

Cell autonomous and systemic factors in progeria development

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Abstract

Progeroid laminopathies are accelerated aging syndromes caused by defects in nuclear envelope proteins. Accordingly, mutations in the *LMNA* gene and functionally related genes have been described to cause HGPS (Hutchinson–Gilford progeria syndrome), MAD (mandibuloacral dysplasia) or RD (restrictive dermopathy). Functional studies with animal and cellular models of these syndromes have facilitated the identification of the molecular alterations and regulatory pathways involved in progeria development. We have recently described a novel regulatory pathway involving *miR-29* and p53 tumour suppressor which has provided valuable information on the molecular components orchestrating the response to nuclear damage stress. Furthermore, by using progeroid mice deficient in *ZMPSTE24* (zinc metalloprotease STE24 homologue) involved in lamin A maturation, we have demonstrated that, besides these abnormal cellular responses to stress, dysregulation of the somatotrophic axis is responsible for some of the alterations associated with progeria. Consistent with these observations, pharmacological restoration of the somatotroph axis in these mice delays the onset of their progeroid features, significantly extending their lifespan and supporting the importance of systemic alterations in progeria progression. Finally, we have very recently identified a novel progeroid syndrome with distinctive features from HGPS and MAD, which we have designated NGPS (Néstor–Guillermo progeria syndrome) (OMIM #614008). This disorder is caused by a mutation in *BANF1*, a gene encoding a protein with essential functions in the assembly of the nuclear envelope, further illustrating the importance of the nuclear lamina integrity for human health and providing additional support to the study of progeroid syndromes as a valuable source of information on human aging.

Introduction

The nuclear envelope is a complex structure that surrounds and protects the genome, playing essential roles in its regulation, organization and maintenance [1]. The nuclear envelope is composed of two membrane bilayers with nuclear pores that control traffic in and out the nucleus [2,3]. The nuclear face of the inner membrane is covered by the nuclear lamina, a protein network that provides scaffold for nuclear envelope proteins and chromatin [4]. In humans, three genes named *LMNA*, *LMNB1* and *LMNB2* encode nuclear lamins. Whereas the two B-type lamins are encoded by two independent genes, *LMNB1* and *LMNB2*, the *LMNA* gene encodes lamin A and lamin C proteins by alternative splicing. Mutations in A-type lamins, lamin B and several lamin-binding proteins (emerin, MAN1 and lamin B receptor) have been found mutated in different human diseases which are collectively known as laminopathies [5]. The range, diversity and tissue-specificity of laminopathy phenotypes are providing valuable clues about the cellular functions of lamins and lamin-related proteins.

Progeroid laminopathies are human syndromes of accelerated aging caused by defects in the nuclear lamina [6,7]. Among them, HGPS (Hutchinson–Gilford progeria syndrome) is the best known. Affected patients show growth impairment, lipodystrophy, dermal and bone abnormalities and cardiovascular alterations, leading to a shortened lifespan [8–10]. HGPS is caused in most cases by a *de novo* point mutation within exon 11 of the *LMNA* gene encoding lamin A (c.1824C>T; p.G608G) [11,12]. Lamin A undergoes a complex maturation process, including the addition of a farnesyl group and a proteolytic processing event carried out by the metalloprotease *ZMPSTE24* (zinc metalloprotease STE24 homologue)/FACE1 (farnesylated proteins-converting enzyme 1) [13]. The G608G mutation activates a cryptic splicing donor site, leading to the accumulation of a truncated form of prelamin A, called LA Δ 50 or progerin, which lacks a 50-residue-long fragment containing the target sequence for the final proteolytic step carried out by *ZMPSTE24*/FACE1. Consequently, this aberrant lamin A isoform remains constitutively farnesylated [14,15].

The use of cellular and murine models of progeroid laminopathies [14–19] has provided valuable information about the molecular alterations involved in progeria, such as the involvement of the p53 tumour suppressor [20], the altered biology of adult stem cells [21] or the presence of metabolic alterations [22,23]. These studies have allowed the

Key words: alternative splicing, laminopathy, microRNA (miRNA), progeria, senescence, tumour suppressor.

