

Targeted Analysis of Tears Revealed Specific Altered Metal Homeostasis in Age-Related Macular Degeneration

Eva Valencia,^{1,2} Montserrat García,^{1,3} Beatriz Fernández-Vega,^{1,3} Rosario Pereiro,^{1,2} Lara Lobo,² and Héctor González-Iglesias^{1,3,*}

¹Ophthalmological Research Foundation, University Institute Fernández-Vega, University of Oviedo, Oviedo, Spain

²Department of Physical and Analytical Chemistry, Faculty of Chemistry, University of Oviedo, Oviedo, Spain

³Ophthalmological Institute Fernández-Vega, Oviedo, Spain

Correspondence: Lara Lobo, Department of Physical and Analytical Chemistry, University of Oviedo, Oviedo, Spain; lobolara@uniovi.es.

Héctor González-Iglesias, Ophthalmological Institute Fernández-Vega, Avda. Dres. Fernández-Vega, 34, 33012 Oviedo, Spain; h.gonzalez@fio.as.

Current affiliation: *Instituto de Productos Lácteos de Asturias, Consejo Superior de Investigaciones Científicas, (IPLA-CSIC) Villaviciosa, Spain.

Received: November 11, 2021

Accepted: March 29, 2022

Published: April 15, 2022

Citation: Valencia E, García M, Fernández-Vega B, Pereiro R, Lobo L, González-Iglesias H. Targeted analysis of tears revealed specific altered metal homeostasis in age-related macular degeneration. *Invest Ophthalmol Vis Sci.* 2022;63(4):10. <https://doi.org/10.1167/iovs.63.4.10>

PURPOSE. Specific altered metal homeostasis has been investigated in the tear film of age-related macular degeneration (AMD) patients considering that metal dyshomeostasis contributes to the production of free radicals, inflammation, and apoptosis and results in conformational changes of proteins.

METHODS. A multitargeted approach based on spectrophotometry and mass spectrometry techniques has been implemented to the multiplexed quantitation of lactoferrin (LF), S100 calcium binding protein A6 (S100A6), metallothionein 1A (MT1A), complement factor H (CFH), clusterin (CLU), amyloid precursor protein (APP), Mg, P, Na, Fe, Cu, Zn, and Ca, in the tear film from 60 subjects, 31 patients diagnosed with the dry form of AMD, and 29 healthy individuals

RESULTS. Significant up-regulations of MT1A (1.9-fold) and S100A6 (1.4-fold) and down-regulations of LF (0.7-fold), Fe (0.6-fold), Mg (0.7-fold), and Cu (0.7-fold) were observed in AMD patients, when compared to control subjects. Of all the studied variables, only APP showed negative correlation with age in the AMD group. Also, positive correlations were observed for the variables Mg and Na, Cu and Mg, and P and Mg in both the AMD and control groups, whereas positive correlations were exclusively determined in the AMD group for Cu and LF, Na and Ca, and Mg and Ca. The panel constituted of MT1A, Na, and Mg predicts AMD disease in 73% of cases.

CONCLUSIONS. The different levels of target metals and (metallo-)proteins in the tear film suggest altered metal homeostasis in AMD patients. These observed pathophysiological changes may be related with the anomalous protein aggregation in the macula.

Keywords: age-related macular degeneration, tear, targeted analysis, (metallo-)proteins, metals, dyshomeostasis

Ageing is an irreversible process inevitably associated with disease, being the predominant risk factor for most conditions that limit health span. According to data from the World Population Prospects 2019,¹ one in eleven people in the world was over the age of 65 in 2019 (9%), which is expected to increase to one in six people by 2050 (16%). The greater aging of the population turns some diseases from occasional to very frequent, with particular impact in neurodegenerative pathologies. Age-related neurodegenerative diseases are a group of disorders sharing common features and molecular patterns characterized by abnormal accumulation of proteins and selective neuronal degeneration and death.² Insoluble oligomerized protein deposition has been identified within the eye during the onset of age-related macular degeneration (AMD). Actually, AMD is the leading cause of noninherited irreversible vision loss in developed countries and a very prevalent cause of legal blindness in the elderly.³ Early AMD initiates with the formation of extracellular deposits, called *drusen*, between the

basal lamina of the retinal pigment epithelium (RPE) and the Bruch's membrane, which progressive degeneration leads to the advanced stages of the disease, the neovascular (or wet) or geographic atrophy (or dry) forms of AMD.⁴ Current therapies exist for neovascular AMD, representing 10% to 15% of cases, but there is no approved treatment for dry AMD, accounting for 85% of cases of advanced disease.^{5,6}

Drusen composition is highly complex, containing lipids, minerals, and proteins abnormally aggregated,⁷⁻¹¹ the formation molecular mechanisms of which include protein misfolding induced by metals.^{12,13} The impairment of intracellular homeostatic control of metals underlies their cytotoxic effects, in the oligomerization of proteins and in the aggregation and consequent loss of functionality of molecules. Also, metal dyshomeostasis contributes to the production of free radicals, inflammation, and apoptosis and results in conformational changes of proteins involved in neurodegeneration.¹⁴⁻¹⁶ Among others, dyshomeostasis of Zn, Fe, Cu, or Ca induces various detrimental intracellu-



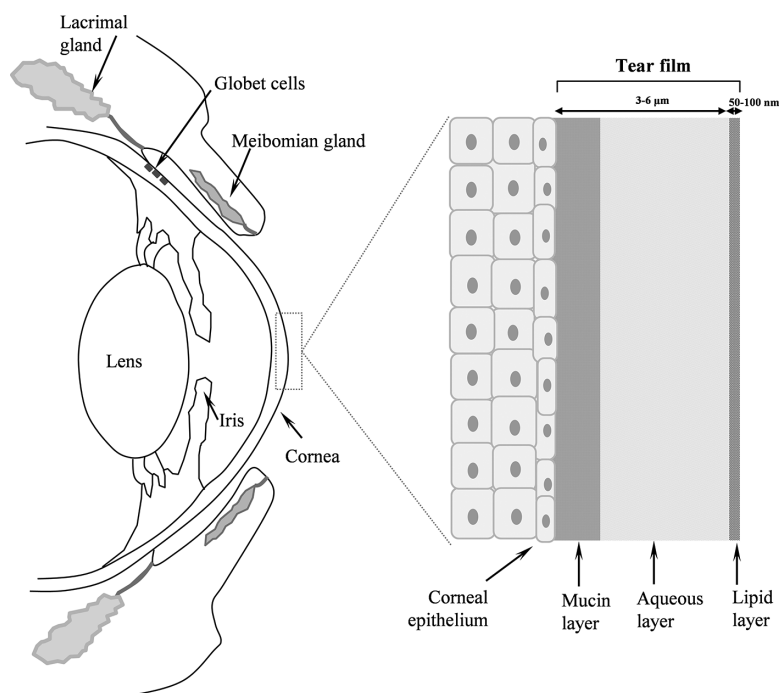


FIGURE 1. Diagram summarizing the lacrimal apparatus and the tear film structure.

lar events, including oxidative stress, DNA fragmentation, protein misfolding, and activation of apoptosis, which leads to neuronal death.^{17,18} Therefore specifically altered metal homeostasis may occur in AMD disease, at both the local and systemic levels. Considering that is required a major understanding of the effects of metals changes during neurodegenerative processes, the application of innovative analytical and biochemical approaches, including (metallo-)proteomic analysis, may contribute to shed light on metal-protein interactions and their pathogenic effects in AMD.

Although neurodegeneration during AMD occurs in the RPE and neurosensory retina, possible changes could be observed in the anterior segment of the eye globe, specifically in readily accessible fluids with noninvasive sampling such as tear film.¹⁹ The tear film, produced by Meibomian, lacrimal, and accessory glands, along with goblet cells,²⁰ is composed of three layers: the inner hydrophilic mucin layer, the middle aqueous layer, and the outer lipid layer (see Fig. 1).²¹ This thin fluid layer (up to 6 μm thickness) is very promising for the discovery and implementation of biomarkers or to study local alterations related to eye diseases, because it contains a comparatively simple proteome composed of a variety of molecules and removes local waste products, drugs, and disease-related media.²² Tear fluid permits a noninvasive procedure for sampling by Schirmer test papers or glass microcapillaries, requiring no incision into the body or tissue removal.²³ However, the conjunctival sac has a capacity of approximately 15 to 30 μL , and the natural tear film volume is 7 to 8 μL , severely limiting the multitargeted analysis. This is most dramatic when studying the elderly population, because tear production or secretion starts to be impaired.^{20,24} So far, limited studies have been carried out for the discovery of specific changes in the tear film associated with AMD neurodegenerative progression.^{25,26} Nevertheless, a major constraint remains in the lack of powerful methodologies for a multidisciplinary

approach, because conventional biological/biomedical techniques only address a specific part of the problem, and tear film represents a great challenge. Therefore methods for the quantitative multiparametric analysis of molecules and metals in tear film are currently mandatory. In this work, we have implemented a multitargeted approach based on spectrophotometry and mass spectrometry techniques to decipher possible existing changes in the homeostasis of metals, metalloproteins, and protein-related metals in AMD patients. To this end, quantitative analysis of lactoferrin (LF), S100 calcium-binding protein A6 (S100A6), metallothionein 1A (MT1A), complement factor H (CFH), clusterin (CLU), amyloid precursor protein (APP), Mg, P, Na, Fe, Cu, Zn, and Ca have been carried out in low volume of tears from control subjects and patients diagnosed with the dry form of AMD.

