www.nature.com/leu

ORIGINAL ARTICLE Age, $JAK2^{V617F}$ and SF3B1 mutations are the main predicting factors for survival in refractory anaemia with ring sideroblasts and marked thrombocytosis

J Broséus¹, T Alpermann², M Wulfert³, L Florensa Brichs^{4,10}, S Jeromin², E Lippert⁵, M Rozman^{6,10}, F Lifermann⁷, V Grossmann², T Haferlach², U Germing³, E Luño^{8,10}, F Girodon⁹ and S Schnittger² for the MPN and MPNr-EuroNet (COST Action BM0902)

Refractory anaemia with ring sideroblasts (RARS) and marked thrombocytosis (RARS-T) is a provisional entity in the World Health Organisation 2008 classification and has previously been shown to have a high proportion of $JAK2^{V617F}$ (*Janus Kinase 2*) and *SF3B1* (*Splicing Factor 3B subunit 1*) mutations. The purpose of the present study was to analyse the frequency of *SF3B1* mutations in a large cohort of 111 patients with RARS-T and 33 patients with RARS and to explore the prognostic impact of *SF3B1* mutational status on RARS-T. The frequency of *SF3B1* mutations in RARS-T (96/111, 86.5%) and RARS (28/33, 84.8%) was similar. In RARS-T, median survival was better in *SF3B1*-mutated patients than in *SF3B1*-non-mutated patients (6.9 and 3.3 years, respectively, P = 0.003). RARS can be differentiated from RARS-T by the frequency of $JAK2^{V617F}$ (0% vs 48.6%). In RARS-T patients, *SF3B1* (P = 0.021) and *JAK2* mutations (P = 0.016) were independent factors for a better prognosis. Altogether, our results confirm that RARS-T is an independent entity that should be recognised by the next World Health Organisation classification. The assessment of *SF3B1* mutations is of prognostic interest in RARS-T patients. Younger age, *JAK2*^{V617F} and *SF3B1* mutations are the main predicting factors for survival in RARS-T.

Leukemia (2013) 27, 1826-1831; doi:10.1038/leu.2013.120

Keywords: refractory anaemia with ring sideroblasts and marked thrombocytosis; SF3B1; prognostic impact; survival

INTRODUCTION

Refractory anaemia with ring sideroblasts (RARS) and marked thrombocytosis (RARS-T) has been proposed in the World Health Organisation 2001 classification of tumours of haematopoietic and lymphoid tissues and retained as a provisional entity in the classification of 2008.¹ RARS-T is classified in the myelodysplastic/myeloproliferative (MDS/MPN) disorders group, because it presents with the dysplastic features of RARS² and the myeloproliferative features of essential thrombocythemia (ET).³ RARS-T is characterised by a high rate of $JAK2^{V617F}$ (Janus Kinase 2) mutations^{4–15} and/or the presence of the mutation $MPL^{W515L/R}$ (*MyeloProliferative Leukemia*).^{16,17} The classification of RARS-T as an entity that is independent from RARS or ET is currently a matter of debate. Several specialists favour the hypothesis that RARS-T is a form of ET with $\geq 15\%$ of ring sideroblasts in the bone marrow¹⁸ while others think that RARS-T develops from RARS with secondary thrombocytosis accompanying the acquisition of the *JAK2^{V617F}* mutation.¹⁹

Recent publication from our group in a European retrospective multicentre collaborative study demonstrated that RARS-T was independent from RARS and ET from a clinical and biological as well as prognostic point of view.²⁰

Our results have recently been strengthened by the discovery of the association between myelodysplastic syndromes and mutations involving components of the RNA splicing machinery, including U2AF35 (U2 small nuclear RNA Auxiliary Factor 35), ZRSR2 (Zinc finger CCCH type, RNA-binding motif and Serine/aRginine rich 2), SRSF2 (Serine/aRginine-rich Splicing Factor 2) and SF3B1 (Splicing Factor 3B subunit 1). SF3B1 mutations (SF3B1^{mut}) are found in about 20% of total MDS and correlate strongly with the presence of \geq 15% of ring sideroblasts (MDS-RS; 64–82.6% in RARS, 57–76% in refractory cytopenia with multilineage dysplasia and ring sideroblasts (RCMD-RS) and 66.7–72% in RARS-T).^{21–27} On the other hand, mutations of SF3B1 are found at a lower frequency in MDS with <15% ring sideroblasts, which confirms the specificity of SF3B1^{mut} in MDS-RS. SF3B1 mutations are rare in myeloproliferative neoplasms and particularly in ET (0-3%).22,28 The high frequency of SF3B1 mutations suggests that these mutations have a pathophysiological role in these diseases, probably through perturbations of RNA splicing. The link between SF3B1-mutated status and ring sideroblasts has been confirmed in a recent experimental study on murine models.²⁹ About one guarter of MDS-RS are SF3B1^{wt} and somatic mutations of SRSF2 or ZRSR2 have been described in about 7% of MDS-RS,²¹