Abbreviations used: BAF, barrier to autointegration factor 1; FACE1, farnesylated proteins-converting enzyme 1; GH, growth hormone; HGPS, Hutchinson–Gilford progeria syndrome; IGF-1, insulin-like growth factor 1; miRNA, microRNA; NGPS, Néstor–Guillermo progeria syndrome; rIGF-1, recombinant IGF-1; *ZMPSTE24*, zinc metalloprotease STE24 homologue.

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development of the first therapeutic approaches for patients with HGPS [24–28]. Currently, two clinical trials are being carried out based on a combination of statins and amino bisphosphonates, alone or complemented with a low dose of a farnesyltransferase inhibitor (<http://clinicaltrials.gov>; NCT00731016 and NCT00916747).

Although murine models of progeroid laminopathies have been essential for understanding the pathways and alterations that drive progeria development, important questions remain to be answered, especially those related to the regulatory mechanisms that control and integrate the altered pathways, the specific contribution of cellular and systemic alterations to the progeroid phenotype, as well as the specific function of each nuclear lamina component. In this regard, three recent reports from our laboratory have shed light on these points, highlighting the importance of nuclear envelope for human health [28–30].

An *miR-29*/p53 regulatory circuit involved in aging and chronic DNA damage response

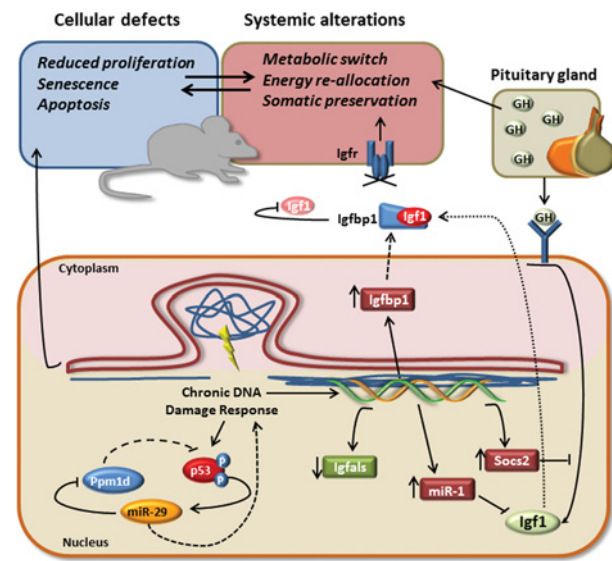
A common feature of aging is the progressive accumulation of cellular damage. Several stressors have been proposed to contribute to this situation, such as oxidative reactions, telomere attrition or the decline of DNA repair and protein turnover systems [31,32]. Progeroid syndromes associated with lamin A alterations show genomic instability as a consequence of the nuclear envelope disruption [33]. This stress triggers a series of cellular and systemic events directed to the restoration of cellular and organismal homeostasis, which are ultimately responsible for many of the alterations characteristic of these syndromes. In this regard, important changes in chromatin organization and transcriptional profiles have been described in murine models of HGPS [20,34], but little is known about the molecular components that orchestrate these changes.

Over the last decade, miRNAs (microRNAs) have emerged as a new and fundamental level of gene regulation. Each miRNA has the potential ability to repress the translation of hundreds of transcripts and their impact on a great number of cellular processes has been broadly proved. To explore the possible involvement of miRNAs in progeria development, we have performed a miRNA transcriptome analysis in the liver of *Zmpste24*-deficient mice [29]. Among the differentially expressed miRNAs, three of them (*miR-29a*, *miR-29b* and *miR-29c*), belonging to the *miR-29* family, are significantly up-regulated in tissues of *Zmpste24* progeroid-knockout mice.