METHODS

Study Subjects

The present observational, prospective and case-controlled study involved 60 subjects: 31 patients diagnosed with the dry form of AMD and 29 healthy individuals (controls). The study adheres to the tenets of the Declaration of Helsinki on Biomedical Research Involving Human Subjects and was approved by the Clinical Research Ethics Committee at Principality of Asturias (Oviedo, Spain). Study subjects were recruited at the Institute of Ophthalmology Fernández-Vega (Oviedo, Spain) based on the agreed inclusion and exclusion criteria, signed informed consent, and the research was conducted by medically qualified personnel. Complete ophthalmic examinations were performed for patients and controls, including slit-lamp biomicroscopy and funduscopy in both eyes. AMD-diagnosed patients were further examined by fluorescence fundus angiography or optical coherence tomography. Only individuals with dry AMD and

evidence of an active form of geographic atrophy in at least one eye and absence of choroidal neovascularization were included. Control subjects were selected from patients over 60 years old and with absence of AMD or glaucoma. Specifically, controls did not show any sign of drusen, including reticular pseudodrusen, or pigment abnormalities in none of the eyes, being in stage 1 of the Clinical Age-Related Maculopathy Staging classification (i.e., no drusen).²⁷ The exclusion criteria for both controls and AMD patients was the absence of other relevant ocular pathologies, such as diabetic retinopathy or any ocular surface disease including dry eye syndrome, eye surface disorders, glaucoma, and previous ocular surgery except for cataract extraction. Those patients or healthy subjects using topical medications or contact lens were also discarded to avoid influences in the tear film composition.

Sample Collection and Preparation

Tear film was collected from each eye, when possible, using calibrated glass microcapillary tubes with 10 μL of capacity (Blaubrand intraMark, Wertheim, Germany). Sample collection was performed in the morning hours, between 9 AM and 12 PM, using sterile gloves. The microcapillaries were placed into the lower conjunctival sacs of each eye avoiding any contact with the ocular surface to minimize its irritation, obtaining the tear samples from the inferior tear meniscus without anesthesia. Once collected, tear samples from each individual eye were immediately placed in Eppendorf tubes using a plastic syringe and stored at -80°C until analysis. The typically collected tear volume reached 1 to 5 μL .

Molecular Assays

Quantitation of six proteins of interest was performed using commercial ELISAs following the instructions of the manufacturer. Commercially available sandwich-type ELISA kits from Abcam (Cambridge, UK) were used for the quantification of four of the candidate proteins, the human LF (ab200015), the human CFH (ab252359), the human CLU (ab174447), and the human APP (ab216944). Commercial ELISA kits purchased from Abynetek Biopharma (Vizcaya, Spain) and LS BIO (Seattle, WA, USA) were used for the analysis of human S100A6 (ABK1-E1028) and MT1A (LS-F10296), respectively. For APP, CFH and S100A6 quantitation the tear samples were diluted 1:200, for MT1A quantitation the tear samples were diluted 1:1,000, for CLU quantitation the tear samples were diluted 1:6,000 and for the LF quantitation tear samples were diluted 1:1,000,000. The concentrations of targeted proteins were expressed in nanograms of the protein per milliliter of tear for APP, CFH, S100A6, MT1 and CLU, and in milligrams of the protein per milliliter of tear for LF. Representative analytical parameters for each of ELISA assays have been included in Supplementary Table S1.

Total protein concentration in tears was determined using the commercial QuantiPro BCA Assay Kit (Sigma Aldrich, Madrid, Spain). The calibration range was 0 to 50 $\mu\text{g mL}^{-1}$ with BSA as standard. The tear samples were diluted 1:2500. A working solution containing copper compound and bicinchoninic acid was added to the samples. The proteins present in tears, reduced Cu^{2+} to Cu^{+} and a complex between copper and bicinchoninic acid was formed, the absorbance of which was measured at 562 nm wavelength. All absorbance measurements were monitored using

the spectrophotometer PerkinElmer 2030 Multilabel Reader VICTORTM X5 (Waltham, MA, USA).

Multielemental Quantitation Using Flow Injection Analysis (FIA) and Inductively Coupled Plasma Mass Spectrometry (ICP-MS) Detection

Multi-elemental analysis of Ca, Mg, P, Na, Zn, Fe and Cu was performed by external calibration using a 7900 ICP-MS (Agilent, Santa Clara, CA, USA) equipped with collision/reaction cell pressurized with He gas ($4.3 \text{ mL} \cdot \text{min}^{-1}$ flow rate) to minimize polyatomic interferences. As internal standard, Ga was used for matrix effect correction. The FIA system, consisting of a six-channel Rheodyne valve 3125 (Rheodyne, Rohnert Park, CA, USA), with a 5 μL loop, was on line coupled to the ICP-MS. A glass syringe of 100 μL volume (Hamilton, Reno, NV, USA) was used to load samples and standards through the system.

The standard stock solution of Mg, P, Fe, Cu, Zn ($1,000 \text{ mg} \cdot \text{L}^{-1}$), Na, Ca ($10,000 \text{ mg} \cdot \text{L}^{-1}$) and Ga ($1,000 \text{ mg} \cdot \text{L}^{-1}$) were purchased from Merck (Darmstadt, Germany) and used for the external calibration curve. The concentration range for calibration was 0 to 200 $\mu\text{g} \cdot \text{L}^{-1}$ for Mg and P, 0 to 500 $\mu\text{g} \cdot \text{L}^{-1}$ for Na, Zn, and Fe, 0 to 60 $\mu\text{g} \cdot \text{L}^{-1}$ for Cu, and 0 to 1 $\text{mg} \cdot \text{L}^{-1}$ for Ca. Each calibration standard was injected five times. To determine the multielemental concentration in tear film, samples were diluted with 0.14 M HNO_3 (trace metal grade; Fisher Scientific, Waltham, MA, USA) using a 1:20 dilution for most of the elements, with the exception of Na requiring a higher dilution of 1:2000. Thus only 1 μL of tear film from each subject was used to determine their multielemental composition.

Statistical Analysis

Demographic characteristics of AMD patients and control subjects were performed with SPSS Statistics version 15.0 (IBM Corp., Armonk, NY, USA). In the case of quantitative variables (age), significant differences were analyzed using the Mann-Whitney U test, and qualitative variables (gender and the presence of cataracts, arterial hypertension or cardiovascular disease) were analyzed using the χ^2 test. Comprehensive studies of all obtained data from both elemental and molecular analysis, reported as mean \pm standard error of the mean, was carried out using SPSS Statistics version 15.0 and GraphPad Prism version 3 for Windows (San Diego, CA, USA). Normal distribution was evaluated by the Shapiro-Wilk's test, and, subsequently, significant differences between control and AMD groups for proteins and elements were analyzed using the Mann-Whitney U test for those without normal tendency, or Student's *t* test was selected for normal distribution. Correlation analysis was carried out using SPSS Statistics version 15.0. A *P* value <0.05 was considered statistically significant. Further stepwise discrimination analysis was carried out with SPSS using the concentrations of the proteins and elements determined by ELISA and FIA-ICP-MS, respectively, in the different sample groups. Additionally, logistic regression analysis using SPSS software was performed for further control of potential confounders using SPSS.

Several statistical tools based on Machine Learning approaches were applied using Orange Canvas software v2.6 (<http://orange.biolab.si>) to assess which method provided the best accuracy for the correct classification of samples

based on the selected panel of proteins or elements. These tools included receiver operating characteristic curve analyses for each of the markers, naive Bayes, k-nearest neighbor, random forest, classification trees, and support vector machine.²⁸

RESULTS

Patients

Demographic characteristics of AMD patients and control subjects are collected in Table 1. The population origin recruited in the present study was Caucasian and mostly from the northernmost regions of Spain. No statistically significant differences were obtained for age, gender, the presence of cataracts, arterial hypertension, or cardiovascular disease, when compared dry AMD cases with controls ($P > 0.05$). Statistically significant differences were observed for dyslipidemia in AMD patients when compared to control subjects ($P = 0.002$). The dry AMD group consisted of 31 individuals, comprising 18 women and 13 men, with a mean age of 80.7 ± 8.0 years, whereas the control group included 29 subjects, formed by 18 females and 11 males, with a mean age of 77.2 ± 8.9 years. In both groups, the percentage of cataractous subjects was similar, specifically 75.9% for controls and 77.4% for AMD patients. The significantly higher percentage of individuals with dyslipidemia in the AMD group (45.2%), compared to controls (6.9%), indicates a possible causal role of plasma lipid levels in the pathogenesis of AMD.^{29,30} Systemic treatments were assessed, including the common oral treatments for both control and AMD subjects (Supplementary Table S2).