E-mail: francois.girodon@chu-dijon.fr or susanne.schnittger@mll.com

¹⁰On behalf of the Spanish Group of Hematological Cytology (GECH).

Received 20 March 2013; accepted 10 April 2013; accepted article preview online 18 April 2013; advance online publication, 14 May 2013

¹Haematology Laboratory, University Hospital, Nancy, France; ²MLL Munich Leukemia Laboratory, Munich, Germany; ³Department of Haematology, Oncology and Clinical Immunology, Heinrich-Heine-Universität, Düsseldorf, Germany; ⁴Laboratorio Citologia Hematològica, Servicio Patologia, Hospital del Mar, Barcelona, Spain; ⁵Haematology Laboratory, University Hospital, Bordeaux, France; ⁶Unidad de Hematopatología, Departamento de Patología, Hospital Clínic, IDIBAPS, Barcelona, Spain; ⁷Department of Internal Medicine, Hospital of Dax, Dax, France; ⁸Servicio de Hematología, Servicio de Salud del Principado de Asturias, Oviedo, Spain and ⁹Haematology Laboratory, University Hospital, Dijon, France. Correspondence: Professor F Girodon, Haematology Laboratory, University Hospital, Plateau technique de biologie, 2 rue Angélique Ducoudray, Dijon, cedex 21079, France or Dr S Schnittger, MLL Munich Leukemia Laboratory, Munich, Germany.

which suggests that other mutant genes may have a role in the appearance of ring sideroblasts. Furthermore, a recent study showed that RARS-T presented with a particular genetic pattern with a high frequency of $JAK2^{V617F}$ and SF3B1 mutations, confirming the classification of RARS-T in the category of myelodysplastic/myeloproliferative neoplasms.³⁰

Finally, as precedent studies have been performed on little RARS-T cohorts, the prognostic impact of *SF3B1*^{mut} status remains controversial,^{22,24,25,27,31} and there is a need for a study on a larger cohort. Our purpose was to analyse the frequency of *SF3B1* mutations in a large cohort of 111 RARS-T and to explore the prognostic impact of *SF3B1* mutations in this disorder.

PATIENTS AND METHODS

Patient selection

According to the World Health Organisation 2008 classification, patients were diagnosed with RARS-T if they fulfilled the following criteria: (i) anaemia (haemoglobin level <125 g/l for females and <135 g/l for males) with erythroid dysplasia and \geq 15% ring sideroblasts; (ii) thrombocytosis of \geq 450 × 10⁹ platelets/l; (iii) <5% blast cells in the bone marrow; (iv) the presence of large atypical megakaryocytes similar to those observed in *BCR-ABL1*-negative myeloproliferative neoplasms; (v) no secondary cause of ring sideroblasts; and (vi) no karyotype abnormalities, such as del(5q), t(3;3)(q21;q26) or inv(3)(q21q26).¹

To obtain a comprehensive data set of this rare entity, samples from seven European centres in three European countries were collected. The total cohort comprised 111 cases of RARS-T and 33 cases of RARS.

Data record

For each patient, demographic (gender, age at diagnosis, date of death or last follow-up) and biological data (blood cell count, bone marrow exploration, ring sideroblasts, karyotype, molecular explorations) were collected.

The *SF3B1* mutations were analysed with a sensitive next-generation amplicon deep-sequencing assay (454 Life Sciences, Branford, CT, USA) with a median coverage of 500 reads. This approach was able to detect mutations with a sensitivity < 1%.