Through a series of functional analysis, we have found that *miR-29* plays a pivotal role in the regulation of cell survival and proliferation through the modulation of the DNA damage response in a p53-dependent way. Thus the p53-mediated activation of the DNA damage response in *Zmpste24*-deficient cells [35] would trigger multiple effector pathways, including an increase in *miR-29* expression (Figure 1). Among the potential targets of

Figure 1. | Cellular and systemic alterations involved in progeroid laminopathies

The accumulation of farnesylated forms of prelamin A at the nuclear envelope causes severe alterations in nuclear dynamics, triggering an adaptive response aimed at preserving organism viability under compromising circumstances. At a cellular level, the p53 signalling pathway orchestrates a chronic DNA damage response that involves *miR-29* transcriptional activation. Thus *miR-29*-mediated repression of *Ppm1d* phosphatase reinforces this stress response, favouring a decrease in cellular proliferation rates accompanied by an increase in apoptosis and senescence. These processes result in the loss of tissue and organism homeostasis. In parallel, a chronic stress response cause changes in the transcriptional profiles of several somatotroph axis key regulators, such as *Igfbp1* (IGF-binding protein 1), *Socs2* (suppressor of cytokine signalling 2), *miR-1* or *Igfals* (IGF-binding protein, acid-labile subunit). These alterations dramatically reduce the levels of circulating IGF-1, which, together with the increased production of GH at the pituitary gland, favours a systemic metabolic switch towards somatic maintenance at the expense of somatic growth.



miR-29, Ppm1d/Wip1 phosphatase has been proposed as the key mediator for this effect. Ppm1d is a phosphatase that acts as a negative regulator of DNA damage response by dephosphorylating important components of this process such as p53, Chk1 (checkpoint kinase 1), Chk2 (checkpoint kinase 2), p38, γ -H2AX (phosphorylated histone H2AX) or ATM (ataxia telangiectasia mutated) [36,37]. Thus a decrease in Ppm1d levels mediated by *miR-29* would contribute to the activation of the DNA damage response. In agreement with these results, *miR-29* has been described as a tumour-suppressor miRNA in several human cancers. This tumour-suppressive function could be consistent with a pro-aging role for this miRNA, since a growing number of tumour-suppressor genes have been reported to be aging promoters, which could be illustrative examples of the antagonistic pleiotropy phenomenon [38].

Somatotroph suppression in *Zmpste24*-deficient mice

Although the dynamics of aging are far from being completely understood, our knowledge of the systemic factors involved in this process has considerably increased in recent years [39–41]. Somatotroph signalling has been identified as a major regulator of longevity from nematodes to humans [42]. Paradoxically, studies in different organisms have shown that the reduction of this signalling is a common feature of both long-lived model organisms and different progeroid mice [43,44].

In this sense, *Zmpste24*-deficient mice show a profound dysregulation of GH (growth hormone)/IGF-1 (insulin-like growth factor 1) balance, with a progressive reduction of blood IGF-1 levels, accompanied by a progressive increase in GH levels and a marked transcriptional alterations in key genes for somatotroph signalling [28] (Figure 1). Thus somatotroph alterations would be responsible for important features of progeroid phenotype such as reduced growth rate and body size. In this case, the observed alterations in somatotrophic axis seem to constitute a detrimental phenomenon, rather than a successful adaptive strategy, as demonstrated by the fact that the treatment of *Zmpste24*-deficient mice with rIGF-1 (recombinant IGF-1) is able to ameliorate some of the progeroid features of these mice. rIGF-1-treated mice showed improved body weight, increased amounts of subcutaneous fat deposits, reduced degree of lordokyphosis and alopecia, and significantly extended longevity. Accordingly, rIGF-1 treatment could be a therapeutic approach to slow down disease progression in children with progeria [45].