Targeted Molecular Analysis of Tear Film

Total concentrations of LF, S100A6, CFH, CLU, APP and MT1A were individually quantified in 29 control subjects and

31 dry AMD patients, using ELISA assays (Table 2), dot plots of which are shown in Figure 2. Tear samples from both eyes of same individual were pooled, when possible. Because of the low available sample volume, molecular composition was determined in most, but not all, study subjects. Of all the analyzed proteins, only LF, S100A6, and CLU have been previously quantified in tears.³¹⁻³⁴ In our experiments, statistical significant differences were observed for the proteins LF (0.7-fold change, $P = 0.004$), S100A6 (1.4-fold change, $P = 0.03$) and MT1A (1.9-fold change, $P = 0.006$), when comparing AMD patients and control subjects (see Figs. 2A, 2B, 2F). By contrast, no significant differences were observed for CFH, CLU, and APP (Figs. 2C-E; $P > 0.05$). Correlations between the proteins analyzed and the age of the recruited individuals were evaluated using Pearson coefficient, showing only significant negative correlation for APP exclusively in the AMD group (Fig. 2G; $r = -0.523$, $P = 0.005$). Averaged total protein concentration in tears for control individuals reached $8.87 \pm 4.84 \text{ mg} \cdot \text{mL}^{-1}$, whereas for AMD patients it was $9.26 \pm 5.28 \text{ mg} \cdot \text{mL}^{-1}$, with no statistically significant differences observed between groups, being in the range of reference published.³⁵

Targeted Multielemental Analysis of Tear Film

Quantitative multiparametric elemental analysis of Ca, Mg, P, Na, Cu, Fe and Zn was carried out in individual human tears by FIA-ICP-MS (Table 2). Dot plots depicting changes in the concentrations of each element, when comparing AMD and control groups are shown in Figure 3, in which normal distribution and unpaired t test was selected to examine possible variations between cohorts. Significant down-regulation was observed for Mg (0.7-fold change, $P = 0.01$, $9.74 \pm 3.68 \text{ } \mu\text{g} \cdot \text{mL}^{-1}$ control group vs. $6.50 \pm 3.27 \text{ } \mu\text{g} \cdot \text{mL}^{-1}$ AMD group), Fe (0.6-fold change, $P = 0.03$, $3.53 \pm 2.01 \text{ } \mu\text{g} \cdot \text{mL}^{-1}$ control group vs. $2.13 \pm 1.37 \text{ } \mu\text{g} \cdot \text{mL}^{-1}$ AMD group) and Cu (0.7-

TABLE 1. Demographic Characteristics of AMD Patients and Controls

Study Population (n)	Age (Mean \pm SD)	Age Range	Gender (Female/Male)	Cataracts	AHT	Dyslipidemia	Cardiovascular Disease
Controls (n = 29)	77.2 \pm 8.9	63–89	18 (62.1%)/11	22 (75.9%)	13 (44.8%)	2 (6.9%)	2 (6.9%)
AMD (n = 31)	80.7 \pm 8.0	62–101	18 (58.1%)/13	24 (77.4%)	14 (45.1%)	14 (45.2%)	3 (9.7%)

n, number of subjects; SD, standard deviation; AHT, arterial hypertension.

TABLE 2. Concentrations of Proteins and Elements Determined in the Tear Film of AMD Patients and Control Subjects

	Control	AMD	Fold-change	P Value
Protein concentrations				
Lactoferrin (average \pm SD, mg/mL)	11.81 \pm 3.98	8.11 \pm 3.47	0.7	0.004
S100A6 (average \pm SD, ng/mL)	409.8 \pm 158.9	568.59 \pm 321.9	1.4	0.03
CFH (average \pm SD, ng/mL)	1692 \pm 1167	2051 \pm 1573	1.2	0.3
Clusterin (average \pm SD, mg/mL $\mu\text{g}/\text{mL}$)	29.33 \pm 13.01	35.99 \pm 14.94	1.2	0.09
APP (average \pm SD, ng/mL)	176.71 \pm 76.04	161.88 \pm 69.86	0.9	0.3
MT1A (average \pm SD, ng/mL)	154.45 \pm 36.56	293.55 \pm 214.00	1.9	0.006
Element concentrations				
Na (average \pm SD, $\mu\text{g}/\text{mL}$)	2125 \pm 524	1783 \pm 678	0.8	0.09
Mg (average \pm SD, $\mu\text{g}/\text{mL}$)	9.74 \pm 3.68	6.50 \pm 3.27	0.7	0.01
P (average \pm SD, $\mu\text{g}/\text{mL}$)	3.95 \pm 2.25	2.94 \pm 2.14	0.7	0.2
Ca (average \pm SD, $\mu\text{g}/\text{mL}$)	23.98 \pm 9.15	22.46 \pm 12.91	0.9	0.7
Fe (average \pm SD, $\mu\text{g}/\text{mL}$)	3.53 \pm 2.01	2.13 \pm 1.37	0.6	0.03
Cu (average \pm SD, $\mu\text{g}/\text{mL}$)	0.17 \pm 0.07	0.11 \pm 0.04	0.7	0.009
Zn (average \pm SD, $\mu\text{g}/\text{mL}$)	0.36 \pm 0.26	0.27 \pm 0.21	0.7	0.3

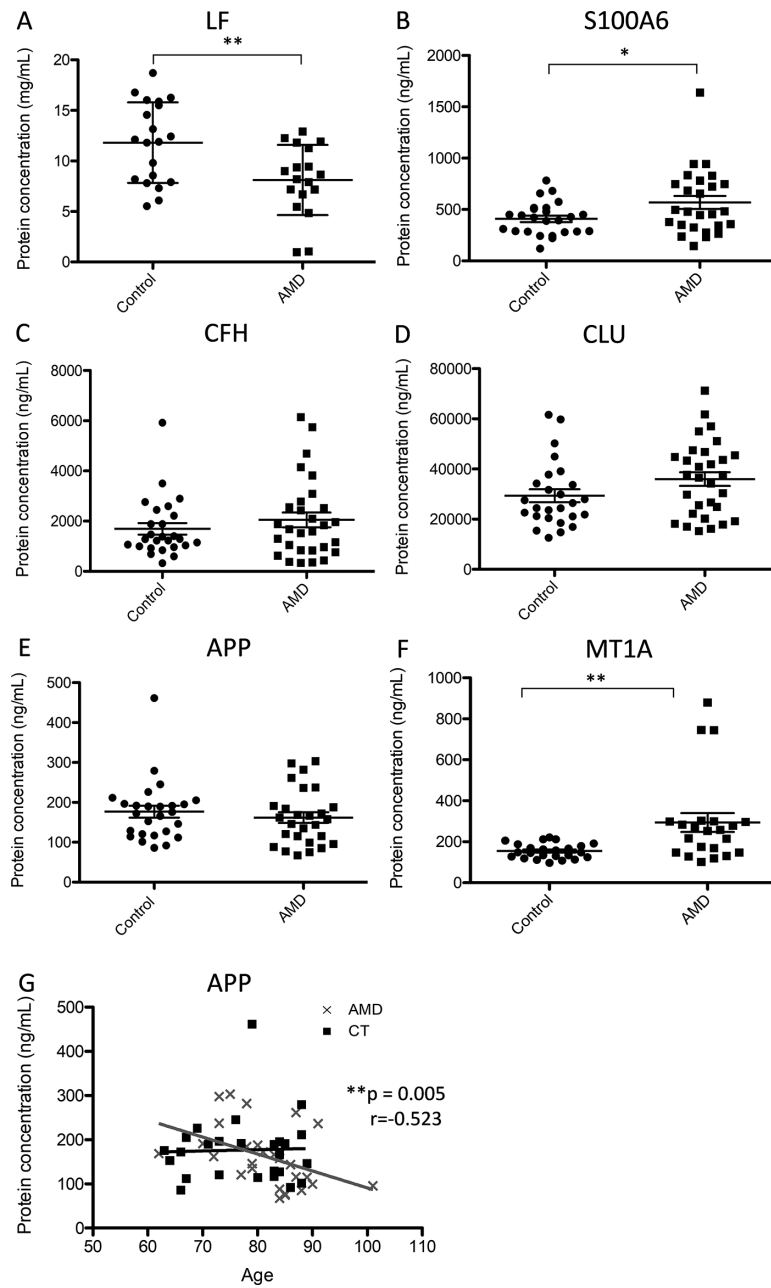


FIGURE 2. Dot plots of (A) LF, (B) S100A6, (C) CFH, (D) CLU, (E) APP, and (F) MT1A protein concentrations determined in the tear film of dry AMD patients and control subjects. (G) Correlation between APP concentrations of AMD patients and control individuals and aging. r value, Pearson correlation coefficient. * P value < 0.05; ** P value < 0.01; *** P value < 0.001.

fold change, $P = 0.009$, $0.17 \pm 0.07 \mu\text{g} \cdot \text{mL}^{-1}$ control group vs. $0.11 \pm 0.04 \mu\text{g} \cdot \text{mL}^{-1}$ AMD group), when comparing AMD patients and control subjects (see Figs. 3B, 3E, and 3G). Correlations between all the elements analyzed and the age of the recruited individuals were also evaluated using Pearson coefficient, showing no statistically significant correlation for none of the variables.

Study of Variable Associations

A correlation study among elements and proteins was carried out in AMD patients and control subjects using the Pearson coefficient, shown in Figure 4 the obtained signifi-

cant associations. Higher negative or positive r values indicate the grade of correlation between variables. Positive correlations were observed for both AMD and control groups when association of the variables Mg and Na ($r = 0.716$, $P = 0.003$ in the AMD group; $r = 0.711$, $P = 0.0002$ in the control group), Cu and Mg ($r = 0.556$, $P = 0.038$ in the AMD group; $r = 0.706$, $P = 0.0003$ in the control group), and P and Mg ($r = 0.705$, $P = 0.007$ in the AMD group; $r = 0.756$, $P = 0.00007$ in the control group) were studied (see Figs. 4A–C). Highly significant positive correlations were observed for the AMD group when association of the variables Cu and LF ($r = 0.839$, $P = 0.002$), Na and Ca ($r = 0.710$, $P = 0.006$), and Mg and Ca ($r = 0.835$, $P = 0.0003$) were evaluated (Figs. 4D–F).