The JAK2^{V617F} mutation was analysed by allele-specific real-time PCR to estimate allele burden according to methods published by Lippert *et al.*³² and Schnittger *et al.*³³ with a sensitivity of 1%. JAK2^{exon12} analysis was performed according to the method of Schnittger *et al.*³⁴ and the MPL^{W515} mutations were analysed by high-resolution melting curve analyses followed by Sanger sequencing if positive, as previously published by Schnittger *et al.*³⁵

Statistical analyses

Standardised overall survival was estimated by the Kaplan–Meier method and based on the time from diagnosis to death or last contact. Survival curves for the different groups were compared using the log rank test. Multivariate analysis was performed using Cox's proportional hazards model.

Approval for the study was obtained from the ethics committee of each institution, and the procedures were carried out in accordance with the Helsinki Declaration of 1975, as revised in 2000.

RESULTS

Demographic and biological features

A total of 144 cases (111 RARS-T and 33 RARS including 72 males and 72 females) were recorded in the study. Median age at diagnosis was 73.9 years (range: 44.4–96.1 years). The median follow-up was 37.5 and 55.2 months for the RARS-T and RARS cohort, respectively (Table 1). Survival data were available in 130 (97 RARS-T and 33 RARS) of the 144 patients.

Frequencies and characterisation of mutations

A karyotype was available in 128 cases. One hundred and ten (85.9%) patients carried a normal karyotype, whereas 18 (14.1%) patients showed aberrant karyotypes, which was equally

Table 1. Demographic and biological characteristics of RARS-T and RARS patients				
		RARS-T patients	RARS patients	

	nono i patiento	nuno patiento
n Male (%)	111 46.8	33 60.6
Age at diagnosis (years) Median Range 20–50 years, n (%) 50–70 years, n (%) >70 years, n (%) Available survival data (n) Median follow-up (years)	74.3 44.4–92.1 4 (3.6) 36 (32.5) 71 (63.9) 97 3.1	71.1 48.4–96.1 2 (6.1) 9 (27.2) 22 (66.7) 33 4.6
WBC (× 10 ⁹ /l) Median Range	7.6 2.1–60.0	5.2 1.6–17.3
Hb (g/l) Median Range	96.5 51.0–131.0	91.0 69.0–128.0
Platelets (× 10 ⁹ /l) Median Range 450–600, n (%) >600, n (%)	646 452–1500 54 (48.6) 57 (51.4)	314 61-444
Ring sideroblasts (%) Median Range	52 16–97	40 19–85
SF3B1 mutations (%) Tested (n) Mutated (n) p.Lys700Glu p.Lys666Glu/Thr/Asp/Asn p.His662Asp/Gln p.Glu622Asp p.Arg625Gys/Leu/Gly p.Thr663Pro p.Met784_Lys785delinslle p.Asp781Gly Two different mutations	86.5 111 96 51 16 11 7 5 2 1 0 3	84.8 33 28 16 2 2 2 3 2 0 1 0
JAK2 ^{V617F} mutations (%) Tested (<i>n</i>)	48.6 111	0 33
MPL mutations (%) Tested (n)	1 102	0 27
IPSS 0 0.5 1	104 5 2	29 3 1

Abbreviations: Hb, haemoglobin; IPSS, International Prognostic Scoring System; *JAK2*, Janus Kinase 2; *MPL*, MyeloProliferative Leukaemia; RARS, refractory anaemia with ring sideroblasts; RARS-T, refractory anaemia with ring sideroblasts and marked thrombocytosis; *SF3B1*, Splicing Factor 3B subunit 1; WBC, white blood cells.

distributed between RARS-T and RARS patients. Even if the IPSS (International Prognostic Scoring System) score can only be applied to MDS *de novo*, we calculated it to check if we had a homogeneous group of patients. Most of the patients of the total cohort (133 out of 144) had an IPSS score of 0.