Interestingly, many pathological features of GH resistance, also known as Laron syndrome, are characteristic of progeroid mice. Both *Zmpste24*^{-/-} mice and patients with this syndrome show reduced muscle development, strength and endurance, as well as decreased bone mineral density, alopecia, skin atrophy and hypoglycaemia [14,22,46]. Some of these alterations could be consequence of an adaptive stress response aimed at preserving organism viability under compromising circumstances by reallocating resources from growth to somatic preservation. In *Zmpste24*-knockout mice, this systemic response could represent an attempt to reduce replication defects, chromosomal instability, nuclear envelope abnormalities and, finally, the risk of developing cancer by decreasing metabolic activity. This hypothesis is supported by the fact that patients with Laron syndrome or other somatotroph-related pathologies such as GH receptor deficiency exhibit a notable reduction in the incidence of malignancies [47,48].

NGPS, a new hereditary progeroid syndrome caused by *BANF1* mutation

The recent availability of high-throughput sequencing technologies has made it possible to address personal genome projects that could uncover the precise causes of human genetic diseases [49,50]. Thus exome sequencing of two unrelated

patients that exhibit a progeroid syndrome without mutations in *LMNA* or *ZMPSTE24* has allowed the identification of a homozygous mutation in *BANF1* (c.34G>C; p.A12T), encoding BAF (barrier to autointegration factor 1), as the molecular abnormality responsible for this syndrome [30].

Affected patients of this disease, called NGPS (Néstor-Guillermo progeria syndrome) (OMIM #614008), partially phenocopy HGPS and MAD (mandibuloacral dysplasia), but also exhibit distinctive features, including the absence of cardiovascular alterations and metabolic anomalies, a very severe osteolysis and a relatively long lifespan of affected individuals [51]. NGPS can be considered as a chronic progeria because of its early onset but slow clinical course, which leads to a relatively long survival of patients.

BAF is a small protein (89 amino acids) highly conserved among metazoans [52,53]. BAF binds directly to double-stranded DNA, chromatin, nuclear lamina proteins (including lamin A and emerin), histones and transcription factors, these being required for higher-order organization of chromatin, nuclear envelope assembly, retrovirus infectivity and transcription of specific genes [54].

The A12T mutation in NGPS patients affects a highly conserved residue located at the surface of the protein, decreasing its stability. Skin fibroblasts from these patients show very low protein levels of BAF compared with control fibroblasts and exhibit profound nuclear abnormalities, including blebs and other aberrations associated with progeroid laminopathies. In fact, BAF reduction is correlated with mislocalization of emerin, which shows a predominant cytoplasmic localization in mutant cells. Taken together, these findings demonstrate the relevance of BAF in nuclear envelope dynamics, providing new insights about the relationship of nuclear envelope to aging.

Conclusions and perspectives

Over the last few years, the generation of experimental murine models of progeroid laminopathies has been crucial for a deeper understanding of the molecular basis of these diseases. This is the case for HGPS, where a fast progress has been made in the last 8 years since the identification of *LMNA* mutations to the first clinical trial in HGPS patients. However, new murine models that fully recapitulate all the disease phenotypes of HGPS [55] are necessary to boost the development of *in vivo* approaches directed to the correction of *LMNA* aberrant splicing [25]. Besides, several questions remain to be answered concerning important aspects such as the relative contribution to the progeroid phenotype of cell-autonomous compared with systemic alterations or the involvement of nuclear envelope dynamics during normal aging [56–58].

Funding

This work was supported by grants from Ministerio de Ciencia e Innovación-Spain, PCTI (Plan de Ciencia, Tecnología e Innovación del

Principado de Asturias)–FICYT (Fundación para el Fomento en Asturias de la Investigación Científica Aplicada y la Tecnología) Asturias, and the European Union Framework Programme 7 (MicroEnviMet). The Instituto Universitario de Oncología is supported by Obra Social Cajastur and Acción Transversal del Cáncer-RTICC (Red Temática de Investigación Cooperativa en Cáncer). C.L.-O. is an Investigator of the Botin Foundation.

