage in bile duct ligated rats.** *Am J Physiol Gastrointest Liver Physiol* 2006, **291**:G307-G317.
  120. Heuvel MC Van Den, Gouw ASH, Boot M, Slooff MJH, Poppema S, De Jong KP: **The finest branches of the biliary tree might induce biliary vascularization necessary for biliary regeneration.** *J Hepatol* 2006, **44**:947-955.
  121. Hynes RO: **Integrins: bidirectional allosteric signalling machines.** *Cell* 2002, **110**:673-687.
  122. Sanchez-Patan F, Anchuelo R, Corcuera MT, Casado I, Gomez-Aguado F, Aller MA, Cruz A, Alonso MJ, Arias J: **Biliary fibrosis in microsurgical extrahepatic cholestasis in the rat.** *Microsurgery* 2008, **28**:361-366.
  123. Aube C, Moal F, Oberti F, Roux J, Croquet V, Gallois Y, Argand C, Caron C, Cales P: **Diagnosis and measurement of liver fibrosis by MRI in bile duct ligated rats.** *Dig Dis Sci* 2007, **52**:2601-2609.
  124. Kershenobich SD, Weissbrod AB: **Liver fibrosis and inflammation. A review.** *Ann Hepatol* 2003, **2**:159-163.
  125. Li MK, Crawford JM: **The pathology of cholestasis.** *Sem Liver Dis* 2004, **24**:21-42.
  126. Proell V, Carmona-Cuenca I, Murillo MM, Huber H, Fabregat I, Mikulits W: **TGF- $\beta$  dependent regulation of oxygen radicals during transdifferentiation of activated hepatic stellate cells to myofibroblastoid cells.** *Comp Hepatol* 2007, **6**:1.
  127. Sedlaczek N, Jia JD, Bauer M, Herbst H, Ruelh M, Hahn EG, Schuppan D: **Proliferating bile duct epithelial cells are a major source of connective tissue growth factor in rat biliary fibrosis.** *Am J Pathol* 2001, **158**:1239-1244.
  128. Cassiman D, Libbrecht L, Desmet V, De Nef C, Roskams T: **Hepatic stellate cell/myofibroblast subpopulations in fibrotic human and rat livers.** *J Hepatol* 2002, **36**:200-209.
  129. Sicklick JK, Choi SS, Bustamante M, McCall SJ, Hernandez-Perez E, Huang J, Li Y-X, Rojkind M, Diehl AM: **Evidence for epithelial-mesenchymal transitions in adult liver cells.** *Am J Physiol Gastrointest Liver Physiol* 2006, **291**:G575-G583.
  130. Zimmermann A, Zhao D, Reichen J: **Myofibroblasts in the cirrhotic rat liver reflect hepatic remodeling and correlate with fibrosis and sinusoidal capillarization.** *J Hepatol* 1999, **30**(4):646-652.
  131. Harty MW, Papa EF, Huddleston HM, Young E, Nazaret S, Riley CA, Ramm GA, Gregory SH, Tracy TF: **Hepatic macrophages promote the neutrophil-dependent resolution of repairing cholestatic rat livers.** *Surgery* 2008, **143**:667-78.
  132. Kilicoglu B, Gencay C, Kismet K, Kilicoglu SS, Erguder I, Erel S, Sunay AE, Erdemli E, Durak I, Akku AM: **The ultrastructural research of liver in experimental obstructive jaundice and effect of honey.** *Am J Surg* 2008, **195**:249-256.
  133. Black DM, Behrns KE: **A scientist revisits the atrophy-hypertrophy complex: Hepatic apoptosis and regeneration.** *Surg Oncol Clin N Am* 2002, **11**:849-864.
  134. Boger PT, Larusso NF: **Cholangiocyte biology.** *Curr Opin Gastroenterol* 2007, **23**:299-305.
  135. Xia X, Francis H, Glaser S, Alpini G, Lesage G: **Bile acid interactions with cholangiocytes.** *World J Gastroenterol* 2006, **12**:3553-3563.
  136. Marzioni M, Fava G, Benedetti A: **Nervous and neuroendocrine regulation of the pathophysiology of cholestasis and of biliary carcinogenesis.** *World J Gastroenterol* 2006, **12**:3471-3480.