A SF3B1^{mut} was noted in 124 out of the 144 patients (86.1%). A total of 127 SF3B1 mutations were detected in these 124 patients (28 RARS and 96 RARS-T). Three RARS-T cases carried two Impact of SF3B1 mutations on the prognosis of RARS-T J Broséus *et al*

1828

different mutations. With the exception of one p.Arg549Cys in exon 12 and two in exon 16, all mutations were located in exons 14 and 15. All but one del/ins mutations (p.Met784_Lys785del/inslle) were missense mutations. In detail, the most frequent mutation was p.Lys700Glu (68/127 53.5%), followed by p.Lys666-Glu/Thr/Asp/Asn mutations (n = 18, 14.2%), p.His662Asp/Gln (n = 13, 10.2%), p.Arg625Cys/Leu (n = 10, 7.9%), p.Glu622Asp (n = 9, 7.1%) and p.Thr663Pro (n = 4, 3.1%). Five further mutations were detected in single cases only. Frequencies and positions of mutation/wildtype load was 40% (range: 15–78%). Small subclones with SF3B1^{mut} were not detected.

Frequency of mutations in RARS and RARS-T

The frequency of *SF3B1* mutations in RARS-T (96/111, 86.5%) was similar to that in RARS (28/33, 84.8%). By contrast, both entities differed by the presence of the *JAK2*^{V617F} mutation, which was detected in 54/144 (37.5%) in the total cohort but in 54/111 (48.6%) in RARS-T compared with none (0/33) in RARS (*P*<0.001). Among the RARS-T *SF3B1*^{mut}, 46/96 (47.9%) harboured a *JAK2*^{V617F}

mutation. $JAK2^{V617F}$ allele burden was very heterogeneous with a median of 49% (range: 1–100%). No $JAK2^{exon12}$ mutation (111 tested) was observed, whereas only one case with the MPL^{W515L} mutation was noted in a RARS-T (102 tested; Table 1 and Figure 2).

Biological association

The presence of *SF3B1*^{mut} was analysed with respect to age, sex, white blood cell count, haemoglobin levels, platelet counts, blast counts, percentage of ring sideroblasts, karyotype and *JAK2*^{V617F} allele burden. In RARS-T, *SF3B1* mutations were more frequent in females (56/59, 94.9%) than in males (40/52, 76.9%) (*P*=0.010), and mean ring sideroblast counts were higher in *SF3B1*^{mut} than in *SF3B1*^{wt} (55% vs 38%) (*P*=0.007). No further correlations were detected for these parameters.

Impact of mutations on outcome

The difference in survival between RARS-T and RARS was not statistically significant (median survival 10.7 vs 6.2 years, respectively, P > 0.05). On the other hand, in the total cohort, patients with *SF3B1*^{mut} had longer survival than those with



Figure 1. Positions and characterisation of mutations in the HEAT domains 1–6 (exons 12–16) of the *SF3B1* gene. Missense mutations are indicated in green and the rare ins/del mutation in yellow. HEAT, Huntingtin, elongation factor 3, protein phosphatase 2A, Tor1; PPP1R8, protein phosphatase 1, regulatory subunit 8; aa, amino acid.



Figure 2. Distribution of SF3B1, JAK2^{V617F} and MPL^{W515L} mutations in RARS-T and RARS.

Impact of *SF3B1* mutations on the prognosis of RARS-T J Broséus *et al*

1829



Figure 3. Kaplan–Meier analysis for survival since diagnosis according to *SF3B1* and *JAK2*^{V617F} mutation status. *SF3B1* in (**a**) total cohort, (**b**) RARS-T, (**c**) RARS and (**d**) *JAK2*^{V617F} in RARS-T.

SF3B1^{wt} (8.0 vs 1.8 years, respectively, P < 0.001; Figure 3a). When restricted to RARS-T (85 *SF3B1*^{mut} and 12 *SF3B1*^{wt}), median overall survival was 6.9 years in *SF3B1*^{mut} vs 3.3 years in *SF3B1*^{wt} (P = 0.003; Figure 3b). In RARS, survival was 10.7 years in the *SF3B1*^{mut} (n = 28) and 1.8 years in *SF3B1*^{wt} patients (n = 5; P = 0.001; Figure 3c). In RARS-T patients, the survival rates at 2, 4 and 6 years within the *JAK2*^{v617F} sub-cohort were 94.9, 84.3 and 67.4%, respectively, while within the *JAK2*^{v617F} (n = 50) was then associated with a more favourable outcome compared with *JAK2*^{wt} (n = 47; P = 0.019; Figure 3d).

Cox regression analysis

In the total cohort including RARS-T and RARS cases in univariate analysis, age (P = 0.020), ring sideroblast count (P = 0.008) and *SF3B1* mutational status (P < 0.001) were prognostically significant, but *SF3B1* mutational status was the only independent prognostic factor (P = 0.001) in multivariable analysis.