References

- Dechat, T., Pflieger, K., Sengupta, K., Shimi, T., Shumaker, D.K., Solimando, L. and Goldman, R.D. (2008) Nuclear lamins: major factors in the structural organization and function of the nucleus and chromatin. *Genes Dev.* **22**, 832–853
- Mekhail, K. and Moazed, D. (2010) The nuclear envelope in genome organization, expression and stability. *Nat. Rev. Mol. Cell Biol.* **11**, 317–328
- Fiserova, J. and Goldberg, M.W. (2010) Nucleocytoplasmic transport in yeast: a few roles for many actors. *Biochem. Soc. Trans.* **38**, 273–277
- Gruenbaum, Y., Margalit, A., Goldman, R.D., Shumaker, D.K. and Wilson, K.L. (2005) The nuclear lamina comes of age. *Nat. Rev. Mol. Cell Biol.* **6**, 21–31
- Worman, H.J., Ostlund, C. and Wang, Y. (2010) Diseases of the nuclear envelope. *Cold Spring Harbor Perspect. Biol.* **2**, a000760
- Ramirez, C.L., Cadiñanos, J., Varela, I., Freije, J.M. and López-Otin, C. (2007) Human progeroid syndromes, aging and cancer: new genetic and epigenetic insights into old questions. *Cell. Mol. Life Sci.* **64**, 155–170
- Burtner, C.R. and Kennedy, B.K. (2010) Progeria syndromes and ageing: what is the connection? *Nat. Rev. Mol. Cell Biol.* **11**, 567–578
- Hennekam, R.C. (2006) Hutchinson–Gilford progeria syndrome: review of the phenotype. *Am. J. Med. Genet. A* **140**, 2603–2624
- Merideth, M.A., Gordon, L.B., Clauss, S., Sachdev, V., Smith, A.C., Perry, M.B., Brewer, C.C., Zalewski, C., Kim, H.J., Solomon, B. et al. (2008) Phenotype and course of Hutchinson–Gilford progeria syndrome. *N. Engl. J. Med.* **358**, 592–604
- Pereira, S., Bourgeois, P., Navarro, C., Esteves-Vieira, V., Cau, P., De Sandre-Giovannoli, A. and Lévy, N. (2008) HGPS and related premature aging disorders: from genomic identification to the first therapeutic approaches. *Mech. Ageing Dev.* **129**, 449–459
- De Sandre-Giovannoli, A., Bernard, R., Cau, P., Navarro, C., Amiel, J., Boccaccio, I., Lyonnet, S., Stewart, C.L., Munnich, A., Le Merrer, M. and Lévy, N. (2003) Lamin A truncation in Hutchinson–Gilford progeria. *Science* **300**, 2055
- Eriksson, M., Brown, W.T., Gordon, L.B., Glynn, M.W., Singer, J., Scott, L., Erdos, M.R., Robbins, C.M., Moses, T.Y., Berglund, P. et al. (2003) Recurrent *de novo* point mutations in lamin A cause Hutchinson–Gilford progeria syndrome. *Nature* **423**, 293–298
- Freije, J.M., Blay, P., Pendás, A.M., Cadiñanos, J., Crespo, P. and López-Otin, C. (1999) Identification and chromosomal location of two human genes encoding enzymes potentially involved in proteolytic maturation of farnesylated proteins. *Genomics* **58**, 270–280
- Pendás, A.M., Zhou, Z., Cadiñanos, J., Freije, J.M., Wang, J., Hultenby, K., Astudillo, A., Wernerson, A., Rodríguez, F., Tryggvason, K. and López-Otin, C. (2002) Defective prelamin A processing and muscular and adipocyte alterations in Zmpste24 metalloproteinase-deficient mice. *Nat. Genet.* **31**, 94–99