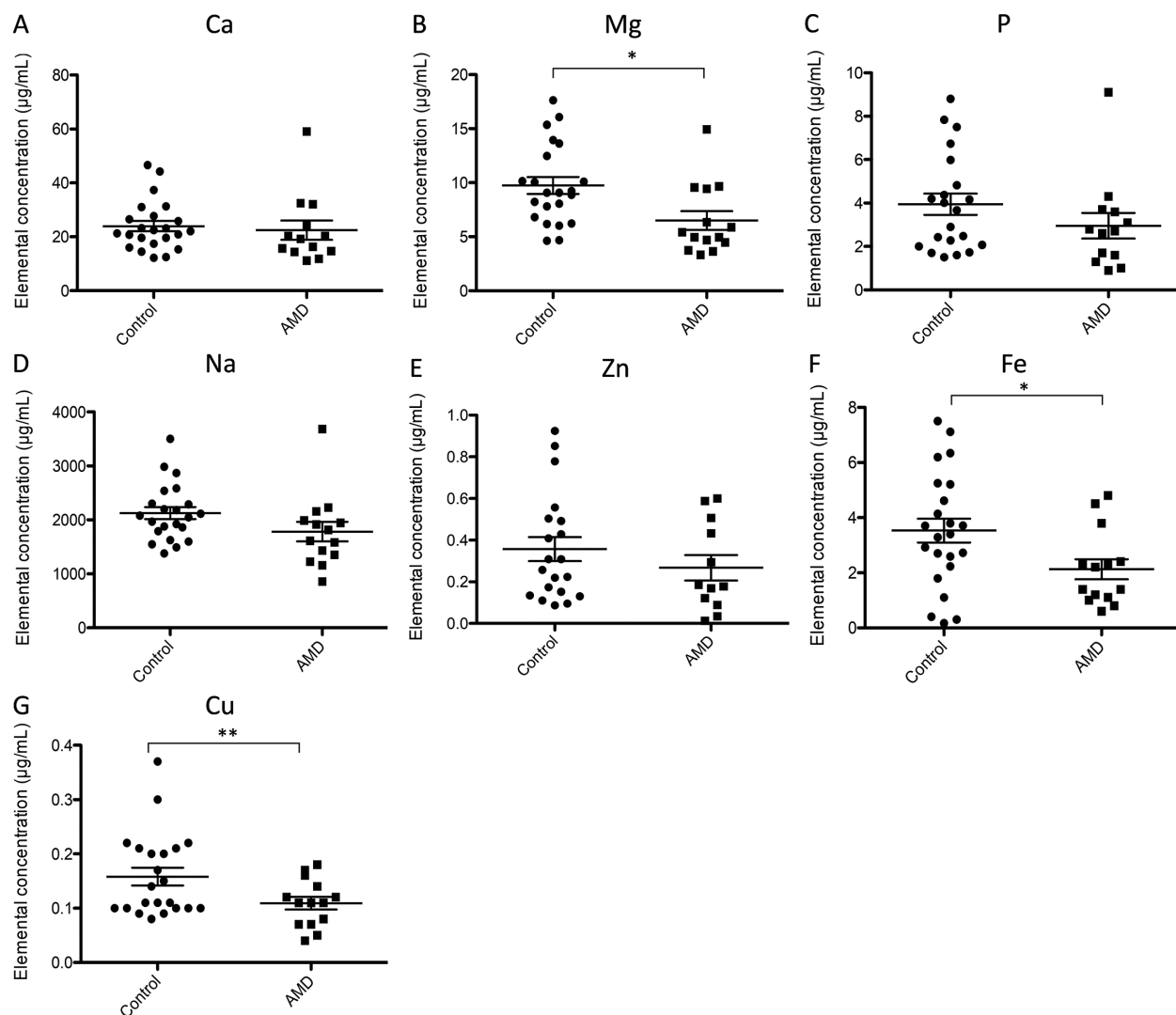


FIGURE 3. Dot plots of (A) Ca, (B) Mg, (C) P, (D) Na, (E) Zn, (F) Fe, and (G) Cu elemental concentrations determined in the tear film of dry AMD patients and control subjects. * P value < 0.05; ** P value < 0.01; *** P value < 0.001.

Data Filtering and Logistic Regression

The observed significant differences between AMD and control groups including individuals with dyslipidemia were further explored in a two-step comprehensive statistical analysis to evaluate the influence of this systemic risk factor in the multitargeted results obtained. First, the dyslipidemic patients were excluded and filtered data and parameters were re-analyzed. Total concentrations of LF, S100A6, CFH, CLU, APP, MT1A, Ca, Mg, P, Na, Cu, Fe and Zn obtained for control subjects ($n = 27$) and AMD patients ($n = 17$), all of them without dyslipidemia, are shown in Supplementary Table S3. Overall, comparable results to those considering all subjects (Table 2) were obtained, with the exception of Mg. When considering all patients, MT1A, S100A6, LF, Fe, and Cu showed similar fold change (1.9-, 1.4-, 0.7-, 0.6-, 0.7-fold, respectively) to when dyslipidemic subjects were excluded (2.1-, 1.4-, 0.5-, 0.5-, 0.6-fold, respectively), with even smaller P values in the latter case (see Supplementary Table S3). Only Mg lost the significant differences when dyslipidemic subjects were excluded (0.7-fold, $P = 0.07$), as compared

with unfiltered data (0.7-fold, $P = 0.01$), although the fold-change was the same. This difference in Mg may be related to the lower number on individuals resulted after filtering subjects with of dyslipidemia as a risk factor.

Second, a multivariate logistic regression was carried out in all recruited subjects (31 AMD patients and 29 control subjects), using SPSS with the unfiltered data, to evaluate the role of several confounders, including dyslipidemia, in the differences observed between the AMD group in comparison to control subjects. This analysis confirmed that the risk factor dyslipidemia ($P = 0.989$) did not contribute to the statistically significant differences observed between groups, discarding possible confounders among the demographic conditions and the studied proteins and elements, and therefore the altered levels of MT1A, S100A6, LF, Fe, and Cu in the tear film between both cohorts are result of AMD disease.

Machine Learning Models

The concentrations determined for the six proteins and the seven elements in tear samples from AMD patients and

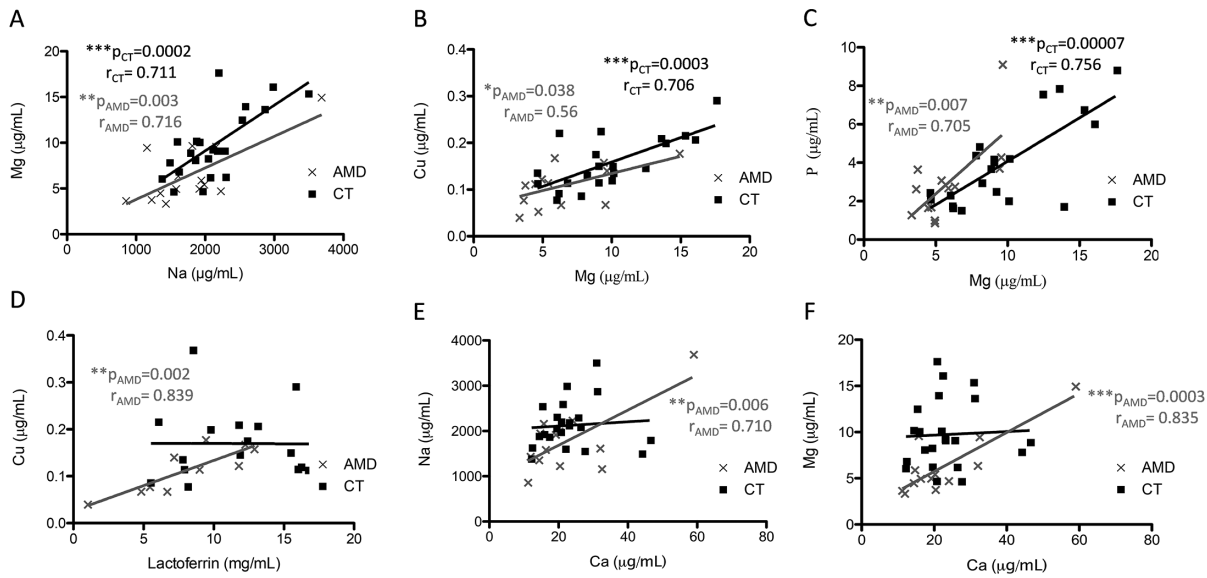


FIGURE 4. Correlation analysis between (A) Na and Mg, (B) Mg and Cu, (C) Mg and P, (D) LF and Cu, (E) Ca and Na, and (F) Ca and Mg, respectively. *r* value, Pearson correlation coefficient. **P* value < 0.05; ***P* value < 0.01; ****P* value < 0.001.

TABLE 3. Best Classifying Machine-Learning Models Using the Variables MT1A, Na, and Mg, When Comparing AMD Patients and Control Subjects

Method	CA	AUC	Sensitivity	Specificity	Precision
Random forest	0.63	0.73	0.82	0.41	0.61
Naive Bayes	0.73	0.86	0.58	0.90	0.87
k-Nearest	0.54	0.59	0.39	0.71	0.60
Neighbors					
Classification tree	0.68	0.70	0.69	0.67	0.70
Support vector machine	0.64	0.71	0.77	0.50	0.73

AUC, area under the curve; CA, correct assignment.

control subjects were used to perform a stepwise discriminant analysis by SPSS to determine the best variables classifying both groups. The panel consisting of MT1A, Na, and Mg correctly assigns the 89% of the samples group and consequently was used to generate multivariate predictive models. The whole data set (test on train data) was used for training and then for testing with distinct machine learning algorithms, including random forest, naive Bayes, k-nearest neighbor, classification tree, and support vector machine. Later, the classification method was validated using the remaining 30% of data to make predictions on future cases for which the target answer is unknown, providing the correct assignment (CA), sensitivity, specificity, area under the curve, and precision of the model (see Table 3). The best classification efficiency was obtained when using the MT1A, Na, and Mg panel and training the data with classification tree and naive Bayes, providing values of correct assignment of 68% and 73%, respectively, when comparing AMD and control groups, the percentage of which represents the correctly classified predictions. The obtained area under the curve reached 0.70 to 0.86 when using classification tree and naive Bayes, respectively.