In the RARS-T cohort in univariate analysis, age (P = 0.038), ring sideroblast count (P = 0.014), $JAK2^{V617F}$ (P = 0.024) and $SF3B1^{mut}$ (P = 0.006) were prognostically significant but only age (P = 0.044), $JAK2^{V617F}$ (P = 0.016) and $SF3B1^{mut}$ (P = 0.021) were independent prognostic parameters in multivariable analysis. Survival was better in patients with age ≤ 80 years, $JAK2^{V617F}$ and SF3B1 mutations. Taking into account these three prognostic factors, a model for survival in RARS-T patients was constructed in which

each risk factor (that is, age >80 years, $SF3B1^{wt}$ and $JAK2^{wt}$) was worth 1 point and defined two groups of risk: high risk for patients with a score of 2 or 3 (n = 13) and low risk when the score is 0 or 1 (n = 84). Median survival in the high-risk group was 1.6 vs 8.0 years in the low-risk group (P < 0.001; corresponding survival rates at 2, 4 and 6 years are 31.4, 31.4 and 0% for high-risk patients vs 95.8, 84.4 and 62.8% for low-risk patients; Figure 4).

DISCUSSION

Up to now, *SF3B1* mutations have been studied in small series of RARS-T patients. In this study, we provide data on *SF3B1* mutations in a large cohort of RARS-T patients, which is, to the best of our knowledge, the largest published to date.

SF3B1 mutations were observed with a high frequency in both RARS-T and RARS patients (86.5% and 84.8%, respectively). These proportions are slightly higher than those already published, as *SF3B1* mutations have been found in 64–82.6% of RARS²¹⁻²⁴ and 66.7–72% in RARS-T.^{23,24} The slightly higher frequency of *SF3B1* mutations in the current study may be due to the larger cohort of RARS-T patients than in other studies. There may also be differences in methodology, for example, direct sequencing vs non-sequencing-based screening strategies.

However, RARS differs from RARS-T in that there are no $JAK2^{V617F}$ mutations in RARS,^{4,14} whereas there is a high frequency in RARS-T.^{4–13,15,19} We recently showed that RARS-T differed from RARS and ET from a clinical, biological and



1830

Figure 4. Kaplan–Meier analysis for survival according to two risk groups. High risk comprising patients with at least two of the following risk factors: age > 80 years, *SF3B1*wt, *JAK2*wt; and low risk: one or less of the risk factors.

prognostic point of view, suggesting that RARS-T could be considered as a unique entity.²⁰ The results of our current study showing the presence of both *SF3B1* and *JAK2*^{V617F} mutations in a high proportion of RARS-T confirm that RARS-T is a unique entity. Indeed, RARS-T was associated with high rates of *SF3B1* mutations (86.5%) and *JAK2*^{V617F} mutations while ET patients have a low frequency of *SF3B1* mutations, and in RARS, no *JAK2*^{V617F} mutations were detected.

A minority of RARS-T patients (13.5%) presented without *SF3B1* mutations. This could be due to another mutation affecting components of the RNA splicing machinery as mutations of *SRSF2*, *U2AF35* and *ZRSR2* have already been described in 12.4, 7.3 and 3.1% of MDS, respectively,³⁶ or due to mutations of proteins associated with SF3B1 (SF3B4 and SF3B14).

These results are in line with our hypothesis that RARS-T is an independent entity characterised by a particular molecular combination associating mutations that give a myeloproliferative advantage (*JAK2*^{V617F}, *MPL*^{W515R/L} or other unknown mutations) and mutations of components of the splicing machinery responsible for myelodysplastic features (*SF3B1* in most cases, and possibly *SRSF2*, *U2AF35* or *ZRSF2* in the remaining cases).