- Bergo, M.O., Gavino, B., Ross, J., Schmidt, W.K., Hong, C., Kendall, L.V., Mohr, A., Meta, M., Genant, H., Jiang, Y. et al. (2002) Zmpste24 deficiency in mice causes spontaneous bone fractures, muscle weakness, and a prelamin A processing defect. *Proc. Natl. Acad. Sci. U.S.A.* **99**, 13049–13054
- Yang, S.H., Bergo, M.O., Toth, J.I., Qiao, X., Hu, Y., Sandoval, S., Meta, M., Bendale, P., Gelb, M.H., Young, S.G. and Fong, L.G. (2005) Blocking protein farnesyltransferase improves nuclear blebbing in mouse fibroblasts with a targeted Hutchinson–Gilford progeria syndrome mutation. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 10291–10296
- Varga, R., Eriksson, M., Erdos, M.R., Olive, M., Harten, I., Kolodgie, F., Capell, B.C., Cheng, J., Faddah, D., Perkins, S. et al. (2006) Progressive vascular smooth muscle cell defects in a mouse model of Hutchinson–Gilford progeria syndrome. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 3250–3255
- Wang, Y., Panteleyev, A.A., Owens, D.M., Djabali, K., Stewart, C.L. and Worman, H.J. (2008) Epidermal expression of the truncated prelamin A causing Hutchinson–Gilford progeria syndrome: effects on keratinocytes, hair and skin. *Hum. Mol. Genet.* **17**, 2357–2369
- Sagelius, H., Rosengarten, Y., Hanif, M., Erdos, M.R., Rozell, B., Collins, F.S. and Eriksson, M. (2008) Targeted transgenic expression of the mutation causing Hutchinson–Gilford progeria syndrome leads to proliferative and degenerative epidermal disease. *J. Cell Sci.* **121**, 969–978
- Varela, I., Cadiñanos, J., Pendás, A.M., Gutiérrez-Fernández, A., Folgueras, A.R., Sánchez, L.M., Zhou, Z., Rodríguez, F.J., Stewart, C.L., Vega, J.A. et al. (2005) Accelerated ageing in mice deficient in Zmpste24 protease is linked to p53 signalling activation. *Nature* **437**, 564–568
- Espada, J., Varela, I., Flores, I., Ugalde, A.P., Cadiñanos, J., Pendás, A.M., Stewart, C.L., Tryggvason, K., Blasco, M.A., Freije, J.M. and López-Otin, C. (2008) Nuclear envelope defects cause stem cell dysfunction in premature-aging mice. *J. Cell Biol.* **181**, 27–35
- Marino, G., Ugalde, A.P., Salvador-Montoliu, N., Varela, I., Quirós, P.M., Cadiñanos, J., van der Pluijm, I., Freije, J.M. and López-Otin, C. (2008) Premature aging in mice activates a systemic metabolic response involving autophagy induction. *Hum. Mol. Genet.* **17**, 2196–2211
- Worman, H.J., Fong, L.G., Muchir, A. and Young, S.G. (2009) Laminopathies and the long strange trip from basic cell biology to therapy. *J. Clin. Invest.* **119**, 1825–1836
- Yang, S.H., Meta, M., Qiao, X., Frost, D., Bauch, J., Coffinier, C., Majumdar, S., Bergo, M.O., Young, S.G. and Fong, L.G. (2006) A farnesyltransferase inhibitor improves disease phenotypes in mice with a Hutchinson–Gilford progeria syndrome mutation. *J. Clin. Invest.* **116**, 2115–2121
- Scaffidi, P. and Misteli, T. (2005) Reversal of the cellular phenotype in the premature aging disease Hutchinson–Gilford progeria syndrome. *Nat. Med.* **11**, 440–445