DISCUSSION

This study revealed evidences of a specific altered metal homeostasis in the course of AMD disease, including up-regulation of MT1A and S100A6, and down-regulation of LF, Fe, Mg, and Cu in the tear film of individuals with macular disorder. AMD is characterized by a progressive loss of central vision caused by age-related neurodegenerative changes in the highly specialized region of the central retina responsible for fine and color vision, the macula. During aging, the systemic metabolism of metals, including Fe, Cu, Zn, and Ca, changes, and their homeostatic deregulation may play an important role in the progressive neurodegeneration. Proteins/peptides identified in extracellular deposits characteristic of AMD undergo pathogenic aggregation promoted by metals.^{36,37} The anomalous protein aggregation observed in AMD may be related to the existence of metal dyshomeostasis, as a cause or consequence.²⁶ Hence, the understanding of structural and functional interactions between metals and intracellular and extracellular components becoming pathogens may shed light to the etiology of AMD disorder. Therefore the identification of potential altered levels of metals, metalloids, and proteins interacting with metals in the tear film of AMD patients may contribute to the understanding of the pathophysiological changes observed in this irreversible disorder and, at the same time, propose candidate biomarkers for an early diagnosis or better prognosis of the disease.

We consequently designed a multitargeted analysis based on immunoassays and elemental mass spectrometry for the quantitative determination of LF, S100A6, CFH, CLU, APP, Ca, Mg, P, Na, Zn, Fe and Cu in the tear film of dry AMD patients and control subjects. Tears are produced by the lacrimal apparatus to moisten the ocular surface, protecting the cornea against desiccation, facilitating non-friction-bearing movements of the lids on the globe to improve the refractive effect and preserving against irritation caused by the environment.³⁸ The lacrimal glands are classic exocrine

acinar glands secreting a dilute aqueous solution containing proteins, small molecular mass components and electrolytes, constituting the tear film. Therefore, the target-biofluid tear film is mainly composed of H₂O (≈98%), with normal pH = 7.5 and Na, Cl, HCO₃, K, Mg, Ca, glucose, retinol and urea as major solutes. The most abundant tear proteins include immunoglobulin A, LF, tear-specific prealbumin, G protein, and lysozyme. Also, traces of other proteins, enzymes, and inhibitors from serum origin are present. The normal tear turnover occurs at approximately 16%/min at normal blink rate but is highly dependent on temperature and humidity and can be induced by a variety of mechanical and psychophysical stimuli.^{39,40} Accordingly, one of the main difficulties of this study deals with the limited sample volume, especially considering the age of the recruited patients. In the case of ELISA assays, tear sample dilution was carried out, permitting the multiparametric quantitation of selected molecules. Also, considering that the multielemental analysis by ICP-MS using conventional nebulization is dramatically limited by high sample volume consumption (up to 200 μL · min⁻¹), and even the use of micronebulizers typically permit uptakes of 50 to 100 μL · min⁻¹, we decided to optimize and implement the FIA-ICP-MS system, together allowing the quantitation of the 13 selected variables in this biofluid.

Normal composition of human tear has been widely characterized using proteomic,^{41–43} lipidomic,^{44–46} metabolomic,⁴⁷ and metallomic⁴⁸ approaches. Proteomic or metabolomic methodologies have been developed for the study of the lacrimal film and the identification of candidate biomarkers in patients diagnosed with glaucoma, dry eye, diabetic retinopathy, keratoconus, or cancer, among others.^{19,25,49} However, limited studies addressing the tear film analysis of AMD patients have been published to date. Among them, Yu et al.⁵⁰ observed, using ELISA assays, that the tear (along with serum) of AMD patients (n = 15) had increased IgA titers than control subjects (n = 15), which may imply a role of overactive IgA responses in AMD pathogenesis. Winiarczyk et al.⁵¹ attempted to define the AMD tear film proteome by MALDI-TOF/TOF using Schirmer strips for sampling in eight patients with wet AMD, six patients with dry AMD, and eight control individuals. From the identification of 342 proteins, shootin-1, histatin-3, fidgetin-like protein 1, SRC kinase signaling inhibitor, Grave's disease carrier protein, actin cytoplasmic 1, prolactin-inducible protein 1 and protein S100-A7A were found up-regulated in the tear film of dry AMD patients, involving specific pathways related to oxidative stress, inflammation, and proteolysis. Recently, the tear film of 15 patients with wet AMD and 15 age-matched healthy controls was comparatively analyzed by two-dimensional gel electrophoresis and MALDI-TOF/TOF, which identified altered proteins belong to pathways involving oxidative stress, protein clearance and chronic inflammation pathways, highlighting annexins A1 and A4 are part of the calcium-dependent phospholipid-binding family that regulate inflammation and autophagy.^{52,53} However, none of the identified altered proteins in those works coincide with those evaluated in our study showing significant variations.

The significant up-regulation of S100A6 protein observed in the tear film of our AMD group can be related to its role as Ca homeostasis regulator. The S100A6 belongs to the S100 family of Ca²⁺-binding proteins of the EF-hand type (i.e., with conserved calcium-binding motif).⁵⁴ In the eye, S100A6 is expressed in adult corneal endothelial cells but not in the

fetal corneal endothelial cells.⁵⁵ The protein levels of S100A6 were previously observed up-regulated in dry eye disease and Meibomian gland dysfunction^{52,56} and down-regulated in the tear film of patients diagnosed with keratoconus⁵⁷ and were therefore proposed as candidate biomarkers of these ocular surface disorders. Interestingly, S100A6 was found to be up-regulated in experimental corneal neovascularization models⁵⁸ and expressed in pterygium tissue removed from patient,^{59,60} but no studies related this protein with AMD disease to date.

For the first time, LF down-regulation was observed in AMD, when comparing patients with sex- and age-matched healthy subjects. This glycoprotein is normally presented in most mucosal secretions (i.e. nasal secretion, saliva, etc., in tears, vitreous humor, RPE, and retina).^{61–63} LF regulates iron homeostasis⁶⁴ and also presents anti-inflammatory, antibacterial, reactive oxygen species modulator, antiviral, and antitumor immunity effects.⁶⁵ This protein has been previously proposed as a candidate biomarker of dry eye disease,⁶⁶ in which oral LF administration preserves lacrimal gland function in aged mice, reducing the oxidative damage and inflammation.⁶⁷ The administration of LF also reduces the choroidal neovascularization in a laser-induced mouse model that mimics the AMD wet form through the inhibition of hypoxia-inducible factors.⁶⁸ Notably, LF concentration in the tear film of control individuals seems to remain invariant through age and independently of sex,^{62,69} as the current study shows. Moreover, the lower levels of LF observed in the tear film of AMD patients may be related to lower Fe concentration and increased oxidative stress observed during the course of the disease.

The MT1A subisoform belongs to the superfamily of metallothioneins, cysteine rich and metal-binding low molecular mass proteins (<7 kDa), involved in cellular Cu and Zn homeostasis, metal detoxification, defense against oxidative damage through free radical scavenging and neuroprotection.^{70–72} The MT1A, codified by the *MT1A* gene and expressed in all the human ocular tissues, was observed up-regulated in the tear of AMD patients, highlighting the high variability in this group in contrast to the control subjects (see Fig. 2F). Conversely, previous studies showed that levels of metallothioneins decreased with age in isolated retinal pigmented cells from postmortem donors with signs of AMD, but without determining individual subisoforms.⁷³ Remarkably, studies carried out using *in vivo* and *in vitro* models of retinal damage showed overexpression of MT1 and MT2 isoforms, suggesting that their up-regulation may be protective against acute retinal insults,^{74–76} which may explain the higher levels of MT1A observed in the AMD patients of this work.

From the different studies published so far, age-related changes of the tear composition have been observed, including increased expression of proinflammatory cytokines and matrix remodeling factors⁷⁷ and higher plasma-derived albumin levels.⁷⁸ Considering that aging is one of the main risk factors for AMD onset, we additionally studied possible correlation between targeted proteins and age in our cohort, without observing any age-related change in the tear composition, with the exception of APP protein. A negative correlation was obtained for APP, but only in the AMD group ($r = -0.523$, $P < 0.01$), indicating that the levels of this protein decreased with age in AMD patients but not in healthy controls. This observation may be associated with the age-related increase of the activity of APP cleaving enzymes, which contributes to APP degradation and

the increased production and accumulation of extracellular deposits,⁷⁹ being markedly declined in the AMD disease group. Consequently, the follow-up of this protein in the tear film of AMD patients during development and advanced stages of the disease warrants additional research.