Conflicting results on the prognostic impact of *SF3B1* mutations in MDS have been reported as several studies noted a good prognostic impact in MDS, whereas others hypothesised that *SF3B1* mutations were associated with good-prognosis MDS subgroups but lost their prognostic impact in RARS and in RCMD-RS.²⁷

In our large cohort of RARS-T patients, *SF3B1* mutations were associated with female sex, higher ring sideroblast counts and a longer overall survival than in *SF3B1*^{wt} patients. In a multivariable analysis, age > 80 years at diagnosis, *SF3B1*^{wt} as well as *JAK2*^{wt} were independent factors of a worse prognosis. Based on these three independent parameters, a prognostic score for RARS-T patients was created to define two risk groups: high risk when there were two or three risk factors, low when there was only one or no risk factor. Median survival was 1.6 vs 8.0 years in the high-and low-risk group, respectively, underlying the relevance of such score in RARS-T patients.

Exploring *SF3B1* mutations in MDS associated with ring sideroblasts is of interest from a prognostic point of view, particularly as specific treatment will be available.^{37,38} Also, allele-specific PCR have been designed, and these could be useful for monitoring minimal residual disease in MDS.³⁹

In summary, this study confirms that RARS-T should be considered an independent entity. In RARS-T patients, age <80 years at diagnosis, *SF3B1* and *JAK2* mutations are independent factors for better survival and may be used to stratify patients.

CONFLICT OF INTEREST

SS and TH declare part ownership of the MLL Munich Leukemia Laboratory. TA, SJ and VG are employed by the MLL Munich Leukemia Laboratory. All the other authors declare no conflict of interest.

ACKNOWLEDGEMENTS

We thank the Medical Doctors from the Haematology Department and Laboratory of the University Hospital of Dijon, the Spanish Group of Hematological Cytology (GECH), as well as Philip Bastable for revising the manuscript. EL and FL are grateful to the Tumor Bank of the CHU of Bordeaux. This work was supported by grants from the association 'Tulipes contre le cancer' (Châlon s/Saône, Burgundy, France) and from FEHH (Spain) and 2009 SGR 541 (Generalitat de Catalunya).

REFERENCES

- 1 Vardiman JW, Bennett JM, Bain BJ, Baumann I, Thiele J, Orazi A. Myelodysplastic/ myeloproliferative neoplasms, unclassifiable. In: Swerdlow SH, Campo E, Lee Harris N, Jaffe ES, Pileri SA, Stein H *et al.* (eds) WHO Classification of Tumours of Haematopoietic and Lymphoid Tissue. 4th edn. IARC: Lyon, France, 2008, pp 85–86.
- 2 Hasserjian RP, Gatterman N, Bennett JM, Brunning RD, Thiele J. Refractory anemia with ringed sideroblasts. In: Swerdlow SH, Campo E, Lee Harris N, Jaffe ES, Pileri SA, Stein H *et al.* (eds) WHO Classification of Tumours of Haematopoietic and Lymphoid Tissue. 4th edn. IARC: Lyon, France, 2008, pp 96–97.
- 3 Thiele J, Kvasnicka HM, Orazi A, Tefferi A, Gisslinger H. Essential thrombocythaemia. In: Swerdlow SH, Campo E, Lee Harris N, Jaffe ES, Pileri SA, Stein H *et al.* (eds) *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissue.* 4th edn. IARC: Lyon, France, 2008, pp 48–50.
- 4 Ceesay MM, Lea NC, Ingram W, Westwood NB, Gaken J, Mohamedali A *et al.* The JAK2 V617F mutation is rare in RARS but common in RARS-T. *Leukemia* 2006; **20**: 2060–2061.
- 5 Boissinot M, Garand R, Hamidou M, Hermouet S. The JAK2-V617F mutation and essential thrombocythemia features in a subset of patients with refractory anemia with ring sideroblasts (RARS). *Blood* 2006; **108**: 1781–1782.
- 6 Flach J, Dicker F, Schnittger S, Kohlmann A, Haferlach T, Haferlach C. Mutations of JAK2 and TET2, but not CBL are detectable in a high portion of patients with refractory anemia with ring sideroblasts and thrombocytosis. *Haematologica* 2010; **95**: 518–519.
- 7 Gattermann N, Billiet J, Kronenwett R, Zipperer E, Germing U, Nollet F et al. High frequency of the JAK2 V617F mutation in patients with thrombocytosis (platelet count > 600 × 109/L) and ringed sideroblasts more than 15% considered as MDS/ MPD, unclassifiable. *Blood* 2007; **109**: 1334–1335.
- 8 Hellstrom-Lindberg E, Cazzola M. The role of JAK2 mutations in RARS and other MDS. *Hematology Am Soc Hematol Educ Program* 2008; 52–59.
- 9 Raya JM, Arenillas L, Domingo A, Bellosillo B, Gutierrez G, Luno E *et al.* Refractory anemia with ringed sideroblasts associated with thrombocytosis: comparative analysis of marked with non-marked thrombocytosis, and relationship with JAK2 V617F mutational status. *Int J Hematol* 2008; **88**: 387–395.
- 10 Remacha AF, Nomdedeu JF, Puget G, Estivill C, Sarda MP, Canals C et al. Occurrence of the JAK2 V617F mutation in the WHO provisional entity: myelodysplastic/ myeloproliferative disease, unclassifiable-refractory anemia with ringed sideroblasts associated with marked thrombocytosis. *Haematologica* 2006; **91**: 719–720.
- 11 Renneville A, Quesnel B, Charpentier A, Terriou L, Crinquette A, Lai JL et al. High occurrence of JAK2 V617 mutation in refractory anemia with ringed sideroblasts associated with marked thrombocytosis. *Leukemia* 2006; 20: 2067–2070.
- 12 Schmitt-Graeff AH, Teo SS, Olschewski M, Schaub F, Haxelmans S, Kirn A et al. JAK2V617F mutation status identifies subtypes of refractory anemia with ringed sideroblasts associated with marked thrombocytosis. *Haematologica* 2008; **93**: 34–40.