- Varela, I., Pereira, S., Ugalde, A.P., Navarro, C.L., Suárez, M.F., Cau, P., Cadiñanos, J., Osorio, F.G., Foray, N., Cobo, J. et al. (2008) Combined treatment with statins and aminobisphosphonates extends longevity in a mouse model of human premature aging. *Nat. Med.* **14**, 767–772
- Cao, K., Graziotto, J.J., Blair, C.D., Mazzulli, J.R., Erdos, M.R., Krainc, D. and Collins, F.S. (2011) Rapamycin reverses cellular phenotypes and enhances mutant protein clearance in Hutchinson–Gilford progeria syndrome cells. *Sci. Transl. Med.* **3**, 89ra58
- Mariño, G., Ugalde, A.P., Fernández, A.F., Osorio, F.G., Fueyo, A., Freije, J.M. and López-Otin, C. (2010) Insulin-like growth factor 1 treatment extends longevity in a mouse model of human premature aging by restoring somatotroph axis function. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 16268–16273
- Ugalde, A.P., Ramsay, A.J., de la Rosa, J., Varela, I., Marino, G., Cadiñanos, J., Lu, J., Freije, J.M. and López-Otin, C. (2011) Aging and chronic DNA damage response activate a regulatory pathway involving miR-29 and p53. *EMBO J.* **30**, 2219–2232
- Puente, X.S., Quesada, V., Osorio, F.G., Cabanillas, R., Cadiñanos, J., Fraile, J.M., Ordóñez, G.R., Puente, D.A., Gutiérrez-Fernández, A., Fanjul-Fernández, M. et al. (2011) Exome sequencing and functional analysis identifies *BANF1* mutation as the cause of a hereditary progeroid syndrome. *Am. J. Hum. Genet.* **88**, 650–656
- Kirkwood, T.B. (2005) Understanding the odd science of aging. *Cell* **120**, 437–447
- Kourtis, N. and Tavernarakis, N. (2011) Cellular stress response pathways and ageing: intricate molecular relationships. *EMBO J.* **30**, 2520–2531
- Liu, B., Wang, J., Chan, K.M., Tjia, W.M., Deng, W., Guan, X., Huang, J.D., Li, K.M., Chau, P.Y., Chen, D.J. et al. (2005) Genomic instability in laminopathy-based premature aging. *Nat. Med.* **11**, 780–785
- Osorio, F.G., Varela, I., Lara, E., Puente, X.S., Espada, J., Santoro, R., Freije, J.M., Fraga, M.F. and López-Otin, C. (2010) Nuclear envelope alterations generate an aging-like epigenetic pattern in mice deficient in Zmpste24 metalloproteinase. *Ageing Cell* **9**, 947–957
- Cadiñanos, J., Varela, I., López-Otin, C. and Freije, J.M. (2005) From immature lamin to premature aging: molecular pathways and therapeutic opportunities. *Cell Cycle* **4**, 1732–1735
- Lu, X., Nguyen, T.A., Moon, S.H., Darlington, Y., Sommer, M. and Donehower, L.A. (2008) The type 2C phosphatase Wip1: an oncogenic regulator of tumour suppressor and DNA damage response pathways. *Cancer Metastasis Rev.* **27**, 123–135
- Cha, H., Lowe, J.M., Li, H., Lee, J.S., Belova, G.I., Bulavin, D.V. and Fornace, Jr. A.J. (2010) Wip1 directly dephosphorylates γ -H2AX and attenuates the DNA damage response. *Cancer Res.* **70**, 4112–4122
- Campisi, J. (2005) Aging, tumour suppression and cancer: high wire-act! *Mech. Ageing Dev.* **126**, 51–58