  137. Marzioni M, Glaser S, Francis H, Marucci L, Benedetti A, Alvaro D, Taffetani S, Ueno Y, Roskams T, Phinizia JL, Venter J, Fava G, Lesage GD, Alpini G: **Autocrine/paracrine regulation of the growth of the biliary tree by the neuroendocrine hormone serotonin.** *Gastroenterology* 2005, **128**:121-137.
  138. Moezi L, Shafaroodi H, Sarkar S, Emami-Razavi SH, Sanatkar M, Mirazi N, Dehpour AR: **Involvement of nitroergic and opioidergic systems in the hypothermia induced by cholestasis in rats.** *Pathophysiology* 2006, **13**:227-232.
  139. Mombeini T, Roushanzamin F, Jorjani M, Pourpak Z, Gaskari SA, Mehr SE, Dehpour AR: **Alteration in specific opioid-receptor labeling on peripheral blood leukocytes of bile duct-ligated rat.** *Pathophysiology* 2006, **13**:111-117.
  140. Nelson L, Vergnolle N, D'Mello C, Chapman K, Le T, Swain MG: **Endogenous opioid-mediated antinociception in cholestatic mice is peripherally, not centrally, mediated.** *J Hepatol* 2006, **44**:1141-1149.



141. Marzioni M, Baroni GS, Alpini G, Benedetti A: **Endogenous opioid peptides and chronic liver disease: from bedside to bench.** *J Hepatol* 2007, **46**:583-586.
142. Bogorad RL, Ostroukhva TY, Orlova AN, Rubtsov PM, Smirnova OV: **Long isoform of prolactin receptor predominates in rat intrahepatic bile ducts and further increases under obstructive cholestasis.** *J Endocrinol* 2006, **188**:345-354.
143. Lopez L, Begega A, Arias JL, Lorente L, Aller MA, Duran M, Arias J: **Hepatic cytochrome oxidase in rats with microsurgical cholestasis or portocaval shunt.** *Rev Esp Enferm Dig* 1999, **91**:365-373.
144. Wong-Riley MTT: **An endogenous metabolic marker for neuronal activity.** *Trends Neurosci* 1989, **12**:94-101.
145. Georgiev P, Jochum W, Heinrich S, Jang JH, Nocito A, Dahm F, Clavien PA: **Characterization of time-related changes after experimental bile duct ligation.** *Br J Surg* 2008, **95**:646-656.
146. Gehring S, Dickson EM, San Martin ME, Van Rooijen N, Papa EF, Harty MW, Tracy TF, Gregory SS: **Kupffer cells abrogate cholestatic liver injury in mice.** *Gastroenterology* 2006, **130**:810-822.
147. Li Z, Mizuno S, Nakamura T: **Antinecrotic and antipoptotic effects of hepatocyte growth factor on cholestatic hepatitis in a mouse model of bile-obstructive diseases.** *Am J Physiol Gastrointest Liver Physiol* 2007, **292**:G639-G646.
148. Wang M, Chen M, Zheng G, Dillard Iii B, Tallarico M, Ortiz Z, Holterman AX: **Transcriptional activation by growth hormone of HNF-6-regulated hepatic genes, a potential mechanism for improved liver repair during biliary injury in mice.** *Am J Physiol Gastrointest Liver Physiol* 2008, **295**(2):G357-G366.
149. Hoekstra H, Tian Y, Jocum W, Stieger B, Graf R, Porte RJ, Clavien PA: **Dearterialization of the liver causes intrahepatic cholestasis due to reduced bile transporter expression.** *Transplantation* 2008, **85**:1159-1166.
150. Semenza GL: **Targeting HIF-1 for cancer therapy.** *Nat Rev Cancer* 2003, **3**:721-732.
151. Lopez-Lazaro M: **HIF-1: hypoxia-inducible factor or dysoxia-inducible factor?** *FASEB J* 2006, **20**:828-832.
152. Brunengraber H, Roe CR: **Anaplerotic molecules: current and future.** *J Inherit Metab Dis* 2006, **29**:327-331.
153. Deberardinis RJ, Mancuso A, Daikhin E, Nissim I, Yudkoff M, Wehrli S, Thompson CB: **Beyond aerobic glycolysis: transformed cells can engage in glutamine metabolism that exceeds the requirement for protein and nucleotide synthesis.** *Proc Natl Acad Sci USA* 2007, **104**:19345-19350.
154. Marcel M, Leroy A: **Clinical, cellular and molecular aspects of cancer invasion.** *Physiol Rev* 2003, **83**:337-376.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