Current literature does not include any study tackling the multielemental analysis of tear film of AMD patients. We solely identified limited publications attempting to quantify several elements in the tear film by different sampling and analysis approaches. A seminal work quantified the levels of Zn in tears of young healthier volunteers by atomic absorption spectrometry, obtaining a mean value of $1.537 \pm 0.146 \mu\text{g} \cdot \text{mL}^{-1}$.⁸⁰ A multielemental analysis of Na, K, Mg, and Ca was carried out in the tear film of 10 healthy humans by ICP-MS or ICP atomic emission spectroscopy, with levels of 104.5 ± 4.27 , 18.1 ± 1.04 , 0.49 ± 0.02 , and $0.33 \pm 0.03 \text{ mM}$, respectively.⁸¹ The multielemental analysis of tears from 60 volunteers from different geographic origins (urban vs rural areas), by ICP-MS, provided averaged concentrations of $18.40 \pm 16.53 \text{ ng} \cdot \text{mL}^{-1}$ for Cu and $29.61 \pm 31.46 \text{ ng} \cdot \text{mL}^{-1}$ for Zn.⁴⁸ Also, the possible relationship between tear Zn levels and helminths parasitic infection was evaluated in 81 individuals using a colorimetric assay, observing higher levels of this element in the infected population ($4.78 \mu\text{g} \cdot \text{mL}^{-1}$) when compared to control subjects ($2.42 \mu\text{g} \cdot \text{mL}^{-1}$).⁸² Finally, a multielemental analysis of Cu, Zn, Se, Rb, Ba, Pb, Mn, and Co was carried out by ICP-MS in 47 patients diagnosed with type II diabetes and 50 healthy controls, observing significant differences for Zn, Cr, Co, Mn, Ba, and Pb, with averaged concentration of Zn reaching $66.00 \text{ ng} \cdot \text{mL}^{-1}$ in diabetic patients and $33.25 \text{ ng} \cdot \text{mL}^{-1}$ in nondiabetic subjects.⁸³

The important variability observed for the above quantitative results of trace elements in tears must be stressed, which differs with our results for Cu and Zn (averaged of 0.16 and $0.36 \mu\text{g} \cdot \text{mL}^{-1}$, respectively). Experimental differences may be attributed to the tear sampling procedure or methodological determination. It seems that sampling using Schirmer strips provided lower levels of Zn and Cu,^{48,82,83} when compared to the use of glass microcapillary.^{80,81} Schirmer strips are routinely used in clinic to determine tear production, where the aqueous fraction of the tear travels further along the strips than non-polar proteins, lipids, or metabolites.⁴² This fact, along with the requirement of a quantitative extraction of the analytes from the strips for further analysis, may contribute to the observed differences in sample composition. In fact, the tear composition is affected by the method used for sample collection,⁸⁴ as discussed for proteins,⁸⁵ metabolites,²⁶ and lipids.⁴⁴ The use of microcapillary tubes is less invasive, safer, and avoids reflex tearing, leading to the identification of a huge number of extracellular proteins, when compared to the use of Schirmer strips,²⁵ but requires trained personal, which could explain the low number of studies using microcapillary sampling. Also, particular care must be taken with all the methodologies to avoid sample contamination during uptake and pretreatment.

The observed lower levels for Fe, Mg and Cu in the AMD cohort of our study may be related to disease onset or progression. The essential element Fe is the most abundant redox-active metal in the human body,⁸⁶ the local accumulation of which has been implicated in the pathogenesis of neurodegenerative diseases, including AMD.⁸⁷ Specifically, higher content of Fe has been detected in the photoreceptors, RPE and its melanosomes, Bruch's membrane, and

extracellular deposits of AMD patients,⁸⁷⁻⁹⁰ where its retinal accumulation may contribute to AMD pathogenesis by inducing oxidative stress and inflammation damage. Interestingly, elemental analysis of aqueous humor showed higher levels of Fe in AMD patients when compared to control subjects.⁹¹ Nonetheless, according to our results, decreased concentration of Fe in the recruited AMD patients seems to be directly related with the observed lower levels of LF, one of the main iron binding proteins in the tear film, which suggest the underlying interplay between both variables.

Likewise, Cu is an essential element for the visual cycle and photoreceptors survival, the deficiency of which has been associated with morphological changes in the retina and optic neuropathy.⁹² The lower Cu concentration determined in the tear film of our recruited patients diagnosed with AMD is in line with the lower levels of Cu observed in the aqueous humor of individuals with nonexudative AMD.⁹¹ With aging, increased systemic levels of Cu (blood or serum) was observed,⁹³ but the local Cu levels in the RPE and choroid were reduced,⁹⁴ specifically in AMD patients,⁹⁵ reflecting lower levels of this element within the affected eye. On the other hand, Mg is the second most abundant intracellular cation involved in maintenance of cell membrane function, synthesis of nucleic acids, and energy metabolism.⁹⁶ Heesterbeek et al.⁹⁷ determined by ICP-MS that Mg levels were higher in the plasma of AMD patients, but without statistical significance. Recently, Chang et al.⁹⁸ reported that multiple dietary nutrients, including Mg, were associated with decreased risk for neovascular AMD. Considering its physiological role in restoring the blood-brain barrier integrity, Mg deficiency has been associated with age-related neurodegenerative diseases, including amyotrophic lateral sclerosis (ALS) and Alzheimer's or Parkinson diseases.^{17,98} Specifically, Mg levels were lower in the blood of patients with ALS, when compared to control subjects, similarly to the significantly lower concentrations of this element determined in the tear film of AMD patients.

The higher prevalence of dyslipidemia in the AMD group is in agreement with previous studies, showing that pathogenesis may be related to circulating lipids, local lipid transport, or both.^{28,99} To discard possible confounders of demographic and clinical conditions, including the risk factor dyslipidemia, in the altered levels of target metals and (metallo-)proteins obtained in the tear film of AMD patients, when compared to control subjects, an additional statistical analysis was carried out. Exclusion of the subjects with dyslipidemia from the multivariate analysis provided similar results, as compared to statistical analysis obtained including individuals with that risk factor, with the exception of Mg, probably because of the individual's reduction in the studied cohort, indicating the low influence of this risk factor on the obtained differences. Interestingly, the multivariate logistic regression analysis confirmed that dyslipidemia did not contribute to the observed differences, and therefore the differences observed for MT1A, S100A6, LF, Fe and Cu are as result of AMD disease.

Additionally, the possible correlations between proteins and elements were also investigated, observing a statistically significant positive trend for Mg when comparing with Na, Cu and P (Figs. 4A-C), for both AMD and control subjects. Conversely, positive correlations for Mg and Ca (Fig. 4D), for Ca and Na (Fig. 4E), and for LF and Cu, were exclusively observed for the AMD group and not for the healthy controls. Notably, LF was the only protein showing high positive

correlation with Cu (Fig. 4 D), but not for Fe, suggesting a possible interdependence in AMD disease. The highly positive correlation observed for Mg with Na, P and Ca may suggest an agonist behavior of Mg,¹⁰⁰ and also for Na and P and for LF and Cu. However, the specific positive interaction in the AMD group needs to be elucidated. Interestingly, the quantitative data obtained for the thirteen variables in both AMD and control groups were used to establish a three-protein-metal panel, consisting of MT1A, Na and Mg. This panel was used to generate machine-learning models (see Table 3), obtaining the classification accuracy when comparing AMD patients versus control subjects, which predicts the disease in the 73% of cases. However, the diagnostic power for predicting AMD is quite low using these variables, and it seems that their alterations in tears are not directly associated between themselves.

Even though this is a comprehensive targeted study on trace metals and (metallo-)proteins in patients with dry AMD, this work should be considered as exploratory. This study is limited by the restricted number of samples used for statistical analysis, which would require additional recruited patients to obtain more than 300 samples for each group to reach adequate power of the tests. Hence, the low number of samples used for classifying groups may affect the discriminating power of the studied panel of MT1A, Na and Mg, using the proposed algorithms. Also, evaluation of tear film composition throughout the day and in the long time would be desirable in future studies to determine intra- and/or inter-daily variations. Finally, issues regarding the relevant correlation of the tear film composition with macular lesions are open, considering that although the tear film is not directly connected with the retina, it can be altered by the partial blood-retinal barrier breakdown in the course of AMD.

The multitargeted approach developed for the analysis of limited volume from the tear film permitted us to obtain quantitative information for 13 variables, including LF, S100A6, CFH, CLU, APP, MT1A, Ca, Mg, P, Na, Cu, Fe, and Zn, combining spectrophotometry and mass spectrometry analytical techniques. The specific altered levels of metals and (metallo-)proteins (i.e., MT1A, S100A6, LF, Fe, Mg and Cu) in the tear film of AMD patients suggest compromised metal homeostasis during the progression of this neurodegenerative disease and may contribute to shedding light into the pathophysiology of macular degeneration. The possible relation of observed pathophysiological changes with the anomalous protein aggregation deserves further research.

Acknowledgments

The authors thank Esther Vázquez for her technical support during tear sampling collection.

Supported through project PID2019-107838RB-I00/Agencia Estatal de Investigación (AEI)/10.13039/501100011033). E. Valencia acknowledges the FPI Grant (Ref. BES-2017-080518) from the Ministry of Economy and Competitiveness. The Instituto Oftalmológico Fernández-Vega and Fundación de Investigación Oftalmológica acknowledge financial support from the Fundación Rafael del Pino (<http://www.frdelpino.es>), through the “Cátedra Rafael del Pino.”