- 39 Houtkooper, R.H., Williams, R.W. and Auwerx, J. (2010) Metabolic networks of longevity. *Cell* **142**, 9–14
- 40 Panowski, S.H. and Dillin, A. (2009) Signals of youth: endocrine regulation of aging in *Caenorhabditis elegans*. *Trends Endocrinol. Metab.* **20**, 259–264
- 41 Haigis, M.C. and Yankner, B.A. (2010) The aging stress response. *Mol. Cell* **40**, 333–344
- 42 Russell, S.J. and Kahn, C.R. (2007) Endocrine regulation of ageing. *Nat. Rev. Mol. Cell Biol.* **8**, 681–691
- 43 Niedernhofer, L.J., Garinis, G.A., Raams, A., Lalai, A.S., Robinson, A.R., Appeldoorn, E., Odijk, H., Oostendorp, R., Ahmad, A., van Leeuwen, W. et al. (2006) A new progeroid syndrome reveals that genotoxic stress suppresses the somatotroph axis. *Nature* **444**, 1038–1043
- 44 Hoeijmakers, J.H. (2009) DNA damage, aging, and cancer. *N. Engl. J. Med.* **361**, 1475–1485
- 45 Ugalde, A.P., Marino, G. and López-Otín, C. (2010) Rejuvenating somatotrophic signaling: a therapeutical opportunity for premature aging? *Aging* **2**, 1017–1022
- 46 Laron, Z. (2004) Laron syndrome (primary growth hormone resistance or insensitivity): the personal experience 1958–2003. *J. Clin. Endocrinol. Metab.* **89**, 1031–1044
- 47 Steuerman, R., Shevah, O. and Laron, Z. (2011) Congenital IGF1 deficiency tends to confer protection against post-natal development of malignancies. *Eur. J. Endocrinol.* **164**, 485–489
- 48 Guevara-Aguirre, J., Balasubramanian, P., Guevara-Aguirre, M., Wei, M., Madia, F., Cheng, C.W., Hwang, D., Martin-Montalvo, A., Saavedra, J., Ingles, S. et al. (2011) Growth hormone receptor deficiency is associated with a major reduction in pro-aging signaling, cancer, and diabetes in humans. *Sci. Transl. Med.* **3**, 70ra13
- 49 Roach, J.C., Glusman, G., Smit, A.F., Huff, C.D., Hubley, R., Shannon, P.T., Rowen, L., Pant, K.P., Goodman, N., Bamshad, M. et al. (2010) Analysis of genetic inheritance in a family quartet by whole-genome sequencing. *Science* **328**, 636–639
- 50 Lupski, J.R., Reid, J.G., Gonzaga-Jauregui, C., Rio Deiros, D., Chen, D.C., Nazareth, L., Bainbridge, M., Dinh, H., Jing, C., Wheeler, D.A. et al. (2010) Whole-genome sequencing in a patient with Charcot-Marie-Tooth neuropathy. *N. Engl. J. Med.* **362**, 1181–1191
- 51 Cabanillas, R., Cadiñanos, J., Villameytide, J.A.F., Perez, M., Longo, J., Richard, J.M., Alvarez, R., Duran, N.S., Illan, R., Gonzalez, D.J. et al. (2011) Néstor-Guillermo progeria syndrome: a novel premature aging condition with early onset and chronic development caused by *BANF1* mutations. *Am. J. Med. Genet. A* **155**, 2617–2625
- 52 Segura-Totten, M. and Wilson, K.L. (2004) BAF: roles in chromatin, nuclear structure and retrovirus integration. *Trends Cell Biol.* **14**, 261–266
- 53 Margalit, A., Brachner, A., Gotzmann, J., Foisner, R. and Gruenbaum, Y. (2007) Barrier-to-autointegration factor: a BAFfling little protein. *Trends Cell Biol.* **17**, 202–208
- 54 Segura-Totten, M., Kowalski, A.K., Craigie, R. and Wilson, K.L. (2002) Barrier-to-autointegration factor: major roles in chromatin decondensation and nuclear assembly. *J. Cell Biol.* **158**, 475–485
- 55 Osorio, F.G., Obaya, A.J., López-Otín, C. and Freije, J.M. (2009) Accelerated ageing: from mechanism to therapy through animal models. *Transgenic Res.* **18**, 7–15
- 56 Scaffidi, P. and Misteli, T. (2006) Lamin A-dependent nuclear defects in human aging. *Science* **312**, 1059–1063
- 57 Ragnauth, C.D., Warren, D.T., Liu, Y., McNair, R., Tajsic, T., Figg, N., Shroff, R., Skepper, J. and Shanahan, C.M. (2010) Prelamin A acts to accelerate smooth muscle cell senescence and is a novel biomarker of human vascular aging. *Circulation* **121**, 2200–2210
- 58 Cao, K., Blair, C.D., Faddah, D.A., Kieckhafer, J.E., Olive, M., Erdos, M.R., Nabel, E.G. and Collins, F.S. (2011) Progerin and telomere dysfunction collaborate to trigger cellular senescence in normal human fibroblasts. *J. Clin. Invest.* **121**, 2833–2844

Received 27 July 2011
doi:10.1042/BST20110677