- 13 Szpurka H, Tiu R, Murugesan G, Aboudola S, Hsi ED, Theil KS *et al.* Refractory anemia with ringed sideroblasts associated with marked thrombocytosis (RARS-T), another myeloproliferative condition characterized by JAK2 V617F mutation. *Blood* 2006; **108**: 2173–2181.
- 14 Steensma DP, Tefferi A. JAK2 V617F and ringed sideroblasts: not necessarily RARS-T. *Blood* 2008; **111**: 1748.
- 15 Wang SA, Hasserjian RP, Loew JM, Sechman EV, Jones D, Hao S et al. Refractory anemia with ringed sideroblasts associated with marked thrombocytosis harbors JAK2 mutation and shows overlapping myeloproliferative and myelodysplastic features. *Leukemia* 2006; 20: 1641–1644.
- 16 Pardanani AD, Levine RL, Lasho T, Pikman Y, Mesa RA, Wadleigh M *et al.* MPL515 mutations in myeloproliferative and other myeloid disorders: a study of 1182 patients. *Blood* 2006; **108**: 3472–3476.
- 17 Schnittger S, Bacher U, Haferlach C, Dengler R, Krober A, Kern W *et al.* Detection of an MPLW515 mutation in a case with features of both essential thrombocythemia and refractory anemia with ringed sideroblasts and thrombocytosis. *Leukemia* 2008; **22**: 453–455.
- 18 Wardrop D, Steensma DP. Is refractory anaemia with ring sideroblasts and thrombocytosis (RARS-T) a necessary or useful diagnostic category? *Br J Haematol* 2009; **144**: 809–817.
- 19 Malcovati L, Della Porta MG, Pietra D, Boveri E, Pellagatti A, Galli A *et al*. Molecular and clinical features of refractory anemia with ringed sideroblasts associated with marked thrombocytosis. *Blood* 2009; **114**: 3538–3545.
- 20 Broseus J, Florensa L, Zipperer E, Schnittger S, Malcovati L, Richebourg S *et al.* Clinical features and course of refractory anemia with ring sideroblasts associated with marked thrombocytosis. *Haematologica* 2012; **97**: 1036–1041.
- 21 Yoshida K, Sanada M, Shiraishi Y, Nowak D, Nagata Y, Yamamoto R *et al.* Frequent pathway mutations of splicing machinery in myelodysplasia. *Nature* 2011; **478**: 64–69.
- 22 Papaemmanuil E, Cazzola M, Boultwood J, Malcovati L, Vyas P, Bowen D *et al.* Somatic SF3B1 mutation in myelodysplasia with ring sideroblasts. *N Engl J Med* 2011; **365**: 1384–1395.
- 23 Visconte V, Makishima H, Jankowska A, Szpurka H, Traina F, Jerez A *et al.* SF3B1, a splicing factor is frequently mutated in refractory anemia with ring sideroblasts. *Leukemia* 2012; **26**: 542–545.
- 24 Malcovati L, Papaemmanuil E, Bowen DT, Boultwood J, Della Porta MG, Pascutto C et al. Clinical significance of SF3B1 mutations in myelodysplastic syndromes and myelodysplastic/myeloproliferative neoplasms. Blood 2011; 118: 6239–6246.
- 25 Damm F, Thol F, Kosmider O, Kade S, Loffeld P, Dreyfus F *et al.* SF3B1 mutations in myelodysplastic syndromes: clinical associations and prognostic implications. *Leukemia* 2012; **26**: 1137–1140.
- 26 Damm F, Kosmider O, Gelsi-Boyer V, Renneville A, Carbuccia N, Hidalgo-Curtis C *et al.* Mutations affecting mRNA splicing define distinct clinical phenotypes and