Disclosure: **E. Valencia**, none; **M. García**, none; **B. Fernández-Vega**, none; **R. Pereiro**, none; **L. Lobo**, none; **H. González-Iglesias**, none

References

- Desa UN. *World population prospects 2019: highlights*. New York: United Nations Department for Economic and Social Affairs. 2019;11(1):125.
- Jucker M, Walker LC. Propagation and spread of pathogenic protein assemblies in neurodegenerative diseases. *Nat Neurosci*. 2018;21:1341–1349.
- Wong WL, Su X, Li X, et al. Global prevalence of age-related macular degeneration and disease burden projection for 2020 and 2040: a systematic review and meta-analysis. *Lancet Glob Heal*. 2014;2(2):e106–e116.
- Age-Related Eye Disease Study Research Group. The age-related eye disease study system for classifying age-related macular degeneration from stereoscopic color fundus photographs: The age-related eye disease study report number 6. *Am J Ophthalmol*. 2001;132:668–681.
- Sadda SVR, Guymer R, Monés JM, Tufail A, Jaffe GJ. Anti-vascular endothelial growth factor use and atrophy in neovascular age-related macular degeneration: systematic literature review and expert opinion. *Ophthalmology*. 2020;127:648–659.
- Chakravarthy U, Bailey CC, Johnston RL, et al. Characterizing disease burden and progression of geographic atrophy secondary to age-related macular degeneration. *Ophthalmology*. 2018;125:842–849.
- Crabb JW, Miyagi M, Gu X, et al. Drusen proteome analysis: an approach to the etiology of age-related macular degeneration. *Proc Natl Acad Sci USA*. 2002;99:14682–14687.
- Lengyel I, Flinn JM, Peto T, et al. High concentration of zinc in sub-retinal pigment epithelial deposits. *Exp Eye Res*. 2007;84:772–780.
- Flinn JM, Kakalec P, Tappero R, Jones B, Lengyel I. Correlations in distribution and concentration of calcium, copper and iron with zinc in isolated extracellular deposits associated with age-related macular degeneration. *Metallomics*. 2014;6:1223–1228.
- Thompson RB, Reffatto V, Bundy JG, et al. Identification of hydroxyapatite spherules provides new insight into subretinal pigment epithelial deposit formation in the aging eye. *Proc Natl Acad Sci USA*. 2015;112:1565–1570.
- Wang L, Clark ME, Crossman DK, et al. Abundant lipid and protein components of drusen. *PLoS One*. 2010;5(4):e10329.
- Astrid Sigel, Sigel H, Roland KO, ed. *Neurodegenerative diseases and metal ions*. Vol 1. Hoboken, NJ: John Wiley & Sons; 2014.
- Quintanar L, Lim MH. Metal ions and degenerative diseases. *J Biol Inorg Chem*. 2019;24:1137–1139.
- Tan ACS, Pilgrim MG, Fearn S, et al. Calcified nodules in retinal drusen are associated with disease progression in age-related macular degeneration. *Sci Transl Med*. 2018;10(466):1–12.
- Everett J, Lermyte F, Brooks J, et al. Biogenic metallic elements in the human brain? *Sci Adv*. 2021;7(24):eabf6707.
- Gilbert R, Peto T, Lengyel I, Emri E. Zinc nutrition and inflammation in the aging retina. *Mol Nutr Food Res*. 2019;63(15):1–16.
- Cicero CE, Mostile G, Vasta R, et al. Metals and neurodegenerative diseases. A systematic review. *Environ Res*. 2017;159(July):82–94.
- White AR, Kanninen KM, Crouch PJ. Editorial: Metals and neurodegeneration: restoring the balance. *Front Aging Neurosci*. 2015;7(JUL):1–2.
- von Thun und Hohenstein-Blaul N, Funke S, Grus FH. Tears as a source of biomarkers for ocular and systemic diseases. *Exp Eye Res*. 2013;117:126–137.

20. Sullivan DA, Stern ME, Tsubota K, Dartt DA, Sullivan RM, Bromberg B, ed. *Lacrimal Gland, Tear Film, and Dry Eye Syndromes 3*. 3rd ed. Boston: Springer; 2002.
21. King-Smith PE, Fink BA, Hill RM, Koelling KW, Tiffany JM. The thickness of the tear film. *Curr Eye Res*. 2004;29(4-5):357–368.
22. Cwiklik L. Tear film lipid layer: a molecular level view. *Biochim Biophys Acta - Biomembr*. 2016;1858:2421–2430.
23. Hagan S, Martin E, Enríquez-de-Salamanca A. Tear fluid biomarkers in ocular and systemic disease: potential use for predictive, preventive and personalised medicine. *EPMA J*. 2016;7:1–20.
24. Johnson ME, Murphy PJ. Changes in the tear film and ocular surface from dry eye syndrome. *Prog Retin Eye Res*. 2004;23:449–474.
25. Ponzini E, Santambrogio C, De Palma A, Mauri P, Tavazzi S, Grandori R. Mass spectrometry-based tear proteomics for noninvasive biomarker discovery [published online ahead of print March 24, 2021]. *Mass Spectrom Rev*, doi:10.1002/mas.21691.
26. Brown CN, Green BD, Thompson RB, Den Hollander AI, Lengyel I. Metabolomics and age-related macular degeneration. *Metabolites*. 2019;9(1):4.
27. Seddon JM, Sharma S, Adelman RA. Evaluation of the clinical age-related maculopathy staging system. *Ophthalmology*. 2006;113:260–266.
28. Curk T, Demsar J, Xu Q, et al. Microarray data mining with visual programming. *Bioinformatics*. 2005;21:396–398.
29. Fan Q, Maranville JC, Fritsche L, et al. HDL-cholesterol levels and risk of age-related macular degeneration: a multiethnic genetic study using Mendelian randomization. *Int J Epidemiol*. 2017;46:1891–1902.
30. Fernández-Vega B, García M, Olivares L, et al. The association study of lipid metabolism gene polymorphisms with AMD identifies a protective role for APOE-E2 allele in the wet form in a Northern Spanish population. *Acta Ophthalmol*. 2020;98(3):e282–e291.
31. Ponzini E, Scotti L, Grandori R, Tavazzi S, Zambon A. Lactoferrin concentration in human tears and ocular diseases: a meta-analysis. *Invest Ophthalmol Vis Sci*. 2020;61(12):9.
32. Benitez-Del-Castillo JM, Soria J, Acera A, Muñoz AM, Rodríguez S, Suárez T. Quantification of a panel for dry-eye protein biomarkers in tears: a comparative pilot study using standard ELISA and customized microarrays. *Mol Vis*. 2021;27(May):243–261.
33. Soria J, Acera A, Merayo-Llodes J, et al. Tear proteome analysis in ocular surface diseases using label-free LC-MS/MS and multiplexed-microarray biomarker validation. *Sci Rep*. 2017;7(1):1–15.
34. Yu V, Bhattacharya D, Webster A, et al. Clusterin from human clinical tear samples: Positive correlation between tear concentration and Schirmer strip test results. *Ocul Surf*. 2018;16:478–486.
35. Gachon AM, Richard J, Dastugue B. Human tears: normal protein pattern and individual protein determinations in adults. *Curr Eye Res*. 2009;2(5):301–308.
36. Tamás MJ, Sharma SK, Ibstedt S, Jacobson T, Christen P. Heavy metals and metalloids as a cause for protein misfolding and aggregation. *Biomolecules*. 2014;4:252–267.
37. Lee JC, Kim JE, Pletneva E V, Faraone-Mennella J, Gray HB, Winkler JR. Protein Folding, Misfolding, and Disease. In: *Neurodegenerative Diseases and Metal Ions*. Hoboken, NJ: John Wiley & Sons, Ltd; 2006:9–60.
38. Pflugfelder SC, Stern ME. Biological functions of tear film. *Exp Eye Res*. 2020;197:108115.
39. Forrester J, Dick A, McMenamin P, Roberts F, Pearlman E. *The Eye. Basic Science in Practice*. 4th ed. St. Louis: Elsevier Health Sciences; 2015.
40. Dartt DA, Willcox MDP. Complexity of the tear film: Importance in homeostasis and dysfunction during disease. *Exp Eye Res*. 2013;117:1–3.
41. Zhou L, Zhao SZ, Koh SK, et al. In-depth analysis of the human tear proteome. *J Proteomics*. 2012;75:3877–3885.
42. Grus FH, Joachim SC, Pfeiffer N. Proteomics in ocular fluids. *Proteomics - Clin Appl*. 2007;1:876–888.
43. Karnati R, Laurie DE, Laurie GW. Lacritin and the tear proteome as natural replacement therapy for dry eye. *Exp Eye Res*. 2013;117:39–52.
44. Lam SM, Tong L, Duan X, Petznick A, Wenk MR, Shui G. Extensive characterization of human tear fluid collected using different techniques unravels the presence of novel lipid amphiphiles. *J Lipid Res*. 2014;55:289–298.
45. Rantamäki AH, Seppänen-Laakso T, Oresic M, Jauhainen M, Holopainen JM. Human tear fluid lipidome: From composition to function. *PLoS One*. 2011;6(5):1–7.
46. Butovich IA. Tear film lipids. *Exp Eye Res*. 2013;117:4–27.
47. Chen L, Zhou L, Chan ECY, Neo J, Beuerman RW. Characterization of the human tear metabolome by LC-MS/MS. *J Proteome Res*. 2011;10:4876–4882.
48. Semeraro F, Costagliola C, Cancarini A, Gilberti E, Tosco E, Apostoli P. Defining reference values of trace elements in the tear film: Diagnostic methods and possible applications. *Ecotoxicol Environ Saf*. 2012;80:190–194.
49. Roda M, Ciavarella C, Giannaccare G, Versura P. Biomarkers in tears and ocular surface: a window for neurodegenerative diseases. *Eye Contact Lens*. 2020;46:S129–S134.
50. Yu H, Yuan L, Yang Y, et al. Increased serum IgA concentration and plasmablast frequency in patients with age-related macular degeneration. *Immunobiology*. 2016;221:650–656.
51. Winiarczyk M, Kaarniranta K, Winiarczyk S, Adaszek Ł, Winiarczyk D, Mackiewicz J. Tear film proteome in age-related macular degeneration. *Graefes Arch Clin Exp Ophthalmol*. 2018;256:1127–1139.
52. Winiarczyk M, Winiarczyk D, Michalak K, et al. Dysregulated tear film proteins in macular edema due to the neovascular age-related macular degeneration are involved in the regulation of protein clearance, inflammation, and neovascularization. *J Clin Med*. 2021;10:3060.
53. Xi Y, Ju R, Wang Y. Roles of annexin A protein family in autophagy regulation and therapy. *Biomed Pharmacother*. 2020;130(August):110591.
54. Donato R, Cannon BR, Sorci G, et al. Functions of S100 Proteins. *Curr Mol Med*. 2012;13:24–57.
55. Chen Y, Huang K, Nakatsu MN, Xue Z, Deng SX, Fan G. Identification of novel molecular markers through transcriptomic analysis in human fetal and adult corneal endothelial cells. *Hum Mol Genet*. 2013;22:1271–1279.
56. Soria J, Durán JA, Etxebarria J, et al. Tear proteome and protein network analyses reveal a novel pentamer panel for tear film characterization in dry eye and meibomian gland dysfunction. *J Proteomics*. 2013;78:94–112.
57. Recalde JI, Duran JA, Rodríguez-Agirretxe I, et al. Changes in tear biomarker levels in keratoconus after corneal collagen crosslinking. *Mol Vis*. 2019;25:12–21.
58. Li C, Zhang F, Wang Y. S100A proteins in the pathogenesis of experimental corneal neovascularization. *Mol Vis*. 2010;16:2225–2235.
59. Riau AK, Wong TT, Beuerman RW, Tong L. Calcium-binding S100 protein expression in pterygium. *Mol Vis*. 2009;15:335–342.
60. Jaworski CJ, Aryankalayil-John M, Campos MM, et al. Expression analysis of human pterygium shows a predominance of conjunctival and limbal markers and genes associated with cell migration. *Mol Vis*. 2009;15:2421–2434.
61. Baker EN, Baker HM. Molecular structure, binding properties and dynamics of lactoferrin. *Cell Mol Life Sci*. 2005;62:2531–2539.

62. Flanagan JL, Willcox MDP. Role of lactoferrin in the tear film. *Biochimie*. 2009;91(1):35–43.
63. Clausen R, Weller M, Wiedemann P, Heimann K, Hilgers RD, Zilles K. An immunochemical quantitative analysis of the protein pattern in physiological and pathologic vitreous. *Graefes Arch Clin Exp Ophthalmol*. 1991;229:186–190.
64. Lawen A, Lane DJR. Mammalian iron homeostasis in health and disease: Uptake, storage, transport, and molecular mechanisms of action. *Antioxidants Redox Signal*. 2013;18:2473–2507.
65. Li Y-Q, Guo C. A review on lactoferrin and central nervous system diseases. *Cells*. 2021;10:1810.
66. Grus FH, Podust VN, Bruns K, et al. SELDI-TOF-MS ProteinChip array profiling of tears from patients with dry eye. *Invest Ophthalmol Vis Sci*. 2005;46:863–876.
67. Kawashima M, Kawakita T, Inaba T, et al. Dietary lactoferrin alleviates age-related lacrimal gland dysfunction in mice. *PLoS One*. 2012;7(3):3–10.
68. Ibuki M, Shoda C, Miwa Y, Ishida A, Tsubota K, Kurihara T. Lactoferrin has a therapeutic effect via HIF inhibition in a murine model of choroidal neovascularization. *Front Pharmacol*. 2020;11(February):1–11.
69. Kijlstra A, Jeurissen SHM, Koning KM. Lactoferrin levels in normal human tears. *Br J Ophthalmol*. 1983;67:199–202.
70. Gonzalez-Iglesias H, Alvarez L, García M, Petrash C, Sanz-Medel A, Coca-Prados M. Metallothioneins (MTs) in the human eye: a perspective article on the zinc-MT redox cycle. *Metallomics*. 2014;6:201–208.
71. Álvarez-Barrios A, Álvarez L, García M, Artime E, Pereiro R, González-Iglesias H. Antioxidant defenses in the human eye: A focus on metallothioneins. *Antioxidants*. 2021;10:1–33.
72. Vasák M, Hasler DW. Metallothioneins: new functional and structural insights. *Curr Opin Chem Biol*. 2000;4:177–183.
73. Tate DJ, Newsome DA, Oliver PD. Metallothionein shows an age-related decrease in human macular retinal pigment epithelium. *Invest Ophthalmol Vis Sci*. 1993;34:2348–2351.
74. Rodríguez-Menéndez S, García M, Fernández B, et al. The zinc-metallothionein redox system reduces oxidative stress in retinal pigment epithelial cells. *Nutrients*. 2018;10:1874.
75. Chen L, Wu W, Dentchev T, et al. Light damage induced changes in mouse retinal gene expression. *Exp Eye Res*. 2004;79:239–247.
76. Biswal MR, Ahmed CM, Ildefonso CJ, et al. Systemic treatment with a 5HT1a agonist induces anti-oxidant protection and preserves the retina from mitochondrial oxidative stress. *Exp Eye Res*. 2015;140:94–105.
77. Micera A, Di Zazzo A, Esposito G, et al. Age-related changes to human tear composition. *Invest Ophthalmol Vis Sci*. 2018;59:2024–2031.
78. Kondkar AA, Azad TA, Almobarak FA, Kalantan H, Al-Obeidan SA, Abu-Amro KK. Elevated levels of plasma tumor necrosis factor alpha in patients with pseudoexfoliation glaucoma. *Clin Ophthalmol*. 2018;12:153–159.
79. Fukumoto H, Rosene DL, Moss MB, Raju S, Hyman BT, Irizarry MC. β -Secretase activity increases with aging in human, monkey, and mouse brain. *Am J Pathol*. 2004;164:719–725.
80. Saatçi AO, Irkeç M, Özgüneş H. Zinc in tears. *Ophthalmic Res*. 1991;23:31–32.
81. Wei XE, Markoulli M, Millar TJ, Willcox MDP, Zhao Z. Divalent cations in tears, and their influence on tear film stability in humans and rabbits. *Invest Ophthalmol Vis Sci*. 2012;53:3280–3285.
82. Framroze Z, Conroy C. Measuring the change in zinc ion concentration in eye tear fluid between healthy and parasite infected individuals: relationship between zinc ions in tear fluid and parasitic infection by soil-transmitted helminths. *J Med Diagnostic Methods*. 2016;5(4):8–11.
83. Cancarini A, Fostinelli J, Napoli L, Gilberti ME, Apostoli P, Semeraro F. Trace elements and diabetes: assessment of levels in tears and serum. *Exp Eye Res*. 2017;154:47–52.
84. Rentka A, Koroskenyi K, Harsfalvi J, et al. Evaluation of commonly used tear sampling methods and their relevance in subsequent biochemical analysis. *Ann Clin Biochem*. 2017;54:521–529.
85. Green-Church KB, Nichols KK, Kleinholz NM, Zhang L, Nichols JJ. Investigation of the human tear film proteome using multiple proteomic approaches. *Mol Vis*. 2008;14:456–470.
86. Hirayama T. Fluorescent probes for the detection of catalytic Fe(II) ion. *Free Radic Biol Med*. 2019;133(June 2018):38–45.
87. Biesemeier A, Yoeruek E, Eibl O, Schraermeyer U. Iron accumulation in Bruch's membrane and melanosomes of donor eyes with age-related macular degeneration. *Exp Eye Res*. 2015;137:39–49.
88. Wong RW, Richa DC, Hahn P, Green WR, Dunaief JL. Iron toxicity as a potential factor in AMD. *Retina*. 2007;27:997–1003.
89. Dunaief JL. Iron induced oxidative damage as a potential factor in age-related macular degeneration: the Cogan lecture. *Invest Ophthalmol Vis Sci*. 2006;47:4660–4664.
90. Hahn P, Milam AH, Dunaief JL. Maculas affected by age-related macular degeneration contain increased chelatable iron in the retinal pigment epithelium and Bruch's membrane. *Arch Ophthalmol*. 2003;121:1099–1105.
91. Jünemann AGM, Stopa P, Michalke B, et al. Levels of aqueous humor trace elements in patients with non-exudative age-related macular degeneration: a case-control study. *PLoS One*. 2013;8(2):1–7.
92. Pineles SL, Wilson CA, Balcer IJ, Slater R, Galetta SL. Combined optic neuropathy and myelopathy secondary to copper deficiency. *Surv Ophthalmol*. 2010;55:386–392.
93. Belbraouet S, Biaudet H, Tébi A, Chau N, Gray-Donald K, Debry G. Serum zinc and copper status in hospitalized vs. healthy elderly subjects. *J Am Coll Nutr*. 2007;26:650–654.
94. Wills NK, Ramanujam VMS, Kalariya N, Lewis JR, van Kuijk FJGM. Copper and zinc distribution in the human retina: relationship to cadmium accumulation, age, and gender. *Exp Eye Res*. 2008;87:80–88.
95. Erie JC, Good JA, Butz JA, Pulido JS. Reduced zinc and copper in the retinal pigment epithelium and choroid in age-related macular degeneration. *Am J Ophthalmol*. 2009;147:276–282.e1.
96. Saris NE, Mervaala E, Karppanen H, Khawaja JA, Magnesium. A. An update on physiological, clinical and analytical aspects. *Clin Chim Acta*. 2000;294(1-2):1–26.
97. Heesterbeek TJ, Rouhi-Parkouhi M, Church SJ, et al. Association of plasma trace element levels with neovascular age-related macular degeneration. *Exp Eye Res*. 2020;201:108324.
98. Chang JJ, Mack WJ, Saver JL, Sanossian N. Magnesium: Potential roles in neurovascular disease. *Front Neurol*. 2014;5:1–8.
99. van Leeuwen EM, Emri E, Merle BMJ, et al. A new perspective on lipid research in age-related macular degeneration. *Prog Retin Eye Res*. 2018;67:56–86.
100. Grigoryan R, Costas-Rodríguez M, Vandenbroucke RE, Vanhaecke F. High-precision isotopic analysis of Mg and Ca in biological samples using multi-collector ICP-mass spectrometry after their sequential chromatographic isolation - Application to the characterization of the body distribution of Mg and Ca isotopes in mice. *Anal Chim Acta*. 2020;1130:137–145.