correlate with patient outcome in myelodysplastic syndromes. *Blood* 2012; **119**: 3211–3218.

- 27 Patnaik MM, Lasho TL, Hodnefield JM, Knudson RA, Ketterling RP, Garcia-Manero G et al. SF3B1 mutations are prevalent in myelodysplastic syndromes with ring sideroblasts but do not hold independent prognostic value. *Blood* 2012; **119**: 569–572.
- 28 Visconte V, Makishima H, Maciejewski JP, Tiu RV. Emerging roles of the spliceosomal machinery in myelodysplastic syndromes and other hematological disorders. *Leukemia* 2012; 26: 2447–2454.
- 29 Visconte V, Rogers HJ, Singh J, Barnard J, Bupathi M, Traina F et al. SF3B1 haploinsufficiency leads to formation of ring sideroblasts in myelodysplastic syndromes. *Blood* 2012; **120**: 3173–3186.
- 30 Jeromin S, Haferlach T, Grossmann V, Alpermann T, Kowarsch A, Haferlach C et al. High frequencies of SF3B1 and JAK2 mutations in refractory anemia with ring sideroblasts associated with marked thrombocytosis strengthen the assignment to the category of myelodysplastic/myeloproliferative neoplasms. *Haematologica* 2013; **98**: 15–17.
- 31 Cazzola M, Rossi M, Malcovati L. Biologic and clinical significance of somatic mutations of SF3B1 in myeloid and lymphoid neoplasms. *Blood* 2013; **121**: 260–269.
- 32 Lippert E, Boissinot M, Kralovics R, Girodon F, Dobo I, Praloran V et al. The JAK2-V617F mutation is frequently present at diagnosis in patients with essential thrombocythemia and polycythemia vera. Blood 2006; 108: 1865–1867.
- 33 Schnittger S, Bacher U, Kern W, Schroder M, Haferlach T, Schoch C. Report on two novel nucleotide exchanges in the JAK2 pseudokinase domain: D620E and E627E. *Leukemia* 2006; 20: 2195–2197.
- 34 Schnittger S, Bacher U, Haferlach C, Geer T, Muller P, Mittermuller J *et al.* Detection of JAK2 exon 12 mutations in 15 patients with JAK2V617F negative polycythemia vera. *Haematologica* 2009; **94**: 414–418.
- 35 Schnittger S, Bacher U, Haferlach C, Beelen D, Bojko P, Burkle D et al. Characterization of 35 new cases with four different MPLW515 mutations and essential thrombocytosis or primary myelofibrosis. *Haematologica* 2009; 94: 141–144.
- 36 Thol F, Kade S, Schlarmann C, Loffeld P, Morgan M, Krauter J *et al.* Frequency and prognostic impact of mutations in SRSF2, U2AF1, and ZRSR2 in patients with myelodysplastic syndromes. *Blood* 2012; **119**: 3578–3584.
- 37 Rymond B. Targeting the spliceosome. Nat Chem Biol. 2007; 3: 533-535.
- 38 Webb TR, Joyner AS, Potter PM. The development and application of small molecule modulators of SF3b as therapeutic agents for cancer. *Drug Discov Today* 2013; **18**: 43–49.
- 39 Matsuda K, Ishida F, Ito T, Nakazawa H, Miura S, Taira C et al. Spliceosome-related gene mutations in myelodysplastic syndrome can be used as stable markers for monitoring minimal residual disease during follow-up. Leuk Res 2012; 36: 1393–1397.